Microinjection of the Catalytic Fragment of Myosin Light Chain Kinase into Dividing Cells: Effects on Mitosis and Cytokinesis

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Abstract. Myosin light chain kinase (MLCK) is thought to regulate the contractile activity in smooth and non-muscle cells, and may play an important role in controlling the reorganization of the actin–myosin cytoskeleton during cell division. To test this hypothesis we have microinjected the 61-kD catalytic fragment of MLCK into mitotic cells, and examined the effects of unregulated MLCK activity on cell division. The microinjection of active 61 kD causes both a significant delay in the transit time from nuclear envelope breakdown to anaphase onset, and an increase in motile surface activity during and after metaphase. Control experiments with intact MLCK or with inactive catalytic fragment suggest that these effects are specifically induced by the unregulated myosin light chain kinase activity. Immunofluorescence analysis suggests that delays in mitosis are coupled to disruptions of spindle structures, while increased surface motility may be related to changes in the organization of actin and myosin at the cell cortex. Most importantly, despite the expression of strong phenotypes, 61 kD-injected cells still form functional cleavage furrows that progress through cytokinesis at rates identical to those of control cells. Together, these results suggest that the activity of MLCK can affect mitosis and cortical activities, however additional control mechanisms are likely involved in the regulation of cytokinesis.

ACTIN and myosin undergo precisely regulated reorganizations in cultured animal cells during mitosis, including the disassembly of stress fibers during prophase/prometaphase (Sanger et al., 1989), the increase in cortical association during metaphase/anaphase (Kitanishi-Yumura and Fukui, 1989; Cao and Wang, 1990b), and the redistribution into the equatorial region during cytokinesis (see Mabuchi, 1986; Salmon, 1989 for reviews). Although the exact roles of these activities are unclear, it is generally agreed that the interaction of cortical actin and myosin II generates the contractile force for cytokinesis (Mabuchi, 1986; Salmon, 1989 for reviews). The involvement of myosin II in cytokinesis, in particular, is supported by a large body of evidence, including the presence of myosin in the equatorial region during cell cleavage (Fujiiwara and Pollard, 1976, 1978; Aubin et al., 1979; Nunnally et al., 1980; Yumura et al., 1984; Maupin and Pollard, 1986; Schroeder, 1987; Sanger et al., 1989), and the ability of microinjected antibodies against myosin to inhibit cytokinesis (Mabuchi and Okuno, 1977; Kiehart et al., 1982; Zurek et al., 1990). Moreover, inhibition of cytokinesis has also been observed with genetically manipulated cells lacking myosin II (Knecht and Loomis, 1987) or expressing defective myosin II molecules (De Lozanne and Spudich, 1987).

While the requirement of myosin II for cytokinesis is well established, important questions remain as to how the interactions between actin and myosin II are spatially and temporally regulated in dividing cells. For both smooth- and non-muscle cells, one of the best-characterized mechanisms for the regulation of actin–myosin interactions is through phosphorylation of the 20-kD regulatory myosin light chain (MLC20) by the Ca2+/calmodulin-dependent enzyme myosin light chain kinase (MLCK) (Kamm and Stull, 1985; Sellers and Adelstein, 1987). This enzyme catalyzes the specific phosphorylation of MLC20 on serine 19, which in turn stimulates the actin-activated MgATPase of myosin, the assembly of myosin filaments under physiological conditions, and the ability of myosin to generate force ( Sellers and Adelstein, 1987; Warrick and Spudich, 1987; Korn and Hammer, 1988). Thus, it has been proposed that differences in the phosphorylation state of MLC20 may control the organization and contractile activity of myosin during cell division (Pollard et al., 1990; Mabuchi, 1990).

