Polarized and Functional Epithelia Can Form after the Targeted Inactivation of Both Mouse Keratin 8 Alleles

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Abstract. We have tested the requirement of keratin intermediate filaments for the formation and function of a simple epithelium. We disrupted both alleles of the mouse keratin 8 (mK8) gene in embryonic stem cells, and subsequently analyzed the phenotype in developing embryoid bodies in suspension culture. After the inactivation of the mouse keratin 8 (mK8) gene by a targeted insertion, mK8 protein synthesis was undetectable. In the absence of mK8 its complementary partners mK18 and mK19 were unable to form filaments within differentiated cells. Surprisingly, these ES cells differentiate to both simple and cystic embryonic bodies with apparently normal epithelia. Ultrastructural analysis shows an apparently normal epithelium with microvilli on the apical membrane, tight junctions and desmosomes on the lateral membrane, and an underlying basal membrane. No significant differences in the synthesis or secretion of α-fetoprotein and laminin were observed between the mK8- or wild-type embryoid bodies. Our data show that mK8 is not required for simple epithelium formation of extraembryonic endoderm.

Intermediate filament (IF) genes constitute a superfamily whose members are expressed in very specific spatio-temporal patterns during development. Keratin IFs are found in all mammalian epithelia and are obligate heteropolymers composed of two groups of IF proteins, the type I and II keratins (for review see Steinert and Roop, 1988). Subsetsof 2 to 10 individual keratins of the ~20 members are expressed in pairs in diverse epithelia. During mouse embryogenesis, mK8 and mK18 (the mouse homologue of the human keratin 8 and 18, also named Endo A and B) are the first intermediate filament proteins to be expressed (Jackson et al., 1980; Kemler et al., 1981; Oshima et al., 1983). They are detected just before the morula stage. Subsequently, they become restricted to the simple epithelium of the early embryo, including the trophoderm, the parietal and visceral endoderm, as well as the single-layered epithelia of the adult animal, such as gut and liver (Moll et al., 1982).

Keratins are commonly regarded as structural proteins. However, they differ from other cytoskeletal components by their complex and specific pattern of expression. This diversity is thought to be functionally significant. Despite several attempts to interfere with keratin filament formation, their function remains obscure. Recently, it has been reported that dominant negative mutations of keratin proteins prevented extraembryonic epithelium formation (Trevor, 1990) or resulted in a defective skin epithelium (Vassar et al., 1991).

1. Abbreviations used in this paper: EB, embryoid body; ES, embryonic stem; HAS, hypoxanthine and azaserine; hrpt, hypoxanthine guanine phosphoribosyl transferase; h-LIF, human leukemia inhibiting factor; IF, intermediate filament; mK8, mouse keratin 8; PCR, polymerase chain reaction. These studies supported the generally accepted view that keratins were involved as a structural support during epithelium formation.

An important approach to studying gene function in vivo is the analysis of cells or organisms homozygous for loss-of-function mutations. Embryonic stem (ES) cells have been used to introduce targeted mutations into the germ line of mice (Thompson et al., 1989; Koller et al., 1989; Schwartzberg et al., 1989; Zijlstra et al., 1989; for review see Rossant and Joyner, 1989; Capecci, 1989). In addition to their use for functional studies in vivo, ES cells provide a powerful in vitro model of embryonic development (Martin, 1981; Evans and Kaufmann, 1981; Doetschman et al., 1985; Nagy et al., 1990; for review, Baribault and Kemler, 1990). In suspension culture, ES cells differentiate to organized structures known as embryoid bodies (EBs) containing a outer layer of endoderm and an inner ectodermal layer, separated by a basal lamina. These EBs expand into large cystic structures reminiscent of the visceral yolk sac both morphologically and biochemically. The mK8 type II keratin, with its partners mK18 and mK19 (Endo C) type I keratins, are induced in extraembryonic endoderm in vivo, as well as in vitro after the differentiation of ES cell and embryonal carcinoma cells to EBs (Boller and Kemler, 1983; Oshima, 1981, 1982).

