An Immortalized Osteogenic Cell Line Derived from Mouse Teratocarcinoma Is Able to Mineralize In Vivo and In Vitro

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Abstract. The hybrid plasmid pK4 containing the early genes of the simian virus SV-40, under the control of the adenovirus type 5 Ela promoter, was introduced into the multipotent embryonal carcinoma (EC) 1003. Expression of the SV-40 oncogenes was observed at the EC cell stage, and this allowed the derivation of immortalized cells corresponding to early stages of differentiation.

Among the immortalized mesodermal derivatives obtained, one clone, C1, is committed to the osteogenic pathway. C1 cells have a stable phenotype, synthesize type I collagen, and express alkaline phosphatase activity. Although immortalized and expressing the SV-40 T antigen, the cells continue to be able to differentiate in vivo and in vitro. In vivo, after injection into syngeneic mice, they produce osteosarcomas. In vitro, the cells form nodules and deposit a collagenous matrix that mineralizes, going to hydroxyapatite crystal formation, in the presence of β-glycerophosphate. This clonal cell line, which originates from an embryonal carcinoma, therefore differentiates into osteogenic cells in vivo and in vitro.

This immortalized cell line will be useful in identifying specific molecular markers of the osteogenic pathway, to investigate gene regulation during osteogenesis and to study the ontogeny of osteoblasts.

One current view of the development of bone cells is that osteocytes derive from primitive mesoblastic cell precursors through a cascade of events. These involve, in the case of intramembranous ossification, (a) proliferation of primitive mesoblasts, (b) differentiation of these cells into an osteoprogenitor cell, then into osteoblasts, and (c) maturation of osteoblasts with synthesis of a collagen matrix and mineralization (5, 10, 39, 44). Up until now, models for following bone differentiation in vitro have been of two types: (a) osteoblast-like clones explanted from normal bone, and (b) clones derived from osteogenic osteosarcomas.

(a) The former are normal explanted cells capable of progressively synthesizing a bone-like tissue in the presence of organic phosphate and ascorbic acid (1, 3, 8, 9, 28, 35, 46). This process and its hormonal regulation can therefore be studied in vitro. Collagen I maturation, in particular, has been extensively studied (11). However, the potential for division of such cells remains limited, and this precludes molecular studies. In addition, phenotypic changes with loss of osteoblastic properties often occurs in culture (47).

(b) Clones derived from rat osteosarcomas have also been useful in investigating the effects of hormones (parathyroid hormone; prostaglandins) and vitamins (retinoids and vitamin D3) on bone development (17, 25, 29, 30, 41). The formation of a mineralized matrix has, however, been shown to require either the implantation of the tumoral cells in diffusion chambers within the animal (43), or the growth of the cells in agar (37).

In both model systems, the major problems encountered are the heterogeneity of the cell population within a clone, and the lack of specific markers allowing formal identification of the progressive stages of osteoblast maturation.

In this study, we introduce mouse teratocarcinoma as a new model for studying bone differentiation. This system has already proven useful in the study of determination and differentiation mechanisms in early embryogenesis. Teratocarcinoma tumors contain a wide variety of differentiated tissues derived from a multipotent stem cell, the embryonal carcinoma (EC) (19). The differentiated derivatives, however, have a limited potential for division, preventing the isolation of pluripotent progenitors committed to the various restricted cell lineages. We have recently developed a strategy for immortalizing such stem cells from teratocarcinomas. A recombinant plasmid, pK4, was constructed, in which the SV-40 oncogenes were linked to the adenovirus 5 Ela promoter. This promoter is active in early embryonic cells (6). In a first series of experiments, the plasmid was introduced into the EC F9 (20, 22). Immortalized cells were obtained after retinoic acid induction. From these cells, precursors of different lineages could be selected and cloned (21). Osteogenic cells were not found among the mesodermal derivatives.

In a series of parallel experiments, pK4 was introduced into 1003 (31), a multipotent EC cell line known to undergo

1. Abbreviation used in this paper: EC, embryonal carcinoma.
mesodermal differentiation with high efficiency (7). We show in this paper that expression of the SV-40 oncogenes was observed in 1003-pk4 cells as early as the EC cell stage. Expression did not prevent further differentiation because the tumors obtained after injection to syngeneic animals contained differentiated cells representative of the three embryonic germ layers: neuroectoderm, endoderm, and mesoderm. Among the immortalized mesodermal derivatives, we were able to select, on the basis of its high expression of the bone-specific alkaline phosphatase, a clone, C1, committed to the osteogenic pathway. C1 maintains a stable phenotype during exponential growth. Upon differentiation in vivo, it gives rise to typical osteosarcomas. In the presence of appropriate inducers, it mineralizes in vitro, forming dense calcified nodules.