One way to test this hypothesis is to overcome the dependence of MLCK on Ca2+/calmodulin, and ask whether cell division is influenced by the unregulated catalytic activity of the kinase. Ikebe and colleagues have previously mapped the domain structure of MLCK and characterized a 61-kD tryptic fragment that possesses the catalytic activity even in the

1. Abbreviations used in this paper: MLCK, myosin light chain kinase; NEB, nuclear envelope breakdown; NRK, normal rat kidney.
absence of the Ca\(^{2+}\)/calmodulin complex (Ikebe et al., 1987a, 1989). Microinjection of this fragment into smooth muscle cells induces contraction without the requirement for the release of Ca\(^{2+}\) (Itoh et al., 1989). In this study, we have prepared this catalytic fragment and microinjected it into mitotic normal rat kidney (NRK) cells soon after nuclear envelope breakdown (NEB). Interestingly, the microinjection caused a significant delay in mitosis during prometaphase/metaphase and an increase in surface activities during metaphase, anaphase, and telophase. However, after the onset of anaphase, the cleavage furrow formed at the proper time and location in the cell, at a rate identical to that of controls. We conclude that while the regulation of MLCK activity can affect mitosis and cortical activities, the control of cytokinesis probably involves additional factors.

**Materials and Methods**

**Preparation of MLCK and its Catalytic Fragment**

MLCK was prepared from turkey gizzards by the method of Walsh et al. (1983), as modified by Ikebe et al. (1987a). After phosphocellulose chromatography, MLCK was further purified by calmodulin-affinity chromatography as described by Adelstein and Klee (1982) and dialyzed into a storage buffer of 50 mM KCl, 30 mM Tris- HCl, 0.5 mM DTT, pH 7.5.

The catalytic fragment of MLCK was prepared by limited proteolysis of MLCK in the presence of Ca\(^{2+}\) and calmodulin as detailed by Ikebe et al. (1987a, 1989). Briefly, MLCK (0.5 mg/ml) in the storage buffer containing 0.2 mM CaCl\(_2\) was protelyzed with trypsin (1:100, wt/wt) in the presence of 0.2 mg/ml calmodulin for 7.5 min at 25°C. The reaction was terminated by the addition of a twofold (wt/wt) excess of soybean trypsin inhibitor over trypsin, and the solution was subjected to anion-exchange chromatography on either a DEAE-Sepharose Fast Flow exchanger (Pharmacia LKB Biotechnology, Inc., Piscataway, NJ) or a TSK DEAE-5PW HPLC column (Pharmacia LKB Biotechnology, Inc.). DEAE chromatography was eluted with a 0.2 mMCaCl\(_2\) was proteolyzed with trypsin (1:100, wt/wt) in the presence of 0.2 mM CaCl\(_2\), 25 mM Tris-HCl, 5 mM MgCl\(_2\), 0.5 mM DTT, pH 7.5, and peak fractions containing the 61-kD catalytic fragment of MLCK were pooled and further purified by hydroxylapatite chromatography (Ikebe et al., 1987a). The purified 61-kD fragment was concentrated to 10 μg/ml, dialyzed against storage buffer, and stored at 4°C.

The extent and rate of MLCK phosphorylation was assayed by incubating MLCK (10 μg/ml) with the 61-kD fragment (5 μg/ml) in a buffer of 150 mM NaCl, 25 mM Tris-HCl, 5 mM MgCl\(_2\), 0.5 mM DTT, pH 7.5, containing either 0.2 mM CaCl\(_2\)/5 μg/ml calmodulin or 1 mM EGTA (and 5 μg/ml calmodulin in assays of MLCK). Reactions were initiated by the addition of ATP to 1 mM. 50-μl aliquots of reaction mixtures were removed at various times, precipitated with 3-5 % trichloroacetic acid (TCA), and the supernatant was assayed for phosphorus content using a phosphorus assay kit (Bio-Rad, Hercules, CA) and a low-light level ISIT video camera (Dage-MTI, Inc., Michigan City, IN). Images were stored as digital files in an image-processing computer, which was assembled by G. W. Hannaway and Associates (Boulder, CO) and consisted of a Silicon Graphics 4D/20 workstation (Mountain View, CA) and image-processing boards from Imaging Technology (150 series; Woburn, MA). Samples of phases were used to determine the timing of key stages of mitosis, and motile activities were analyzed using a motion display program as described previously (Wang, 1989).