To analyze the function of mK8 in early embryonic development, and to test a possible involvement of mK8 in epithelium formation and organization, we inactivated both mK8 alleles in ES cells, and thereby impaired filament formation for all three keratins. We analyzed the effect of the targeted mutation in developing EBs, and found that mK8 is dispensable for the formation of extra embryonic endodermal epithelium.
Materials and Methods

Cell Culture

EI4TG2a ES cell line was originally established by Hooper et al. (1987). ES cells were routinely cultured in DME supplemented with 15% FCS, 0.1 mM β-mercaptoethanol and 1,000 U h-LIF on gelatin-coated plates. LIF was produced by transfecting the pC10-6R plasmid (kindly provided by J. Heath) (Smith et al., 1988; Moreau et al., 1988; Williams et al., 1988) into cos cells and the supernatant was subsequently titrated using ES cell colony plating efficiency. For embryoid body formation, ES cells were plated at a density of 10^5 cells/100-mm bacterial petri dish (Fisher) in DME supplemented with 10% PBS and 0.1 mM β-mercaptoethanol. Culture medium was changed every day.

Gene Targeting

The K8TV1 targeting vector contains two arms of homology to the mK8 gene (Endo Aa4), a 1.6-kb Xba1 fragment and a 1.2 Smal fragment, respectively. The neo gene (pMcInlexpO4A, Stratagene, La Jolla, CA) replaces part of the first exon, including the ATG translation initiation codon (Sémat et al., 1988). The modified Herpes simplex virus thymidine kinase gene (Mansour et al., 1988), HSV- tk, was added 3', followed by the Bluescript vector (Stratagene). K8TV2 and K8TV3 are essentially identical to K8TV1 except that the neo gene has been replaced by the pBl(23) and pBl(23) hprt mini-genes (Reid et al., 1990). For electroporation, ES cells were transfected with 40 μg of Not I linearized targeting vector in 0.7 ml of culture medium using a BTX Transfecter 100 at 250 V for 12 h after electroporation, one plate was trypsinized and the number of cells counted. This number was used to calculate cell survival. 150 mg/ml G418 (for K8TV1) or G418 and hypoxanthine (0.7 μg/ml)-azaserine (10 μM) (HAS) (for K8TV2 and K8TV3) as well as 2 μg Ganc were added 24 h after electroporation. One plate was selected in the absence of Ganc to evaluate the transformation efficiency and the enrichment factor of the Ganc selection. The number of colonies without Ganc selection are deduced relative to this number. After 10 d under selection, single colonies were picked and grown in duplicate. PCR analysis was performed using 27-29-mer oligonucleotides, one complementary to the neo R gene (for K8TV1, 5'-GCC ACG GGTGTT GGG GC-3') and onelocated intronone, respectively. After transfection of E14K8-Nl cell clone 1 was used for the inactivation of both alleles are depicted in Fig. 1. They contain 3 kb of the mK8 gene, interrupted by the neo gene for the targeting of the first allele (K8TV1) and by a hprt minigene for the targeting of the second allele (K8TV2). The Herpes simplex virus thymidine kinase, HSV- tk, was inserted to make use of the positive-negative selection with gancyclovir (Ganc) (Mansour et al., 1988). EI4TG2a ES cells were transfected with K8TV1 and four targeting events were identified by PCR screening (Table 1). Three of those clones were expanded and the planned alteration of one mK8 allele was confirmed by Southern blot analysis (Fig. 2).

Immunobiochemical Analysis

For immunofluorescence analysis, ES cells were sedimented at 1 g, mixed with OCT compound, and frozen directly on dry ice. 5-μm sections were fixed in 100% methanol for 10 min, rinsed with PBS, and then incubated with the respective antibodies. For staining in monolayer, ES cells were trypsinized, replated on gelatin-coated coverslips, and methanol-fixed 2 d later. TROMA-1, -2, and -3 rat mAbs are directed against mK8, mK18, and mK19, respectively (Boiler and Kemler, 1987). Three hybridoma cell lines were a gift of Dr. R. Kemler (Max-Planck Institut für Immunobiologie, Freiburg, Germany). mK8 and mK18 antisera have already been described previously (Oshima, 1982). Antivimentin (Graver et al., 1983) and antidesmoplakinI/II (Pasdar and Nelson, 1991) antisera were gifts from Drs. J. Singer (University of California at San Diego, San Diego, CA) and J. Nelson (Stanford University, Stanford, CA), respectively. Laminin antisera was described previously (Oshima and Linney, 1988). Extensive immunoprecipitations and E LISA analysis confirmed that the sample was specific to the laminin A and B chains. Anti-α- fetoprotein was commercially available from Miles Laboratories Inc. (Elkhart, IN). Fluorescein-conjugated rabbit anti-rat IgG, goat anti-rabbit IgG, and rhodamine-conjugated goat anti-rabbit IgG antibodies were purchased from Oracon Teknika (West Chester, PA). For double staining experiments, the species specificity of the second antibody was controlled as follows: cells were incubated with TROMA-2 rat mAb and subsequently with rhodamine anti-rabbit antisera. Also, cells were incubated with desmoplakin I/II rabbit antisera and subsequently with fluorescein anti-rat IgG. Both controls were negative, showing that the second antibodies used here were species specific.

