Abstract. The leukocyte function-associated molecule 1 (LFA-1, CD11a/CD18) is a membrane glycoprotein which functions in cell-cell adhesion by heterophilic interaction with intercellular adhesion molecule 1 (ICAM-1). LFA-1 consists of an α subunit (Mr = 180,000) and a β subunit (Mr = 95,000). We report the molecular biology and protein sequence of the α subunit. Overlapping cDNAs containing 5,139 nucleotides were isolated using an oligonucleotide specified by tryptic peptide sequence. The mRNA of 5.5 kb is expressed in lymphoid and myeloid cells but not in a bladder carcinoma cell line. The protein has a 1,063-amino acid extracellular domain, a 29-amino acid transmembrane region, and a 53-amino acid cytoplasmic tail. The extracellular domain contains seven repeats. Repeats V-VII are in tandem and contain putative divalent cation binding sites. LFA-1 has significant homology to the members of the integrin superfamily, having 36% identity with the Mac-1 and p150,95 α subunits and 28% identity with other integrin α subunits. An insertion of ~200 amino acids is present in the NH2-terminal region of LFA-1. This “inserted/interactive” or I domain is also present in the p150,95 and Mac-1 α subunits but is absent from other integrin α subunits sequenced to date. The I domain has striking homology to three repeats in human von Willebrand factor, two repeats in chicken cartilage matrix protein, and a region of complement factor B. These structural features indicate a bipartite evolution from the integrin family and from an I domain family. These features may also correspond to relevant functional domains.

The leukocyte function-associated molecule 1 (LFA-1) is a member of a family of three leukocyte glycoproteins involved in cell-cell adhesion. This family of proteins, LFA-1, Mac-1, and p150,95, are heterodimers consisting of distinct α subunits (Mr = 180,000, 170,000, and 150,000, respectively) and a common β subunit (Mr = 95,000) (25, 47). NH2-terminal sequencing of the α subunits in the mouse and human has suggested that they are structurally related (33, 48); however, the cell surface expression and function of these molecules differs. LFA-1 is expressed on virtually all leukocytes and is involved in a large number of adhesion-dependent phenomena. mAb directed against LFA-1 inhibit antigen-specific T helper cell function and cytolytic functions such as cytotoxic T lymphocyte-mediated killing, antibody-dependent cytotoxicity by granulocytes, and natural killer activity (47). LFA-1 is also involved in antigen-independent interactions that mediate cell localization to sites of inflammation such as leukocyte adhesion to endothelial cells, fibroblasts, epidermal keratinocytes, and synovial cells (12-14, 17, 30, 53). Mac-1 and p150,95 are expressed on monocytes, granulocytes, and some activated lymphocytes, and function as adhesion molecules in cell-cell and cell-substrate interactions as well as complement receptors for C3bi (1).

The importance of these three glycoproteins is signified by the clinical syndrome known as leukocyte adhesion deficiency (LAD) (1). The primary defect in LAD occurs in the β subunit common to LFA-1, Mac-1, and p150,95 (22) resulting in deficient surface expression of the αβ complexes. LAD patients have recurrent bacterial infections which are sometimes fatal and their leukocytes are deficient in a wide range of adhesion-dependent functions.

The structure of the β subunit common to LFA-1, Mac-1, and p150,95 has been determined (23, 27) and has revealed homology to extracellular matrix (ECM) receptors. These similarities led to the concept of a family of αβ heterodimers designated the integrins (21, 41). The term "integrin" emphasizes the role of these proteins as transmembrane links between the extracellular environment and the cytoskeleton.
Three subfamilies of integrins are defined by their distinct β subunits. The β1 subunit is common to the fibronectin receptor (FN) and some antigens appearing very late in leukocyte activation whereas β2 is common to platelet glycoprotein IIb/IIIa (gpIIb/IIIa) and the vitronectin receptor. β1 and β2 integrins are all ECM receptors and are involved in cell–substrate adhesion, matrix assembly, regulation of cell growth, differentiation, and localization during morphogenesis, and wound healing. We will refer to the β1 and β2 family as the extracellular matrix receptor integrins. The β2 subunit is common to LFA-1, Mac-1, and p150,95, whose expression is limited to leukocytes, and we designate these as the leukocyte integrins.