**Fluorescence Localization of F-actin, Myosin, and Tubulin**

Cells were followed for various periods of time before fixation. Cells stained for F-actin and myosin were fixed for 10 min with 3.2% formaldehyde (EM grade; Electron Microscopy Sciences, Fort Washington, PA) in PHEM buffer (60 mM Pipes, 25 mM Hapes, 10 mM EGTA, 2 mM MgCl\(_2\); pH 6.1; Schliwa and van Blerkom, 1981), followed by a 5-min postfixation/extraction in acetone at 20°C. Samples were then blocked with PBS containing 1% BSA for 30 min and reacted overnight at 4°C with a 1:20 dilution of an anti-myosin mAb (Klotz et al., 1986; Amac Inc., Westbrook, ME). Similar results were obtained with polyclonal anti-platelet myosin antibodies kindly provided by Drs. Ira Herman (Tufts University School of Medicine) and Keigi Fujiwara (National Cardiology Research Center, Osaka, Japan). After extensive washes with PBS/BSA, the samples were reacted with a 1:100 dilution of fluorescein conjugated sheep-anti-mouse secondary antibody (Sigma Chemical Corp., St Louis, MO). Cells were then stained with 0.2 μM rhodamine phalloidin (Molecular Probes) for 45 min, washed with PBS, and examined by fluorescence microscopy. For microtubule staining, fluorescence images were obtained with a Zeiss (Thornwood, NY) IM35 inverted microscope equipped with a 40×/NA 0.65 achromat objective, and a low-light level ISIT video camera (Dage-MTI, Inc., Michigan City, IN). Images were stored as digital files in an image-processing computer, which was assembled by G. W. Hannaway and Associates (Boulder, CO) and consisted of a Silicon Graphics 4D/20 workstation (Mountain View, CA) and image-processing boards from Imaging Technology (150 series; Woburn, MA). Sequences of phase images were used to determine the timing of key stages of mitosis, and motile activities were analyzed using a motion display program as described previously (Wang, 1989).

**Figure 1. Preparation of the Catalytic Fragment of MLCK by Limited Tryptic Digestion.**

**Summary of the purification procedure by SDS-PAGE:** lane 1, MLCK (7 μg); lane 2, digestion of MLCK with trypsin in the presence of Ca\(^{2+}\)/calmodulin; lane 3, pooled fraction from TSK DEAE-SPW HPLC; lane 4, 61-kD fragment after hydroxylapatite chromatography (3 μg). Molecular weight standards (M\(_s\)) are in the far left and far right lanes, as indicated: 205 kD, myosin heavy chain; 116 kD, β-galactosidase; 97 kD, phosphorylase b; 66 kD, BSA; 45 kD, ovalbumin; 30 kD, carbonic anhydrase; 20 kD, soy bean trypsin inhibitor; 14 kD, α-lactalbumin.
Figure 2. Phosphorylation activity of MLCK and the 61-kD catalytic fragment prepared for microinjection studies. MLC\textsubscript{20} is phosphorylated by MLCK in the presence of the Ca\textsuperscript{2+}/calmodulin complex (open triangles), but not in the presence of EGTA and calmodulin (closed triangles). In contrast, the 61-kD catalytic fragment has a strong activity both in the presence of EGTA (closed circles) and in the presence of Ca\textsuperscript{2+} (data not shown). After dialysis into a low ionic strength buffer or extended periods of storage on ice, the 61-kD fragment shows a greatly reduced activity (open circles). Gel insets show the urea-glycerol PAGE analysis of MLC\textsubscript{20} phosphorylation by an active 61-kD (a) and inactive 61-kD (b) preparation of the catalytic fragment of MLCK, at 0, 1, 5, and 10 min after the addition of ATP. LC\textsubscript{20}D, dephosphorylated MLC\textsubscript{20}; LC\textsubscript{20}P, phosphorylated MLC\textsubscript{20}; LC\textsubscript{17}, 17-kD myosin light chain. The band migrating slightly faster than LC\textsubscript{17} is most likely a proteolytic fragment of myosin.