For most immunoprecipitation experiments, EB cultures were labeled overnight with 35S-Met at a concentration of 50 μCi/ml, in methionine-free EB culture medium. Secretion was analyzed by pulse-chase experiments where cells were labeled with 200 μCi/ml of 35S-Met for 1 h and rinsed 3× with normal complete EB culture medium. Both supernatant and EBs were collected at different chase times. Cell lysate, containing 10^7 TCA-insoluble cpm, were incubated for 2-3 h with the respective antibodies, followed by a 50-min incubation with Staphylococcus aureus as described previously (Oshima, 1982). The pelleted proteins were separated on a 12.5% SDS-PAGE and detected by fluorography (Bonner and Laskey, 1974). Gels were exposed to XAR-5 Kodak film. For quantitative analysis, films were scanned with an LKB laser densitometer.

Electron Microscopy

Epon-embedded EBs were sectioned and stained with uranyl acetate and lead citrate, following standard procedures for EM.

Results

Inactivation of mK8 by Gene Targeting

To impair both mK8/mK18 and mK8/mK19 keratin filament formation, we inactivated the type II mK8 keratin. In the mouse genome, there is one functional mK8 gene with eight exons and a single pseudogene (Vasseur et al., 1983). The targeting vectors used for the inactivation of both alleles are depicted in Fig. 1. They contain 3 kb of the mK8 gene, interrupted by the neo gene for the targeting of the first allele (K8TV1) and by a hprt minigene for the targeting of the second allele (K8TV2). The Herpes simplex virus thymidine kinase, HSV- tk, gene was inserted to make use of the positive-negative selection with gancyclovir (Ganc) (Mansour et al., 1988). EI4TG2a ES cells were transfected with K8TV1 and four targeting events were identified by PCR screening (Table 1). Three of those clones were expanded and the planned alteration of one mK8 allele was confirmed by Southern blot analysis (Fig. 2).

The 41K8-N1 cell clone was used for the inactivation of the remaining allele. The parental EI4TG2a ES cell line carries a deletion in the hprt gene and therefore lacks hprt activity completely (Hooper et al., 1987). Two forms of a hprt minigene (Reid et al., 1990) were tested as selective markers in the construction of the targeting vectors for the second allele. K8TV2 (Fig. 1) and K8TV3 (not shown) contain the hprt minigene including the first and second introns or the second intron alone, respectively. After transfection of 41K8-N1 cells with K8TV2 and K8TV3, hprt' clones were selected in medium containing hypoxanthine and azaserine (HAS) in order to inhibit de novo synthesis of purines and make the cells dependent upon the activity of the hprt gene for growth. Azaserine replaced the more conventional aminoopterin and thymidine combination, found in HAT medium, to avoid possible interference with the negative selection using the nucleotide analogue Ganc. G418 selection was continued during the course of the second homologous recombination experiment in order to select only recombination events with the unmodified allele. Six targeting events were identified by...
should inactivate it. This allele becomes mK8- and hprt+. Southern blot analysis should reveal a shift in size of the XbaI restriction fragment from 8.5 to 9.6 kb. Small black boxes represent the oligonucleotides used to identify the targeting events by PCR (see Table 1).

PCR from 153 screened colonies after transfection with K8TV2, but none with K8TV3 (Table I). The targeted modifications were confirmed by Southern blot analysis for all expanded clones, which were positive by PCR (Fig. 2). One HASr clone, which did not contain a second targeted gene, was also picked as a control for further functional assays (Fig. 2, lane 9).