**Material and Methods**

**Cell Culture and Differentiation Conditions**

1003 cells (7) and their derivatives were routinely grown in DME (Gibco Laboratories, Grand Island, NY) supplemented with 10% FBS (Gibco Laboratories).

Cloning by limiting dilution (0.3 cells per 6-mm microtiter wells) was performed in multishes (Costar Corp., Cambridge, MA). Clones 1003 transformed by pk4 were obtained after cotransfection of the embryonal carcinoma 1003 with plasmids pk4 and PSVinKneoα using the calcium phosphate precipitation technique as described (21). An intense staining of >10% of the cells with anti-SV-40 T antibodies was detected 48 h after the transfection. Colonies resistant to Geneticin (Sigma Chemical Co., St. Louis, MO, 400 μg/ml) were isolated and subcloned.

C1 cells were routinely grown on tissue-culture dishes in DME supplemented with 10% FBS. To induce in vitro mineralization, C1 cells were first seeded on untreated plastic dishes (Sterilin, Ltd., Hounslow, UK). After 8 or 15 d, the three dimensional clusters formed were either plated onto tissue-culture dishes on which they reattached, or maintained as aggregates. After 2 d, the medium was shifted to DME supplemented with 1% FBS, and some dishes were treated with ascorbic acid (50 μg/ml. Sigma Chemical Co., St. Louis, MO) and β-glycerophosphate (7 mM, Sigma Chemical Co.). The nodules, which developed rapidly, were fixed for light or electron microscopy.

**Tumor Formation**

2-4 × 10⁶ cells in 1 ml DME were injected subcutaneously to irradiated (600 rad) C3H syngeneic mice. One part of the 1003-pk4-derived tumors was fixed in Bouin's fluid and processed for routine histological analysis. Another part of the tumor was minced and allowed to grow and differentiate in vitro. C1-derived tumors were fixed in 2% glutaraldehyde, dehydrated in alcohol and embedded undecalcified in methacrylate (2, 16). Thick sections (5 μm) were cut according to conventional methods (2).

**Southern Blot Hybridization of Genomic DNA**

Aliquots (10 μg) of genomic and pk4 DNAs were digested by restriction endonucleases, electrophoresed on a 0.8% agarose gel and blot-transferred using zetabind membranes (Cuno, Inc., Meriden, CT). Hybridization was carried out using a pk4 probe nick-translated using an Amersham kit (Amersham International, Amersham, UK) (3 × 10⁶ cpm/μg).

**Immunocytochemistry**

Indirect immunofluorescence studies on cells were carried out using several antisera: (a) mouse mAb Pab 419 (32) detecting viral T antigens and mouse mAb ECMA 7 directed against specific EC cell-surface antigens (23); (b) affinity-purified rabbit antibodies raised against basal membrane components: collagen I and III (14). (c) species-specific secondary antisera coupled to fluorophores (Cappel Laboratories, Malvern, PA) were used to visualize sites of primary antibody binding.

Fixation and permeabilization of the cells are described in the figure legends. Controls consisted of cells incubated without primary antibody or with nonimmune serum.

**Biochemical Measurements of CAMP**

The response of C1 cells to 1-34 human parathyroid hormone (PTH; Sigma Chemical Co.), prostaglandin E2 (PGE2; Sigma Chemical Co.) and salmon calcitonin (CT; kindly provided by Rorer Laboratories, Gennevilliers, France) was evaluated by determination of the intracellular production of CAMP. Confluent cells were treated for 10 min with PTH or the solvent (PBS), PGE2 or the solvent (0.1% ethanol) or CT, or the solvent (PBS). Intracellular CAMP concentrations were determined in cell lysates using a specific RIA (Immunotech, Luminy, France).

**Biochemical Measurements of Alkaline Phosphatase Activity**

This was carried out according to previously reported methods (28). The thermosensitivity of the enzyme was checked according to Goldstein et al. (12).

**Histological Detection of Collagen and Minerals**

After 15 d of culture in the presence or absence of ascorbic acid and β-glycerophosphate, the nodules formed were collected, fixed in 95% ethanol, and embedded in glycolmethacrylate. 5-μm-thick sections were stained with toluidine blue, Von Kossa, or Goldner Trichrome for the detection of collagen and minerals (2, 28).