Table I. Transit Times for Mitosis and Cytokinesis in Control and Microinjected Cells

<table>
<thead>
<tr>
<th>Condition</th>
<th>NEB to Anaphase onset (min)</th>
<th>Anaphase onset to midcytokinesis</th>
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<tbody>
<tr>
<td>Uninjected NRK cells</td>
<td>26.1 ± 3.9 (n = 19)</td>
<td>4.8 ± 0.8 (n = 15)</td>
</tr>
<tr>
<td>Active 61-kD fragment</td>
<td>46.6 ± 17.1 (n = 39)</td>
<td>4.9 ± 1.1 (n = 33)</td>
</tr>
<tr>
<td>Buffer-injected controls</td>
<td>28.1 ± 4.0 (n = 12)</td>
<td>4.6 ± 0.8 (n = 12)</td>
</tr>
<tr>
<td>Inactive 61-kD fragment</td>
<td>24.2 ± 4.1 (n = 7)</td>
<td>4.8 ± 0.9 (n = 28)</td>
</tr>
<tr>
<td>MLCK (Ca\textsuperscript{2+}/CaM-dependent)</td>
<td>23.4 ± 4.3 (n = 13)</td>
<td>4.2 ± 1.3 (n = 10)</td>
</tr>
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</table>
Figure 4. Time-lapse series of phase micrographs of a buffer-injected NRK cell during mitosis. After NEB (a, t = 0 min), the cell was microinjected at t = 5 min and observed through cell division. Images were recorded during prometaphase (b, t = 9.7 min), metaphase (c, t = 19.4 min), anaphase onset (d, t = 28.3 min), anaphase II (e, t = 29.8 min), and midcytokinesis (f, t = 32.2 min). Note that the plasma membrane appears smooth around the cell, except for slender projections anchoring the cell to the substrate (arrows). Bar, 10 μm.

Microinjection of the 61-kD Catalytic Fragment of MLCK into Dividing Cells

To define the timing of different stages of mitosis in NRK cells, we observed normal dividing cells by video microscopy and measured the period from NEB to midcytokinesis under conditions where the culture temperature was precisely maintained (see Materials and Methods). Uninjected cells showed a transit time from NEB to midcytokinesis averaging ~31 min, with the period from NEB to anaphase onset lasting ~26 min, and the period from anaphase onset to midcytokinesis lasting ~5 min (Table 1).

In contrast, cells injected with the active 61-kD fragment showed a significant delay in the progression through prometaphase and metaphase, with transit times between NEB and anaphase onset ranging from 25 to 97 min (Fig. 3; Table 1). In addition, ~10% of the injected cells failed to show normal karyokinesis or cytokinesis (see below). Surprisingly, all of the cells that completed karyokinesis showed no effect on the duration of anaphase or the rate of cytokinesis, as measured from the point of anaphase onset to midcytokinesis (Table 1). Microinjection of cells with buffer alone, the inactive 61-kD fragment, or intact MLCK showed no effect on either the timing of mitosis or the rate of cytokinesis (Table 1). Cells injected with control solutions exhibited typical morphological features of uninjected cells as they progressed through mitosis and cytokinesis, including chromosome condensation and NEB, chromosome congregation and alignment, chromosome separation, and formation of the cleavage furrow (Fig. 4).

In contrast, a large proportion (75%) of cells injected with the active 61-kD fragment showed striking surface blebbing activities during later stages of mitosis. As shown in Fig. 5, the injected cell progressed through prometaphase with a normal phase morphology, however during anaphase highly active blebs showing rapid expansion and retraction activities were observed on the cell surface. The blebbing activity was initiated during metaphase in ~40% of injected cells, and during anaphase in ~35% of injected cells. Once started, the blebbing continued through the end of cell division. Blebs were initially observed over the entire surface during metaphase (Fig. 6), but as cells entered cytokinesis the blebbing activity subsided in the equatorial region where apparently normal cleavage furrows formed (Figs. 5 and 6).

In a small percentage of injected cells that failed to show normal karyokinesis (mentioned above), metaphase chromosomes formed dispersed aggregates, while erratic blebs and constrictions were observed on the cell surface (Fig. 7, a and b). The failure to undergo normal cytokinesis was likely related to the abnormal karyokinesis observed in these cells.