The targeting frequency with K8TV2 (1/25 HASr + GanC colonies or 4.3 x 10⁻⁸ transfected cells) is approximately half of that observed with K8TV1 (1/11 HASr + GanC colonies or 10⁻⁷ transfected cells). Taking into account that only half the number of targets were present after the first inactivation, these results are in agreement with previous observations that the targeting frequency is independent of the length of nonhomologous DNA (Mansouret al., 1990). The unique difference between K8TV2 and K8TV3 is the presence of the hprt first intron in the former construct. We were unable to detect any targeting events with K8TV3, although HASr colonies were obtained. The first intron appears to contain regulatory elements necessary for sufficient levels of hprt expression in the context of the nonexpressed mK8 gene (Reid et al., 1990).

Table I. Targeting Frequency

<table>
<thead>
<tr>
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<th>K8TV1</th>
<th>K8TV2</th>
<th>K8TV3</th>
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<tr>
<td>Transfected cells</td>
<td>10⁵</td>
<td>1.4 x 10⁸</td>
<td>10⁵</td>
</tr>
<tr>
<td>Surviving cells</td>
<td>5 x 10⁷</td>
<td>7 x 10⁷</td>
<td>5 x 10⁷</td>
</tr>
<tr>
<td>Colonies without GanC</td>
<td>6,400</td>
<td>1,526</td>
<td>300</td>
</tr>
<tr>
<td>Transformation efficiency</td>
<td>6.4 x 10⁻⁵</td>
<td>0.9 x 10⁻⁵</td>
<td>0.3 x 10⁻⁵</td>
</tr>
<tr>
<td>Colonies with GanC</td>
<td>320</td>
<td>218</td>
<td>30</td>
</tr>
<tr>
<td>GanC enrichment factor</td>
<td>20-fold</td>
<td>7-fold</td>
<td>10-fold</td>
</tr>
<tr>
<td>Screened colonies</td>
<td>42</td>
<td>153</td>
<td>28</td>
</tr>
<tr>
<td>PCR positives</td>
<td>4</td>
<td>6</td>
<td>0</td>
</tr>
<tr>
<td>Targeted clones (confirmed by Southern blot)</td>
<td>3/3</td>
<td>5/5</td>
<td>–</td>
</tr>
<tr>
<td>Targeting frequency per:</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>GanC/C/G418/HASr colonies</td>
<td>1/11</td>
<td>1/25</td>
<td>–</td>
</tr>
<tr>
<td>transfected cells</td>
<td>10⁻⁷</td>
<td>0.43 x 10⁻⁷</td>
<td>&lt;10⁻⁴</td>
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K8TV1 targeting vector (Fig. 1) was used for the disruption of the first allele, K8TV2 (Fig. 1) and K8TV3 (not shown) for the disruption of the second allele. The number of surviving cells is the number of attached cells 12 h after electroporation. The number of colonies without GanC is the number of colonies if they would have been selected in G418 (for K8TV1) or HAS (for K8TV2 and K8TV3) only. The number of colonies with GanC is the total number of G418 + GanC colonies (for K8TV1) or G418 + HAS + GanC colonies (for K8TV2 and K8TV3) obtained.
Figure 2. Southern blot analysis of the targeted ES cell clones. 5 μg of genomic DNA, digested with XbaI was loaded in each lane. The filter was hybridized with the 2.5-kb EcoRI pseudogene fragment (Endo Aα2 [Vasseur et al., 1985]), which is homologous to the exonic sequences of the mK8 gene. In the parental cell line (lane 2), the 8.5 and the 5-kb bands represent the gene and the pseudogene, respectively. The presence of a 9.5-kb band in lane 3 shows the expected shift in one allele of the mK8 gene after homologous recombination with K8TV1. The presence of the 10.5-kb band in lanes 4-8 shows that the remaining wild-type allele has been targeted with K8TV2. The control clone E14K8-N1-H6 (lane 9) is HASr, but, as expected from the PCR analysis, the second allele was not targeted. Radiolabeled lambda HindIII size markers are shown in lane 1.