The ligand of LFA-1 is an inducible cell surface glycoprotein, intercellular adhesion molecule-1 (ICAM-1) (Mr = 90,000), which is found on cells of many lineages including leukocytes, endothelial cells, fibroblasts, and epithelial keratinocytes (14, 29, 40, 46, 49). ICAM-1 mRNA and surface expression is induced by inflammatory mediators including interferon-gamma, interleukin-1, tumor necrosis factor, and lipopolysaccharide (36, 46, 49); thus, it may regulate cell interaction and localization in inflammation. LFA-1-dependent adhesion of cells to planar lipid membranes containing ICAM-1 requires metabolic energy, a functional cytoskeleton, and divalent cations (29). Cell activation can enhance LFA-1-dependent adhesion without any effect on LFA-1 or ICAM-1 surface expression as shown by phorbol ester–induced homotypic adhesion of B, T, and monocytic cells (39).

ICAM-1 is a member of the immunoglobulin superfamily and consists of five immunoglobulin constant region-like domains. In contrast to most ligands of the ECM receptor integrins which contain the critical recognition sequence arginine-glycine-aspartic acid (RGD), ICAM-1 does not contain an RGD sequence (46, 49). The LFA-1–ICAM-1 receptor–ligand pair is thus far the only known example of a member of the integrin superfamily interacting with a member of the immunoglobulin superfamily.

We have been interested in the structural basis for the important function of LFA-1 in inflammation and the immune response. Furthermore, we wished to define the relationship of LFA-1 to other leukocyte integrins and to the ECM receptor integrins. The lack of an RGD sequence in ICAM-1 raised the question of whether the LFA-1 β subunit has structural homology with other members of the integrin superfamily. A region near the NH2 terminus of the molecule contains an insertion of ~200 amino acids similar to p150,95 (9) and Mac-1 (4, 7, 38). This domain has significant homology to the type A domains of von Willebrand factor (vWF), complement factor B, and the repeats of chicken cartilage matrix protein (CMP). These similarities suggest relevant functional domains within the LFA-1 α subunit as well as novel evolutionary relationships.

Materials and Methods

Protein Purification

The mAb, TSI/22, which is directed against the LFA-1 α subunit, was purified and coupled using cyanogen bromide to CL-4B Sepharose at 2 mg mAb per ml of packed bed. SKW3 cells (42.2 g) were lysed in 300 ml of lysis buffer (25) and the lysate was spun at 5000 g, the pellet discarded, and then spun at 16,000 g for 2 h. The supernatant was then sequentially passed through a precolumn of activated and quenched CL-4B Sepharose and then a TSI/22 mAb Sepharose column. The TSI/22 column was washed sequentially (25), and the LFA-1 molecule was eluted with 0.5 M NaCl, 0.1% Triton X-100, 1 m M iodoacetamide, 10 U/ml aprotinin, and 0.025% NaN3, 50 mM triethylamine, pH 11.5, and the pH immediately neutralized. The fractions containing LFA-1 were pooled, lyophilized, and precipitated in 5 vol ethanol at ~20°C overnight. Purified protein was reduced and alkylated (23) and subjected to preparative SDs-PAGE. The band corresponding to the α subunit was visualized with 1 M KCl, excised, and electrophoresed (20). The purified α subunit was lyophilized and precipitated in 4 vol ethanol at ~20°C overnight. The pellet was resuspended and digested with 1% (wt/wt) trypsin (23). The trypic fragments were then isolated by HPLC (Beckman Instruments Inc., Palo Alto, CA) on a C4 reverse phase column (Vydac, Hesperia, CA). The peptides were eluted on a 0–60% acetonitrile gradient in 1% trifluoroacetic acid. Several peaks were rechromatographed isocratically in a concentration of acetonitrile determined by the equation, F = 0.9E-2, where F is the volume percentage of acetonitrile under isocratic conditions for a peptide that eluted at E percent during the linear gradient (57). Peaks were collected in 1.5-ml polypropylene tubes and concentrated to <50 μl. Eight peaks were subjected to microsequencing. The sequence of one peptide, LE4, was used to synthesize a single sequence oligonucleotide (5′-GGGATGTTTGGGTTACAAT-3) according to suggestions of Lathe (26). cDNA Cloning, Restriction Mapping, and Nucleotide Sequencing