**Electron Microscopy, X-Ray Microanalysis, and X-Ray Diffraction**

The calcified nodules formed in differentiated C1 cultures were fixed in situ with 2% glutaraldehyde in 0.1 M sodium cacodylate (pH 7.2, 20 min). The nodules were post-fixed in 1% osmium tetroxide (15 min), rinsed, dehydrated in acetone, and embedded in Epon. Ultra-thin sections stained with 2.5% uranyl-acetate and lead citrate were examined using a Siemens Elmiskop 101 electron microscope (Siemens-Allis, Inc., Cherry Hill, NJ). Mineralized spots were analyzed on carbon-coated sections using energy dispersive x-ray microanalysis (Tracor Northern, Middleton, WI). X-ray diffraction analysis of the calcifying structure was performed and calibrated using an aluminum standard.
Results

Selection and Cloning of an Immortalized Osteogenic Clone from pK4-transformed Teratocarcinoma

Expression of the pk4 Genome at the EC Cell Stage in Transformed 1003-pK4 Clones. 1003 EC cells were cotransfected with plasmids pk4 and PSVtk-neoβ. Geneticin resistant clones were then selected. Out of 15 clones analyzed, 3 (clones 8120, 8212, and 8129) had integrated the pK4 plasmid, as shown by analysis of the cellular DNA by Southern blot hybridization. As indicated by the restriction fragment pattern, the integrated plasmid DNA was grossly intact. 1-3 integrated copies were detected depending on the analyzed clone (Fig. 1, top). All three clones expressed the SV-40 T antigen in 100% of cell nuclei (Fig. 1, C and D).

Further analysis of the transformed clones (1003-pK4) revealed that the cells had remained undifferentiated, as judged by their morphology, the persistence of the expression of specific markers of EC cells (4, 23) (Fig. 1, A and B), and their potential to differentiate, when submitted to a suitable environment, into derivatives of the three embryonic germ layers. Except for the integration of the plasmid and the expression of the SV-40 oncogenes, the 1003-pK4 cells could not be distinguished from the parental EC 1003. We conclude that the SV-40 T antigen can be expressed in 1003-pK4 cells as early as the EC stage, and that the presence of the T antigen neither causes recognizable phenotypic change nor prevents differentiation in vivo or upon in vitro induction.

Selection of an Osteogenic Clone among Immortalized Mesodermal Derivatives. Each of the three 1003-pK4 clones was injected into irradiated syngeneic mice. Teratocarcinomas were produced within 1-3 mo in all inoculated animals. They were composed of EC cells, and of various differentiated derivatives of the three germ layers. In cultures derived from the tumor, most cells expressed the SV-40 T antigen.

One tumor arising from 1003-pK4 clone 8129 was allowed to grow in vitro. Areas containing mature mesodermal derivatives closely associated with immature cells were selected with the idea that the immature cells might correspond to mesodermal progenitors. Clones harboring various phenotypes were isolated from this mesodermal population by limiting dilution in Costar multidishes (0.3 cell per well). All clones expressed the SV-40 T antigen (Fig. 2, A and B). Among them, myoblast-like clones giving rise at confluence to multinucleated myotubes (Fig. 2, C and D). Some clones were enriched in adipoblasts and adipocyte-like cells containing lipid droplets (Fig. 2 E). Others were fibroblast-like (Fig. 2 F) or exhibited a mixed phenotype (Fig. 2 H). Finally, some clones were composed of polygonal cells of unidentified phenotype (Fig. 2 G). To screen for the presence of an osteogenic precursor among the immature cells, the various subpopulations were injected into syngeneic mice. The tumors formed contained only mesodermal derivatives. The polygonal cells gave rise in all cases to tumors containing mesoblastic cells, osteoblast-like cells oriented around host capillaries, and developing bone. Transitional stages between the immature cells and mineralized bone were present in all the tumors (not shown). The cell population arising from one tumor (87102) was further cloned by limiting dilution. Clones were selected on the basis of their polygonal morphology and of their high level of bone specific heat-labile alkaline phosphatase activity (Table I). One clone, C1, was selected for further analysis.