Polarized Epithelium Forms in mK8- Embryoid Bodies

ES cells, homozygous or heterozygous for the targeted mutation, as well as the parental cell line were allowed to differentiate in suspension culture. All clones formed normal aggregates. Endoderm layers covering the surface of EBs were morphologically recognizable by light microscopy after 2–4 d (Fig. 3 A) and in histological sections (Fig. 3, B and C). Ectoderm-like epithelia formed beneath the outer endodermal layer as shown in sections of 4-d-old EBs culture (Fig. 3 B). Large cystic structures expanded after 8–10 d (Fig. 3 D). Neither the kinetics of embryoid body formation nor the proportion of aggregates that formed cystic structures appeared to be different from the wild-type in all mutant clones analyzed.

Transmission EM revealed apparently typical and normal epithelium in mK8- and wt EBs (compare Fig. 4, A and B). Cells were polarized with microvilli on the apical membrane and a basement membrane underlying the epithelial cell layer. Secretion granules were observed, indicating that these cells were actively secreting. Because keratins are associated with desmosomes we were particularly interested in whether these structures are present in mK8- EBs. Structures resembling desmosomes and tight junctions were observed on the lateral membrane of epithelial cells derived from both mK8- and wild-type ES cells (Fig. 4, C and D and inset to Fig. 4 D).

Lack of mK8/mK18 Filaments in Doubly Targeted Clones

To confirm that the introduced mutations in the mK8 gene functionally inactivated the mK8 gene, we let the parental and doubly targeted ES cells differentiate to EBs and performed immunoprecipitation and immunofluorescence experiments. The mK8 was immunoprecipitated with an mK8-specific antiserum from the parental cell line (Fig. 5 A, lane
Figure 4. Ultrastructural analysis of mK8− and wild-type endoderm. 4-d-old EBs derived from parental (A and C) and doubly targeted (B and D) ES cells were embedded in Epon. Transmission EM of thin sections show that mK8− and wild-type EBs form a polarized epithelium with microvilli (mv), basal membrane (bm), secretion granules (g), tight junctions (tj), and desmosomes (d). Magnifications: (A and B) 6,000×; C is a higher magnification of A at 15,000×; (D) 18,000×; inset in D shows desmosome-like structure in mK8−EB at higher magnification.
Figure 5. Immunoprecipitation of mK8, mK18, laminin, and α1-fetoprotein in wild-type and doubly targeted ES cells. (A) 7-d-old EBs derived from doubly targeted (lanes 2, 4, 6) or parental (lanes 1, 3, 5) ES cells were labeled with 50 μCi/ml [35S]Met overnight. 107 cpm of lysate were incubated with mK8 (lanes 1 and 2), mK18 (lanes 3 and 4), or laminin (lanes 5 and 6) antisera. The immunoprecipitated proteins were separated on a 12.5% SDS-PAGE and exposed to film overnight. Arrows indicate the position of mK8, mK18, and laminin A and B chains. In B, 12-d-old EBs derived from doubly targeted (lanes 3-4, 7-8, 11-12, 15-16) and parental (lanes 1-2, 5-6, 9-10, 13-14) ES cells were labeled with 200 μCi/ml for 30-min pulse. Radioactive medium was replaced with a normal medium and EBs were harvested after 0 h (lanes 1-4), 1.5 h (lanes 5-8), 2.5 h (lanes 9-12), or 5 h (lanes 13-16). α1-Fetoprotein was immunoprecipitated from the cell lysates (lanes 1, 3, 5, 7, 9, 11, 13, 15) or from the culture supernatants (lanes 2, 4, 6, 8, 10, 12, 14, 16). Immunoprecipitated proteins were separated on a 12.5% SDS-PAGE and exposed to film overnight. Fluorograms were scanned and the percentage of secreted α1-fetoprotein (α1-fetoprotein in supernatant/total α1-fetoprotein) was plotted as a function of time. P, cellular pellet; S, culture supernatant.

1.5 h (lanes 5-8), 2.5 h (lanes 9-12), or 5 h (lanes 13-16). α1-Fetoprotein was immunoprecipitated from the cell lysates (lanes 1, 3, 5, 7, 9, 11, 13, 15) or from the culture supernatants (lanes 2, 4, 6, 8, 10, 12, 14, 16). Immunoprecipitated proteins were separated on a 12.5% SDS-PAGE and exposed to film overnight. Fluorograms were scanned and the percentage of secreted α1-fetoprotein (α1-fetoprotein in supernatant/total α1-fetoprotein) was plotted as a function of time. P, cellular pellet; S, culture supernatant.