Fluorescence Localization of F-actin, Myosin, and Tubulin after Microinjection of the 61-kD Fragment

The distribution of F-actin and myosin in cells microinjected with the 61-kD fragment was observed after staining with fluorescent phalloidin or specific antibodies against myosin.
In cells exhibiting dynamic surface activity, blebs were delineated by intense cortical staining of actin (Figs. 8a and 9a). In some cases actin and myosin were clearly localized near the base of the blebs (Fig. 9a and b, arrows). In dividing cells both actin filaments and myosin were present in the cortex of the cleavage furrow, although the degree of staining varied from cell to cell as previously shown by others (Nunnally et al., 1980; Cao and Wang, 1990a). Similar distributions of actin and myosin in the furrow were observed for control cells and cells injected with the 61-kD fragment. Note that the variability of myosin staining in the furrow of NRK cells was observed both with the monoclonal and polyclonal antibodies used in this study, however only the mAb appeared to show a distinct punctate staining pattern as previously observed in lymphoblasts (Bornens et al., 1989) and PtK2 cells (Klotz et al., 1986).

To determine whether the delay of prometaphase and metaphase was related to disruptions of the mitotic spindle, injected cells were processed for immunofluorescence with anti-tubulin antibodies. Disrupted spindle structures, lacking well-defined kinetochore fibers, were observed in injected cells exhibiting prolonged periods of prometaphase.
and metaphase (Fig. 10, d and e). However, upon the onset of anaphase, the organization of spindle microtubules in 61-kD delayed cells (Fig. 10f) appeared similar to that of control cells (Fig. 10, g–i).

Effects of the 61-kD Fragment on Cortical Activities

One of the most dramatic effects of the microinjection of the 61-kD fragment is the induction of active surface blebbing during metaphase and anaphase (Figs. 5 and 6). Although the mechanism for blebbing is poorly understood, it is likely that the 61-kD fragment causes an increase in the phosphorylation of myosin and stimulates contractile activities, which in turn may generate surface blebs through deformation of the cortical cytoskeleton and extrusion of the inner cytoplasm. This explanation is supported by the concentration of actin filaments and myosin in the cortex and near the base of blebs (Figs. 8 and 9; also see Herman et al., 1981, and Zeligs and Wollman, 1977). In addition, a recent study has reported abnormal surface activities in Dictyostelium cells expressing a truncated myosin II, which lacks the ability to undergo disassembly and appears to show enhanced interactions with cortical actin (Egelhoff et al., 1991). Blebbing has also been observed in some transformed cell lines (Porter et al., 1973) and under conditions that presumably weaken the mechanical strength of the cortex, e.g., after the treatment of cells with cytochalasin B (Puck et al., 1972; Miranda et al., 1974; Perry and Snow, 1975; Godman and Miranda, 1978), or in melanoma cells deficient in the actin filament cross-linking protein filamin (Byers et al., 1991).

The apparent lack of blebbing during prometaphase in 61-kD-injected cells suggests that the activation of myosin-dependent motility may depend on multiple factors. For example, other kinases or phosphatases may affect the extent and/or sites of phosphorylation on MLC20 (Lamb et al., 1988; Fernandez et al., 1990). A particularly attractive possibility is that MLCK-dependent phosphorylation of MLC20 may be modulated by the activity of the p34^{cdk2} kinase (Sat-
terwhite et al., 1990), which phosphorylates MLCZo at serines 1, 2, and threonine 9, and inhibits MLCK-mediated phosphorylation of MLCZo at serine 19 (Ikebe et al., 1987b; Sellers and Adelstein, 1987; Pollard et al., 1990). Thus, even though the 61-kD fragment is likely to be active during prometaphase, phosphorylation of MLCZo at serine 19 may be inhibited until the activity of p34°&Z declines later in mitosis. Future studies to test the level and sites of MLCZo phosphorylation in dividing cells will be extremely valuable.

**Effects of the 61-kD Fragment on Cytokinesis**

Several mechanisms for the regulation of cytokinesis have been proposed (Rappaport, 1986; Salmon, 1989; Mabuchi, 1990; Pollard et al., 1990). One attractive hypothesis involves the preferential phosphorylation of myosin in the equatorial region by MLCK, presumably after the dephosphorylation of MLCZo at the p34°&Z directed sites and a localized activation of MLCK by the Ca^{2+}-calmodulin complexes (Pollard et al., 1990). The increase in the phosphorylation of MLCZo at serine 19 may be inhibited until the activity of p34°&Z declines later in mitosis.
lation of MLC\textsubscript{2o} at serine 19 then activates contraction of the cleavage furrow. Based on this model, one would predict that changes in the degree, timing, or location of myosin phosphorylation could disrupt the process of cytokinesis. Specifically, microinjection of the 61-kD fragment should cause an unregulated phosphorylation of myosin throughout the entire cortex of dividing cells, that in turn might stimulate a global contraction and potentially disrupt both the timing and pattern of cell cleavage.

