Mutations in Human Dynamin Block an Intermediate Stage in Coated Vesicle Formation

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Abstract. The role of human dynamin in receptor-mediated endocytosis was investigated by transient expression of GTP-binding domain mutants in mammalian cells. Using assays which detect intermediates in coated vesicle formation, the dynamin mutants were found to block endocytosis at a stage after the initiation of coat assembly and preceding the sequestration of ligands into deeply invaginated coated pits. Membrane transport from the ER to the Golgi complex was unaffected indicating that dynamin mutants specifically block early events in endocytosis. These results demonstrate that mutations in the GTP-binding domain of dynamin block Tfre-endocytosis in mammalian cells and suggest that a functional dynamin GTPase is required for receptor-mediated endocytosis via clathrin-coated pits.

Receptor-mediated endocytosis occurs via the formation of clathrin-coated vesicles and involves coat assembly, receptor recruitment, coated pit invagination, and finally vesicle budding (Smythe and Warren, 1991; Robinson, 1992; Schmid, 1993). The major structural proteins of the coat, clathrin, and AP complexes have been well characterized (Pearse and Robinson, 1990; Keen, 1990; Brodsky, 1988). While proteins that regulate coated vesicle formation have not been identified (Schmid, 1992), a cell-free assay which reconstitutes receptor-mediated endocytosis has recently revealed a requirement for multiple GTP-binding proteins (Carter et al., 1993).

Dynamin, a GTP-binding protein first isolated from bovine brain tissue (Shpetner and Vallee, 1989; Obar et al., 1990; Scaife and Margolis, 1990; Nakata et al., 1991) is a strong candidate for a GTPase that acts early in endocytosis. Dynamin is a member of a subfamily of structurally related GTP-binding proteins which share considerable sequence homology (43–66%) in the amino-terminal GTPase domain, but more limited homology (<10–30%) in their COOH-terminal regions. Other members of this family (for review see Collins, 1991) include the mammalian Mx-1 related, interferon-inducible proteins which confer viral resistance (Arnheiter and Meier, 1990), MGM1 which plays a role in yeast mitochondrial DNA maintenance (Jones and Fangman, 1992) and VPSlp, a protein involved in vacuolar sorting in yeast (Rothman et al., 1990; Yeh et al., 1991).

Whether these proteins have related functions or simply share a common GTPase domain driving divergent functions remains to be determined.

Evidence for the role of dynamin in endocytosis came from the discovery that the Drosophila homologue of dynamin is encoded by the shibire locus (van der Bliek and Meyerowitz, 1991; Chen et al., 1991). Adult flies with a mutation that causes the shibire gene product to be temperature sensitive exhibit rapid and reversible paralysis at elevated temperatures (Grigliatti et al., 1973). Morphological analysis of nerve terminals from affected flies revealed a depletion of synaptic vesicles and an accumulation of membrane invaginations and coated pits at the cell surface (Kosaka and Ikeda, 1983).

Other shibire–derived tissue and cell lines are similarly affected, demonstrating that the defect is pleiotropic. Results from the fluid-phase uptake of morphological tracers also suggest a specific defect in endocytosis but not secretion (Kosaka and Ikeda, 1983; Tsuruhara et al., 1990). However, a direct role for dynamin in receptor-mediated endocytosis has not been demonstrated in either Drosophila or mammalian cells.