I), but was not detected in EBs derived from the doubly targeted ES cells (Fig. 5 A, lane 2). Immunofluorescence staining was performed on EB frozen sections. mK8 was uniformly present throughout the differentiated endoderm of wild-type EBs (Fig. 6 A), but was absent from doubly targeted EBs (Fig. 6 B). In addition, in the mK8- EBs, no mK18 staining was observed (Fig. 6, C and D), even though mK18 protein synthesis was detected by immunoprecipitation (Fig. 5 A, lanes 3 and 4). These results confirm that both mK8 and mK18 are required for keratin filament formation and are in agreement with previous observations that human K18, expressed in the absence of a complementary keratin partner, is rapidly degraded (Kulesh et al., 1989). mK19 is expressed in extraembryonic endoderm and it has been suggested to be a partner keratin for mK8. mK19 was found to be present in the endoderm layer of the cystic structure in wild-type EBs (Fig. 6 E). In the absence of mK8, mK19 was unable to form filaments (Fig. 6 F), which strongly supports the notion that mK8 is the partner of mK19 in extraembryonic endoderm.

To better visualize the staining pattern of these antigens in differentiated ES cells, 10-d-old EBs were replated on coverslips. Endodermal cells as well as other cell types migrated out of the EBs. Immunofluorescence staining for mK8, mK18, and mK19 on EB outgrowths confirmed the results obtained with EB sections (Fig. 7), except for a small subpopulation of cells (from a few to about 10% of differentiated cells) which showed mK18 and mK19 staining at the periphery of mK8- cells. The nature of these cells is unclear, but the punctate staining pattern was reminiscent of desmosome staining by desmoplakin antibodies. To test the hypothesis that the structures associated with mK18 and mK19 were desmosomes, we performed double staining of mK18 (Fig. 8 B) or mK19 (not shown) and desmoplakin I/II (Fig. 8 A). The staining patterns were identical, suggesting that in the absence of mK8, mK18 and mK19 have the ability to associate with desmosomes. The localization of both antigens was not observed in EB sections. Therefore, it is unlikely that this cell population represents endodermal cells. In addition, this phenomenon is not observed when ES cells are induced to differentiate to a homogenous endodermal cell population by retinoic acid (data not shown). Desmoplakin I/II is observed in most extraembryonic endoderm of both mK8- and wild-type EBs (Fig. 6, K and L).

The absence of normal filament staining patterns for mK18 and mK19 indicates that it is unlikely that another type II keratin is expressed in the differentiated cells of mK8- EBs because any other type II keratin would be expected to polymerize with mK18 and mK19 resulting in detectable filamentous structures (Hatzfeld and Franke, 1985). However, vimentin is expressed in most cultured cells including undifferentiated ES cells (Paulin et al., 1982; personal observation) even though it is absent from the inner cell mass from which ES cells are derived (Jackson et al., 1980; Oshima et al., 1983). While vimentin and keratins form distinct IF networks, they are similar in primary structure (Singer et al., 1986; Steinert and Roop, 1988). Thus, vimentin could conceivably complement the absence of keratin filaments. How-
ever, the endodermal layer of mK8- EBs was negative for vimentin staining with a polyclonal antiserum (Fig. 6, I and J).

Functional Activity of the Visceral Endoderm

The large cystic structures of EBs are reminiscent of the visceral yolk sac, both morphologically and biochemically. Visceral endoderm of the yolk sac actively produces and secretes $\alpha_1$-fetoprotein and laminin in vivo. In EBs derived from embryonal carcinoma cells, the production and secretion of $\alpha_1$-fetoprotein appears to be a good indicator of the organization and maturity of the visceral endodermal epithelium (Grover et al., 1983). To test the functional activity of the $mK8^+$ extraembryonic endoderm we monitored $\alpha_1$-fetoprotein synthesis and secretion in EBs culture. No differences in the rate of synthesis or secretion were detectable by pulse-chase immunoprecipitation experiments (Fig. 5 B). Similarly, laminin synthesis (Fig. 5 A, lanes 5 and 6) and secretion (not shown) were not significantly different between the $mK8^+$ and wild-type EBs. Immunofluorescence