Properties of the Osteogenic Clone C1

Characteristics of the Precursor Cell. C1 cells exhibit a polygonal morphology (Fig. 3 A). All the cells of the clone, ex-
Figure 2. Phenotypes of the mesodermal clones derived from a 1003-pK4 teratocarcinoma. (A) Phase-contrast and (B) immunofluorescence labeling with mAb against T antigen of an immature, mesoblastic like clone. Bar, 10 μm ×800. (C and D) Myoblast-like cells and the same clone showing multinucleated myotubes at confluence. (E) Adipocytes. (F) Fibroblasts. (G) Polygonal cells of unidentified phenotype. (H) mixed phenotype. Bar, 50 μm. ×200.
Table I. Alkaline Phosphatase Activity

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<th></th>
<th>Total activity</th>
<th>Heat-labile activity</th>
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<tr>
<td></td>
<td>μmol PNP/mg protein per min</td>
<td></td>
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<tr>
<td>Embryonal carcinoma 1003</td>
<td>0.01 ± 0.001</td>
<td>0.01 ± 0.002</td>
</tr>
<tr>
<td>Embryonal carcinoma 1003-pK4</td>
<td>0.01 ± 0.002</td>
<td>0.01 ± 0.001</td>
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<tr>
<td>Tumor 87102</td>
<td>0.21 ± 0.02</td>
<td>0.20 ± 0.02</td>
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<tr>
<td>Clone F1</td>
<td>0.02 ± 0.003</td>
<td>0.02 ± 0.003</td>
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<tr>
<td>Clone C1</td>
<td>0.23 ± 0.05</td>
<td>0.27 ± 0.03</td>
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Cells were plated in DME containing 10% FBS at 4 × 10^4 cells/cm^2. When the cells were confluent, they were first washed with DME and then cultured for a further 24 h in serum-free DME. The cells were scraped, cell lysates were collected and sonicated, and alkaline phosphatase activity was determined in cell lysates. Values are the mean ± SEM of four independent confluent cultures.

Express the SV-40 T antigen in their nucleus (Fig. 3 B). When growing exponentially, the clonal cell population has a doubling time of 30 h and maintains a stable, homogeneous phenotype for over 100 generations.

C1 cells synthesize type I collagen, a characteristic feature of osteoblastic cells (1). At confluence, collagen represents 8.2 ± 1.4% (n = 4) of total protein, as assessed by collagenase digestion of [3H]proline-labeled C1 cells. SDS-PAGE analysis of the proteins digested with pepsin shows two major bands comigrating with standard α1(1) and α2(1) collagen chains. Collagen type III chains are not detectable (Figs. 4 and 5A).

Table II. Intracellular cAMP Production by C1 Cells

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<thead>
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<th>cAMP</th>
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<tr>
<td></td>
<td>pmol/mg protein</td>
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<tr>
<td>Controls (PBS)</td>
<td>11.8 ± 0.2*</td>
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<tr>
<td>(1-34) hPTH</td>
<td>10^-10</td>
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<tr>
<td></td>
<td>20.6 ± 0.8*</td>
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<td></td>
<td>10^-9</td>
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<tr>
<td></td>
<td>49.7 ± 9.3*</td>
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<td></td>
<td>10^-8</td>
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<tr>
<td></td>
<td>89.1 ± 9.6*</td>
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<td></td>
<td>10^-7</td>
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<tr>
<td></td>
<td>102.7 ± 6.2*</td>
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<tr>
<td>Controls (PBS)</td>
<td>7.0 ± 1.5</td>
</tr>
<tr>
<td>CT</td>
<td>10^-8</td>
</tr>
<tr>
<td></td>
<td>7.1 ± 0.8</td>
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<tr>
<td></td>
<td>10^-9</td>
</tr>
<tr>
<td></td>
<td>4.4 ± 0.8</td>
</tr>
<tr>
<td>Controls (ethanol)</td>
<td>2.7 ± 0.2</td>
</tr>
<tr>
<td>PGE2</td>
<td>10^-4</td>
</tr>
<tr>
<td></td>
<td>3.6 ± 0.4</td>
</tr>
<tr>
<td></td>
<td>10^-7</td>
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<td></td>
<td>4.3 ± 0.25*</td>
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<tr>
<td></td>
<td>10^-6</td>
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<td>7.2 ± 0.6*</td>
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C1 cells were cultured in DME with 10% FBS. At confluence cells were washed, cultured in DME and 0.1 mM isobutylmethylxantine (IBMX) for 1 h. The cells were then washed and treated in DME with 0.1 mM IBMX for 10 min in presence of the compounds or their vehicle. The cells were scraped and cAMP concentrations were determined in cell lysates. Values are the mean ± SEM of three to four independent confluent cultures.