The production and screening of the cDNA library was performed as previously described (9). Restriction maps of the selected clones were determined by double and partial digests (28). Restriction fragments were subcloned into either M13mp8 and mpl9 (31) or pGEM-3Z, 4Z, or 7Z (Promega Biotec, Madison, WI). Deletions of the fragments in pGEM were made using Exonuclease III and S1 nuclease (18). Sequencing was by the dideoxy termination method (43). The coding, 5′-untranslated and 3′-untranslated regions were determined 100, 100, and 36.7% in both orientations, respectively.

Southern and Northern Blot

Southern blots were performed as described elsewhere (8). For Northern blots, 10 μg of poly (A+) RNA isolated from SKW3, U937, IB4, or RJ cells was subjected to electrophoresis on a 1.0% formaldehyde gel and transferred to nitrocellulose (5). The nitrocellulose (Bio-Rad Laboratories, Richmond, CA) was prehybridized and hybridized in 2× SSC, 1× Denhardt’s solution, 0.1% SDS, and 10 μg/ml of herring sperm DNA. A 18-kb Eco RI probe from the 5′ end of the cDNA clone, λ3R1 (Fig. 2), was labeled by nick translation and used as probe.

Computer Analysis

Homology searches and alignment of sequences used the Microgene DNA program (Beckman Instruments Inc.), FASTP (56) on the NBRF and NEW databases (National Biomedical Research Foundation, Washington, DC), and FASTP using the SWISS-PROT data base (Bionet Intelligentics, Mountainview, CA). These alignments were then optimized, and the percent identity and statistical significance were determined using ALIGN (NBRF) (11).

Hydropathy was determined according to Hopp and Woods using the Microgene DNA program (19).

Results and Discussion

Protein Purification and Peptide Sequence

LFA-1 was solubilized from SKW3, a T lymphoma cell line,
Table 1. Sequences of Tryptic Peptides

<table>
<thead>
<tr>
<th>Residues</th>
<th>Amino acid sequence</th>
</tr>
</thead>
<tbody>
<tr>
<td>95–107</td>
<td>XDQ (N) YLSGI (E) YLF</td>
</tr>
<tr>
<td>199–210</td>
<td>HMLLTLTFGA1</td>
</tr>
<tr>
<td>254–260</td>
<td>YIGIGK</td>
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<tr>
<td>405–413</td>
<td>VLLFQEQQ</td>
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<tr>
<td>494–503</td>
<td>GEAITALTXI</td>
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<tr>
<td>541–554</td>
<td>IEGTQVLSGIQXFQ</td>
</tr>
<tr>
<td>564–576</td>
<td>X (L) (E) D (V/G) LADVAVGAE</td>
</tr>
<tr>
<td>803–817</td>
<td>KVEMLKPHSEIXVS</td>
</tr>
<tr>
<td>929–946</td>
<td>(E/Q) PSHDHNPXLEAVXG</td>
</tr>
</tbody>
</table>

Parentheses indicate ambiguity in the sequence. The underlined residues were used to generate an oligonucleotide probe.