Figure 6. Immunofluorescence staining of doubly targeted and wild-type EBs. Frozen sections of EBs derived from the parental (A, C, E, G, I, K) and from doubly targeted (B, D, F, H, J, L) ES cells were stained by immunofluorescence with TROMA-1 (A and B), TROMA-2 (C and D), TROMA-3 (E and F) mAbs directed against the indicated antigens, antilaminin (G and H), antivimentin (I and J), and antidesmoplakin I/II (K and L) antisera. (Magnification 250×.) The white arrows indicate endoderm layers.

Figure 7. Immunofluorescence staining of mK8, mK18, mK19 in differentiated ES cells. 10-d-old EBs derived from parental (A, C, and E) and doubly targeted (B, D, and F) ES cells were trypsinized briefly and replated on gelatin-coated coverslips. After 3 d, cells were stained by immunofluorescence with TROMA-1 (A and B), TROMA-2 (C and D), and TROMA-3 (E and F) mAbs. (Magnification 400×.)
Figure 8. Double immunofluorescence staining of desmoplakin I/II and mK18 in doubly targeted differentiated ES cells. 10-d-old EBs derived from doubly targeted ES cells were trypsinized briefly and replated on gelatin-coated coverslips as in Fig. 5. Double immunofluorescence staining with antidesmoplakin I/II antisera (A) and TROMA-2 mAbs (B) show that both antigens colocalized at the periphery of the cells, in a subpopulation of epithelial cells.

staining revealed that laminin was deposited beneath the endoderm layer in mK8 E8-EBs in the same manner as in wild-type EBs (Fig. 6, G and H). These results demonstrate that an epithelium similar to the functional visceral endoderm can form in the absence of mK8.

Discussion

We show here that a polarized and functional epithelium can form in the absence of the mouse keratin 8. These results are unexpected because keratins are commonly regarded as structural proteins that provide strength to epithelial layers. However, it is clear that the visceral endodermal layer of cystic embryoid bodies that lack keratin filaments are able to withstand the apparent hydrostatic pressure associated with their expansion. In addition, the normal synthesis and secretion of α-fetoprotein, a sensitive marker of mature visceral endoderm, indicates that neither the processes of differentiation nor that of active secretion requires simple epithelial keratins.

Is the Lack of mK8 Requirement a Consequence of Functional Redundancy among IF Genes?

As an increasing number of targeted mutations are analyzed in mice, it has become evident that the pattern of expression of the targeted gene is not necessarily a good indicator of the tissue or time during development when the gene product is essential. The en-2 targeted mutation has an effect restricted to neural cells expressing en-2 but lacking en-1 (Joyner et al., 1991). Despite constitutive expression of c-src, a targeted mutation of c-src results in a defect in bone formation only (Soriano et al., 1991). Both groups suggested that other members of the same multigene family could complement the targeted gene function when coexpressed. Functional redundancy between similar proteins has been shown to occur in the E(spl)-C (enhancer of split complex in Drosophila), where several closely related transcripts have the same expression patterns (for review see Campos-Ortega and Knust, 1990). However, it is very unlikely that the deficiency in keratin filaments is complemented by another member of the keratin gene family, because other type II keratins capable of polymerizing with the normal mK18 and mK19 are absent from the mK8 visceral endodermal layer. However, a low level or an altered polymerized form of vimentin might escape detection by immunofluorescent staining even by polyclonal antiserum and might complement the keratin deficiency. No other intermediate filament proteins are known to be expressed in this tissue.