To our surprise, cytokinesis proceeded normally in the majority of 61-kD injected cells with respect to both the site and rate of cell cleavage (Figs. 5 and 6; Table I). These results indicate that additional factors, independent of the phosphorylation of MLC\textsubscript{2o}, are likely involved in regulating cytokinesis. Based on previous studies, it is clear that cytokinesis involves not only acto-myosin interactions, but also formation of contractile structures in the cleavage furrow through the recruitment and organization of actin filaments and accessory proteins (Fujitwara et al., 1978; Aubin et al., 1979; Nunnally et al., 1980; Sanger et al., 1989; Cao and Wang, 1990a,b; Mabuchi, 1990; Pollard et al., 1990). Thus, the assembly of cross-linking proteins in the equatorial region may account for the suppression in blebbing activity concomitant with the organization of actin–myosin interactions into an effective cleavage force. In contrast, the lack of such organization in other regions may account for the sustained blebbing activities observed outside the furrow.

The regulation of cytokinesis may also involve the concerted activities of multiple kinases and phosphatases. While MLC\textsubscript{2o} may be continually phosphorylated by the 61-kD catalytic fragment in injected cells, it is possible that only the equatorial region is able to maintain a specific pattern and level of phosphorylation during cytokinesis. The activity and localization of myosin in dividing cells may also be regulated by additional factors, such as phosphorylation of the heavy chains (Egelhoff et al., 1991), and/or association with myosin-binding proteins (Yabkowitz and Burgess, 1987). However, the physiological role of these factors in mammalian tissue culture cells remains to be elucidated.

### Effects of the 61-kD Fragment on Mitosis

One of the unexpected results of this study is that injection of the 61-kD catalytic fragment affects not only cortical activities, but also the progression of prometaphase and metaphase. Given the variety of evidence indicating that myosin II heavy chains are not directly involved in chromosome congression or segregation (Mabuchi and Okuno, 1977; Kiehart et al., 1982; De Lozanne and Spudich, 1987; Knecht and Loomis, 1987; Zurek et al., 1990), we are left with the question of how an increase in MLCK activity and elevation of phosphorylated MLC\textsubscript{2o} might cause delays in mitosis.

The most direct explanation for such delays is that the dephosphorylation of MLC\textsubscript{2o} at serine 19 is somehow required for the progression from metaphase to anaphase. This implies that MLC\textsubscript{2o}, in addition to regulating myosin heavy chain activities, may also interact with other molecules involved in mitosis. One possibility is that, since MLC\textsubscript{2o} can serve as a substrate for several kinases as well as phosphatases (Sellers and Adelstein), it is possible that increases in the phosphorylation state of MLC\textsubscript{2o} may alter its interaction with these enzymes, resulting in a change in the balance of activities among different kinases and phosphatases involved in the regulation of mitosis. Alternatively, the catalytic domain of MLCK is known to share some homology with other members of the Ca\textsuperscript{2+}-calmodulin-dependent kinase family (Hanks et al., 1988; Olson et al., 1990; Shoemaker et al., 1990). Thus, the removal of certain noncatalytic domains during the preparation of 61-kD could affect its subcellular localization and/or substrate specificity, and cause delays in mitosis by phosphorylating proteins involved in spindle function. Finally, immunofluorescence studies suggest that MLCK may undergo redistribution from stress fibers in interphase cells (DeLanerolle et al., 1981) to kinetochore spindle-fibers in mitotic cells (Guerrero et al., 1981). This raises the question of whether MLCK may normally have additional substrates in the mitotic spindle, in addition to MLC\textsubscript{2o}. Future experiments to test these ideas should yield new and important insights regarding the mechanism of mitosis and cytokinesis.

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