Figure 1. Protein purification of LFA-1 α subunit. Silver-stained SDS-PAGE gel showing a column fraction of the affinity-purified protein (lane 1) and the electroluted α subunit (lane 2).

and isolated by mAb affinity chromatography using an antibody directed against the α subunit. SDS-PAGE showed the α and β subunits (Fig. 1, lane 1). The α subunit was further purified by preparative SDS-PAGE (Fig. 1, lane 2) and the purified α subunit was digested with trypsin and the peptides were isolated by reverse phase HPLC. The sequence of nine peptides was determined by microsequencing (Table I) and the sequence possessing the lowest codon redundancy was used to specify a single oligonucleotide sequence which used the most commonly occurring human codons (26).

cDNA Cloning and Characterization

The 32-mer oligonucleotide probe was used to isolate 20 clones from a size-selected agt10 cDNA library constructed from PMA-stimulated myeloid cells (9). These cells have been previously shown to synthesize the LFA-1 α subunit (32). The insert size was determined and the longest clone, λ5L5, was restriction mapped, and sequenced (Fig. 2). This clone contained the nucleotide sequence corresponding to the oligonucleotide probe (85% identity to the “best-guess” probe) and agreed perfectly with the tryptic peptide sequence (16 of 16 residues). However, this clone did not encode the entire protein since not all of the tryptic peptide sequences were present in the open reading frame. The 5' 1.0-kb Eco RI fragment of λ5L5 was used to select 14 additional clones. The clone λ3R1 had an identical restriction map in the overlapping regions and contained an additional 1.0-kb 5' fragment (Fig. 2).

The composite sequence of λ5L5 and λ3R1 contains 5,139 nucleotides (Fig. 3). There is an open reading frame of 3,510 nucleotides, a 5' untranslated region of 94 nucleotides, and a 3' untranslated region of 1,535 nucleotides which contains a polyadenylation signal site 15 nucleotides before the poly(A+) tail. Within the 3' untranslated region there is a typical Alul repeat consisting of two tandem related sequences each terminated by an A-rich segment (16).

Northern blots demonstrate an LFA-1 α subunit mRNA of 5.5 kb in SKW3 T lymphoma cells, U937 myelomonocytic cells, and IB4 B lymphoblastoid cells (Fig. 4 A, lanes 1-3); however, no signal was detected in EJ bladder carcinoma cells (Fig. 4 A, lane 4). The mRNA expression is in agreement with the restriction of cell surface expression of LFA-1 to hematopoietic cells. The β subunit showed the same pattern of expression (Fig. 4 B). In Southern blots, the 1.2-kb 5' Eco RI–Bam HI fragment from clone λ2L2 hybridized to two fragments of 10 and 8 kb (8). A genomic clone isolated from a cosmid library possesses the same two fragments...
which are contiguous and hybridize with different regions of the cDNA (unpublished data) showing that the LFA-1 α subunit is a single copy gene.

**Protein Sequence**

All 105 amino acids determined by microsequencing of trypptic peptides were found in the translated open reading frame, confirming the authenticity of the cDNA clones (Fig. 3). Hydrophobicity analysis shows that the LFA-1 α subunit is a typical transmembrane protein with a 25-residue hydrophobic signal sequence, an extracellular domain of 1,063 residues, a single hydrophobic transmembrane region of 29 residues, and a short cytoplasmic tail of 53 residues. The NH2-terminal residue of human LFA-1 was identified by homology to the NH2-terminal sequence of murine LFA-1 (48) (Fig. 6, see below). Human LFA-1 has 55% identity with murine LFA-1 over the first 20 amino acids. A classical signal peptide with a consensus sequence (Ala-X-Ser/Pro) for the cleavage peptidase precedes the NH2-terminal sequence. There are three putative upstream transcription initiation sites (ATG) in frame. The use of the first initiation site (nucleotide position 89) is generally favored (24) and gives a 25-residue signal sequence with several NH2-terminal polar groups as is typically found in signal sequences (54).

The mature protein is \( M_r = 126,193 \). 12 N-linked glycosylation sites (Asn-X-Thr/Ser) are present in the extracellular domain. These findings are consistent with the size previously determined by SDS-PAGE for the in vitro translated murine LFA-1 α subunit \( M_r = 140,000 \), the murine and human LFA-1 α subunit glycoproteins \( M_r = 180,000 \) and 177,000 (44), and with previous studies on the glycosylation of LFA-1 (10, 32).