mK8 Is Dispensable for Endoderm Formation

The results of this study appear to contrast with two recent studies of the effects of dominant-negative mutations of two different keratin proteins. The disruption of mK8/mK18 filaments by a dominant mutation of K18 impaired the formation of an extraembryonic epithelium in an EB assay similar to that used here (Trevor, 1990). However, null and dominant-negative mutations are distinct genetic modifications that can lead to distinct phenotypes. One explanation of the differences of dominant-negative keratin mutant phenotypes with the null mutant phenotype reported here is that the remaining presence of disrupted keratin filaments or bundles found in cells expressing dominant-negative mutations may interfere with components essential for epithelial assembly or integrity. Possible unexpected functions of mutant proteins and consideration of the level of expression necessary for a biological effect are complications of experiments utilizing dominant negative mutations or other similar disrupting agents. The future use of gene targeting to generate dominant negative mutations may minimize the potential complication of obtaining physiological concentrations of the mutant gene product. In a second investigation, the expression of a dominant-negative mutation of an epidermal specific keratin in transgenic mice resulted in the formation of a defective basal epithelial layer with subsequent pathology similar to a group of human genetic disorders known as epidermolysis bullosa simplex (Vassar et al., 1991). In this case, epidermal keratins appear to have an important structural function. However, epidermal and simple epithelial keratins may have
different functions. Alternatively, the differences in phenotype may be due to different requirements of the respective tissues.

The function for these simple epithelial keratins may be much more subtle than previously suggested or than revealed by the in vitro differentiation of ES cells. The analysis of germ line mutation of the keratin 8 gene will extend this analysis to all murine epithelia and provide the most rigorous test of the function of affected tissues. However, the tissue-specific patterns of keratin expression vary considerably among species. For example, the expression of keratin filaments in preimplantation mouse hamster embryos and oocytes appears to differ significantly (Planche et al., 1989). Myocardial and retinal pigment epithelial cells of different species differ with regard to keratin expression (Kuruc and Franke, 1988; Ovaribe et al., 1988; Jahn et al., 1987; Markl and Franke, 1988). These inconsistencies suggest that keratins are fortuitously expressed in some epithelial cells. In addition, disruption of keratin filaments by antibody injection had no effect on cellular morphology or other aspects of PTK epithelial cell behavior in monolayer culture (Klymkowsky et al., 1983). Similarly, mouse embryos develop normally to the blastocyst stage after microinjection of keratin antibodies which disrupted filament organization (Emerson, 1988). These studies are in agreement with our results that mK8/mK18 and mK8/mK19 filaments are dispensable for endoderm formation.

**Further Applications for Double Gene Targeting**

The total loss of expression of mK8 in ES cells permits extensive biochemical analysis because EBs can be cultured in large quantity. Keratin IF bundles radiate from the perinuclear region to the desmosomes (Jones and Goldman, 1985) and hemidesmosomes (Klatte et al., 1989) which contribute to cell adhesion. Desmoplakin, a major component of the desmosome, is associated with keratin filaments before desmosome formation in MDCK cells (Pasarad et al., 1991). However, we found desmosome-like structures in the absence of mK8 filaments. Future detailed analysis of desmoplakin localization in mK8- epithelial cells may reveal differences in desmosome assembly. In that respect, the colocalization of mK18 and mK19 with desmoplakin I/II in a subpopulation is a surprising observation. It seems likely that mK18 and mK19 are able to bind desmosome components without the presence of a partner keratin. A recent analysis of mK8, mK18, and mK19 domain function emphasized the necessity for the absence of the endogenous protein to perform functional and structural analysis (Xu and Lane, 1990). Using a variety of truncated keratin expression vectors ectopically expressed in fibroblasts, the authors analyzed specific domain functions in filament formation. The system developed in this study would be ideal to study the effects of these mutations in an endogenous epithelial cell where keratins could interact with desmosome components.

The method of targeting both alleles of a gene and analyzing the phenotype in vitro (Rieke et al., 1991; Mortensen et al., 1991) may be suitable for any gene that is not essential for ES cell growth and is expressed in differentiated ES cells such as the early embryonic development lineages, hematopoietic lineages (Schmitt et al., 1991; Wiles and Keller, 1991), and cardiac muscle cells (Robbins et al., 1990). In addition, in cases where targeted mutations are embryonic lethal, chimeras of doubly targeted ES cells with wild-type embryos will permit the analysis of the phenotype at later stages (Pevny et al., 1991).

hpr is a bidirectional selective marker (Reid et al., 1990; Valancius and Smithies, 1991). We selected for the presence of HPRT activity. Subsequent selection against HPRT with thioguanine would permit replacement of the modified gene with additional constructs carrying modification of protein domains or regulatory signals. This will permit unlimited modifications of one allele in a null genetic background.

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