* Significant difference with the appropriate control (P < 0.05 or better level of significance by t test).

Alkaline phosphatase activity is revealed in most of the C1 cells. The parental clones 1003 and 1003-pK4 also express alkaline phosphatase, a marker of embryonal carcinomas and of germinal cells (4). However, as illustrated in Table I, alkaline phosphatase activity levels present in extracts of C1 cells are much higher than those in 1003 and 1003-pK4, or in the fibroblastic-like clone F1. The osteosarcoma 87102 from which C1 and F1 were cloned, also shows a high level of alkaline phosphatase activity.

Moreover, most of the C1 enzymatic activity (95%) is lost upon heating to 56°C for 30 min (Table I), a characteristic of bone-specific alkaline phosphatase (12). In addition, 59.6% of the alkaline phosphatase activity (p < 1,000) is inhibited by physiological doses of 1,25 dihydroxyvitamin D3 [1,25(OH)₂D] (10^-4 M). Furthermore, C1 cells are responsive to PTH and PGE2 but not to CT in terms of stimulation of cAMP production, another characteristic feature of osteoblastic cells (Table II) (29, 30).

A significant amount of acid phosphatase activity is found in the cytoplasm of the cells (not shown). Such an activity was recently demonstrated in vivo in rat osteoblasts and osteocytes (5).

Differential In Vivo. When injected subcutaneously into irradiated syngeneic mice, the SV-40-transformed C1 cells develop tumors, found to be typical osteosarcomas. The tumors maintain their histological features after serial passages in vitro and in vivo. They form a dense trabecular bone network with extensive vascularization. Several trabeculae of calcified bone matrix are regularly arranged within the osteosarcoma. Numerous bone forming cells are dispersed along the bone matrix, some of which are embedded like osteocytes (Fig. 6). Bone resorption by osteoclast-like cells appears in some areas, but host marrow cells are not apparent.

C1 cells are therefore able to form and mineralize in vivo a calcified matrix similar to immature spongiosa. The osteosarcoma cells must moreover originate from the in vivo
determination of CI since they express the SV-40 T antigen in the tumor.

**Discussion**

In a recent report (21), we have introduced a method to derive various immortalized precursors of the neuroectodermal, mesodermal, and endodermal pathways from the embryonal carcinoma F9. This method is based upon the transformation of F9 cells by a plasmid, pK4, carrying the SV-40 early genes under the control of the adenovirus Ela promoter. The transformed F9 cells differentiated upon addition of retinoic acid to the cultures (20, 21). The expression of SV-40 oncogenes in the transformed F9 cells accompanied the differentiation.

In this work, the plasmid pK4 is introduced into another EC cell line, 1003. In contrast with the case of F9 EC cells, expression of the SV-40 oncogenes in 1003 cells is realized as soon as the EC stage and does not require the induction of differentiation. Note that the oncogene expression does not induce differentiation (33, 34) and does not alter the multipotentiality of the EC cells, as indicated by the presence of the expected differentiated derivatives of the three germ layers within tumors from mice inoculated with the transformed 1003 clones. In particular, among the derivatives, committed precursors of the mesodermal lineage were cloned.

This study both generalizes and extends our earlier observations. The recombinant plasmid pK4 is clearly of a potential interest for delineating molecular mechanisms leading to the expression of the SV-40 T antigen in embryonal carcinoma cells through comparative studies of 1003 and F9 cells (24, 26, 42). pK4 also appears to be a valuable tool for establishing stable cell lines corresponding to progenitor cells. It promotes immortalization at an early stage, while still allowing the expression of differentiation programs.

This study is devoted to the characterization of an osteogenic clone, CI. CI isolation required several steps: transfection of an embryonal carcinoma cell line known for its high efficiency of mesodermal differentiation, selection of a tumor giving rise to many mesodermal derivatives, cloning from this tumor in search of the presence of osteogenic clones among mesodermal derivatives, and finally selection of the clones for their osteogenic potential in vivo and their high activity of bone-specific alkaline phosphatase in vitro (18, 27).