Within the extracellular domain there are seven internal repeats (Fig. 5). The degree of identity is highest among the three repeats (14.5–33%) located toward the COOH terminus which show a statistically significant relationship \( P < 10^{-6} \). The relationship among the four repeats located toward the NH2 terminus is weaker and is discernible by conservation of flanking sequences (Fig. 5); the homology between repeat IV and V is significant \( P < 10^{-4} \). The central regions of the three COOH-terminal repeats is similar with the EF hand divalent cation binding site motif, perhaps due to convergent evolution (Fig. 5). Previous studies have shown that Mg2+ alone or at lower concentration in conjunction with Ca2+ is necessary for ligand binding func-
Distinct Subfamilies of Integrin α Subunits

We compared the LFA-1 α subunit to other integrin α subunits (Fig. 6). The LFA-1 α subunit has striking and consistently higher homology to other leukocyte integrin α subunits (Mac-1, 35.7%; p150,95, 37.4%) than to ECM receptor integrin α subunits (vitronectin receptor, 26.4%; FNR, 27.8%; gpIIb, 30.2%). The ECM receptor integrin α subunits are more related to one another (~ = 41.9% SD = 3.7%) than to the leukocyte integrins (Fig. 8, see below). Further structural features distinguish the leukocyte and ECM receptor integrins. The leukocyte integrins contain an insertion of ~200 amino acids near the NH2-terminal region of the protein that is not present in the sequenced ECM integrins, data banks revealed that the LFA-1 domain of ~200 amino acids, which is not present in the sequenced ECM integrins, is homologous to the domains of the same size in vWF and a cartilage matrix protein (Figs. 7 and 8). These alignments have 20.4–32.1% identity and are statistically significant (P < 10^{-9} – 10^{-23}). Of the integrins sequenced to date, this 200-amino acid domain is unique to the leukocyte integrins. Homologous domains are inserted in several proteins and in well-studied examples have been documented to mediate interaction with ligand (see below). For this reason we will refer to the 200-residue region of the LFA-1, Mac-1, and p150,95 α subunits as the “I” (inserted/interactive) domain. The homology unit is present in three tandem repeats in vWF (A domains) (45) (P < 10^{-12} – 10^{-23} for comparison of the three repeats to one another) and two repeats separated by an EGF domain in CMP (2) (P < 10^{-23}). With the exception of the NH2-terminal region of the CMP repeat 1 which has not been sequenced, the repeating units in both vWF and CMP correspond precisely to the region homologous to the I domain, supporting the concept that this homology unit is a domain. Similar structural homologies have been noted for the murine and human Mac-1 α subunit (7, 38) and the degree of homology of Mac-1 and p150,95 with vWF and CMP is similar to that found with LFA-1 (17.8–31.6% identity, P < 10^{-9} – 10^{-23}). Factor B has previously been found to be homologous with the vWF A repeats (P < 10^{-9} – 10^{-7}) (45); factor B in turn is homologous but at a lower level with the LFA-1, Mac-1, and p150,95 I domains (P < 10^{-12} – 10^{-4}) (7, 38). In factor B a single homology unit is bounded on one side by the site for the cleavage which activates the Bzymogen to the active Bb fragment, and on the other side by the serine protease domain (6).

An Inserted Domain in the Leukocyte Integrins

Searches of the NBRF and SWISS-PROT protein sequence data banks revealed that the LFA-1 domain of ~200 amino acids, which is not present in the sequenced ECM integrins, is homologous to the domains of the same size in vWF and a cartilage matrix protein (Figs. 7 and 8). These alignments have 20.4–32.1% identity and are statistically significant (P < 10^{-9} – 10^{-23}). Of the integrins sequenced to date, this 200-amino acid domain is unique to the leukocyte integrins. Homologous domains are inserted in several proteins and in well-studied examples have been documented to mediate interaction with ligand (see below). For this reason we will refer to the 200-residue region of the LFA-1, Mac-1, and p150,95 α subunits as the “I” (inserted/interactive) domain. The homology unit is present in three tandem repeats in vWF (A domains) (45) (P < 10^{-12} – 10^{-23} for comparison of the three repeats to one another) and two repeats separated by an EGF domain in CMP (2) (P < 10^{-23}). With the exception of the NH2-terminal region of the CMP repeat 1 which has not been sequenced, the repeating units in both vWF and CMP correspond precisely to the region homologous to the I domain, supporting the concept that this homology unit is a domain. Similar structural homologies have been noted for the murine and human Mac-1 α subunit (7, 38) and the degree of homology of Mac-1 and p150,95 with vWF and CMP is similar to that found with LFA-1 (17.8–31.6% identity, P < 10^{-9} – 10^{-23}). Factor B has previously been found to be homologous with the vWF A repeats (P < 10^{-9} – 10^{-7}) (45); factor B in turn is homologous but at a lower level with the LFA-1, Mac-1, and p150,95 I domains (P < 10^{-12} – 10^{-4}) (7, 38). In factor B a single homology unit is bounded on one side by the site for the cleavage which activates the Bzymogen to the active Bb fragment, and on the other side by the serine protease domain (6).
Figure 6. Alignment and comparison of the human LFA-1 α subunit with the other members of the integrin superfamily (Mac-1 [4, 7], p150,95 [9], vitronectin receptor [50], fibronectin receptor [31], and platelet glycoprotein IIb [37]) and the NH2 terminus of the murine LFA-1 α subunit [48]. The residues common to LFA-1 and at least one other integrin are boxed. The area of the I domain is shown in Fig. 7. The protease cleavage site in the ECM receptor α subunits are indicated with black dots.
similar to the overall percent identity among the leukocyte integrins, implying that a single I domain incorporated into a primordial leukocyte integrin only once rather than independently. Then, the gene duplicated and gave rise to LFA-1 and a Mac-1/p150,95 primordial gene. Further duplication of the Mac-1/p150,95 primordial gene gave rise to Mac-1 and p150,95. This scheme is consistent with the observation that Mac-1 and p150,95 are more closely related to each other than to LFA-1 (35.7, 37.4%, respectively). Since members of the I domain superfamily serve important recognition functions in several proteins. The A1 domain of vWF binds to glycoprotein Ib and heparin (15) while both the A1 and A3 domains are involved in binding to collagen. The domain in factor B is located in a region of the molecule available for interaction with its ligand C3b (6, 34). The domain in CMP may also have an important role in interaction with collagen (2) and cartilage proteoglycan (35). These domains lack glycosylation and cysteines for the most part. Similarly, the I domain and the repeats with divalent cadcu binding sites in LFA-1 contain only one N-glycosylation site and cysteines for the most part. Similarly, the I domain and the repeats with divalent calcium binding sites in LFA-1 contain only one N-glycosylation site and cysteines for the most part.

Structural differences between the ECM receptor integrin α subunits and LFA-1 correlate with differences in recognition specificity; RGD containing peptides block the binding of the VNR, FNR, and gpIIb/IIIa to their ligands (41) but not binding of LFA-1 to ICAM-1 (29). Furthermore, ICAM-1

Figure 7. Alignment and comparison of the LFA-1 α subunit I domain with the homologous domains in Mac-1 (4, 7), p150,95 (9), vWF (45), factor B (34), and CMP (3).
does not contain an RGD sequence and unlike other known integrin ligands, is a member of the immunoglobulin gene superfamily. It will be of interest to determine whether the I domain confers specificity for non-RGD containing ligands.

This study demonstrates that the LFA-1 α subunit belongs to the integrin superfamily but possesses an additional domain. This I domain and homologous domains constitute a protein “domain” family that is of functional importance. Since LFA-1 is involved in a large number of leukocyte functions and may have more than one ligand (39) (Dustin, M. L., and T. Springer, manuscript in preparation), it is possible the more than one functional domain exists in the LFA-1 α subunit. The availability of cDNA clones for the α and β subunits will allow these and other structure-function relationships to be examined.

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