CI cells are clearly committed towards the osteogenic pathway. (a) After injection into syngeneic mice, they produce typical osteosarcomas which indeed originate from the de novo differentiation of CI, as shown by the expression of the SV-40 T antigen in the tumors. (b) In vitro, CI cells express osteoblast phenotypic markers, including synthesis of type I collagen (I), expression of the bone-specific alkaline phosphatase isoenzyme (12) and osteogenic potential in vivo (24, 26). pK4 also appears to be a valuable tool for establishing stable cell lines corresponding to progenitor cells. It promotes immortalization at an early stage, while still allowing the expression of differentiation programs.
Figure 5. In vitro differentiation of CI cells. (A) Immunofluorescence with anticollagen I antibodies showing intracellular staining of exponentially growing cells. Fixation with 3% pFA, then with methanol. Bar, 10 μm. ×800. (B) Calcified nodule grown in the presence of ascorbic acid and β-glycerophosphate. Bar, 50 μm. ×100. (C–F) Transverse sections (5 μm thick) of undecalcified nodules. Culture in suspension for 21 d in the absence (C and E) or in the presence (D and F) of β-glycerophosphate and ascorbic acid. (C) Intense membrane-bound alkaline phosphatase activity without mineralization. (D) Toluidine blue-stained section showing a large mineralized matrix (m) surrounded by alkaline-phosphatase-positive cells. (E) Von Kossa staining showing the absence of mineralization. (F) Calcified matrix and osteocyte-like cells (arrows) embedded within the mineralized matrix. Bar, 50 μm. ×200.

mice, bone formation arising after injection of a teratocarcinoma-derived untransformed cell line has been described (36). However, the resulting osteochondrosarcoma tumors were not formally related to the in vivo differentiation of the injected cells and no clonal cell line expressing in vitro an osteogenic phenotype could be obtained. It is noteworthy to mention also the work of Silbermann et al. (44) who used viruses containing an active v-fos oncogene to transform in vitro embryonic murine condyles. After injection in mice, highly abnormal bone-like metastatic tumors were obtained. However, these metastatic tumors contrast with our CI cell system, because the bone obtained upon CI differentiation has histological features typical of normal immature bone, and since the CI-derived tumors have no metastatic properties despite the expression of the early SV-40 oncogenes. Finally, a recombinant retrovirus containing the SV-40 T antigen was recently used to immortalize rat calvaria cells. The immortalized cell lines obtained were shown to retain osteoblastic properties (15). As compared with the Calvaria cell lines, CI cells have two additional features. Firstly, the
Figure 6. Section of an osteosarcoma derived from C1 cells: dense trabecular bone network with extensive vascularization. Osteocytes embedded in the bone matrix (arrows). Bar, 50 μm. ×200.

cells are osteogenic in vitro, allowing the study of gene regulation during matrix mineralization. Secondly, it should be recalled that the C1 clone derives from an island of cells resembling the somitic mesoderm of the early embryo. This island was composed of mesoblastic cells able to differentiate into either myogenic, fibroadipogenic, or osteogenic derivatives. The study of various clones representative of those mesoblastic cells is now under way. Hopefully, some of these clones may correspond to immortalized stages of bone commitment earlier than the C1 stage. This idea is supported by a recent report (13) showing that progenitor cells in a bone-derived clonal cell population from rat can differentiate into bone as well as into muscle, fat, or cartilage. Therefore, our immortalized teratocarcinoma-derived system appears to be of interest to study also the regulation of mesenchymal stem cell differentiation.

In conclusion, the teratocarcinoma-derived clone C1 offers the advantages associated with both osteosarcoma-derived cell lines having the potential for proliferation, and osteoblastic-like clones derived from normal bone, having the ability to differentiate in vitro into osteogenic cells. The potential of the C1 cells for proliferation and their stable phenotype will give the possibility to obtain large amounts of homogeneous material corresponding either to exponentially growing precursor cells or to cells induced to differentiate. Differential screening with other mesoblastic clones and with the initial embryonal carcinoma may permit the identification of specific molecular markers for the osteogenic pathway (38), the investigation of gene regulation during osteogenesis, and the study of the ontogenesis of osteoblasts.

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Figure 7. Electron micrograph of the mineralized matrix: C1 osteogenic cells are surrounded by banded collagen fibers. The mineralized collagen form a dense calcified matrix. Bar, 1 μm. ×24,000. The deposition of minerals is closely associated with collagen fibers (inset, ×22,600).
Ligue Nationale Française contre le Cancer, and the Fondation André Meyer.

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References


Figure 8. (A) Energy-dispersive x-ray spectrum of a mineralized area of a nodule, showing major peaks for calcium (Ca) and phosphorus (P). (B) Electron diffraction pattern of the mineral shown in Fig. 7. The mineral shows a crystal- line structure characteristic of hydroxyapatite.