The phenotypic analysis of the shibire mutation and the identity of the shibire gene product as dynamin present a paradox. Dynamin has been characterized in vitro as a microtubule-stimulated GTPase and a putative microtubule-based mechanochemical motor protein (Shpetner and Vallee, 1989; 1992), although the latter activity has not been reproduced with highly purified protein (Maeda et al., 1992). Several studies have shown that polymerized microtubules are not required during the early stages of endocytosis (Oka...
that they contained only the intended mutations. Supercoiled plasmid DNA, to remove the existing AUG. Full-length human dynamin cDNA with the recloned into SacII-cut and dephosphorylated pTM1-wild-type construct. 
fused by PCR in a second step with the upstream and downstream primers, of oligonucleotides in which the desired mutation had been incorporated expression construct, pTM1 (kindly provided by Dr. B. Moss), was modified 
sequence analysis using oligonucleotides synthesized on a Cyclone Plus DNA 
NcoI site of the linker followed by selection of a clone with the proper orien-
nce of the cDNA was added to make the cDNA full-length. The Vaccinia ex-
ous exposure times were quantitated using a scanning densitometer (Model 
samples from the resuspended cells used in transport assays were lysed by the addition of 1% Triton X-100 in PBS, containing 5 mM MgCl2 and 1 mM AEBSF (Calbiochem, La Jolla, CA). The samples were pellets to clear the nuclei, precipitated with 10% TCA, washed with ice cold acetone, resuspended in Laemmli sample buffer, and then analyzed by SDS-PAGE. The gels were electroblotted onto nitrocellulose, blocked with 5% nonfat dry milk powder in TBS containing 0.1% Tween 20 incubated with a 1:1,500 dilution of the rabbit anti-dynamin antibody (748) followed by detection with an HRP-conjugated secondary antibody (Pierce, Rockford, IL), and 
dilution of the rabbit anti-dynamin antibody (748) followed by detection 
with an HRP-conjugated secondary antibody (Pierce, Rockford, IL), and 
and then infected with vaccinia T7 RNA polymerase recombinant virus (vTF7-3, Fuerst et al., 1986), at a multiplicity of 10 PFU/cell for 30 min with inter-
mittent rocking in a 37°C incubator. The inouculum was removed and the 
cells transfected with 7 µg pTM1 plasmid DNA containing dynamin con-
structs prepared as described above that had been previously mixed for 15 
min at room temperature in 1 ml of DMEM and 20 µl of transfection reagent (TransfectACE) prepared as described (Rose et al., 1991). 2 h after infec-
tion, rifampicin (100 µg/ml final concentration) was added to the cells and 
expression was allowed to proceed for the indicated times.

Anti-Dynamin Antibodies and Western Analysis

E. coli carrying the expression vector pET1ID containing a Drosophila dyn-
imin fragment (residues 359–615) produced an abundant 30-kD protein 
upon induction by IPTG. The protein was gel-purified, electroeluted, and 
used as antigen to generate polyclonal antibodies (748) from rabbits by stan-
dard procedures. Serum from this rabbit contained an antibody immuno-
reactive against the 30-kD protein and a 100-kD protein in HeLa cells and 
bovine brain extracts (data not shown). The antibody also recognized 
recombinant wild-type and mutant molecules. For Western blot analysis, 
samples from the resuspended cells used in transport assays were lysed by the addition of 1% Triton X-100 in PBS, containing 5 mM MgCl2 and 1 mM AEBSF (Calbiochem, La Jolla, CA). The samples were pellets to clear the nuclei, precipitated with 10% TCA, washed with ice cold acetone, resuspended in Laemmli sample buffer, and then analyzed by SDS-PAGE. 

Subcellular Fractionation

Cells from one 60-mm dish were harvested at room temperature in PBS 
containing 5 mM EDTA and washed at 4°C by centrifugation, and 
resuspended in 0.5 ml 0.25 M sucrose, 1 mM MgCl2, 2 mM EGTA, 25 
Mm Hepes, pH 7.4. Cells were then lysed by rapid freezing in liquid nitro-
gen, followed by slow thawing in a room temperature water bath. Soluble 
and particulate fractions were obtained after centrifugation for 30 min at 
50 K rpm (110 k g) in a TLAO100.2 Beckman Instrs. Inc., Fullerton, CA) 
rotor. Pellets were resuspended in 0.5 ml and equal amounts of pellet and 
supernatant fractions were solubilized in 0.5% TX-100 and precipitated 
with 10% TCA. SDS-PAGE and Western blot analysis were performed as 
described above.

Assay of Tfn Endocytosis

After the appropriate period of protein expression (usually 5 h), the cells

Materials and Methods

Materials

Anti-Tf receptor monoclonal antibody, D65 was a kind gift of I. S. Tow-
bridge and S. White (Salk Institute, San Diego, CA). Anti-clathrin mono-
clonal antibody, X22, and anti-a-adaptin antibody, AP6, were generous 
gifts of E M. Brodsky (University of California, San Francisco). Human 
diferric transferrin (Tfn) 1 was biotinylated as previously described (Smythe 
et al., 1992). All other reagents were obtained from Sigma Chem. Co. (St. 
Louis, MO).

Molecular Analysis and Preparation of Constructs

A human dynamin cDNA fragment had previously been isolated using Dro-
sophila cDNA as a heterologous probe (van der Bliek and Meyerowitz, 1991). This fragment was used to screen a cDNA library made from human 
temporal cortex (Stratagene, La Jolla, CA) after standard protocols (Sam-
brook et al., 1988). The newly isolated cDNAs were characterized by se-
quence analysis using oligonucleotides synthesized on a Cyclone Plus DNA 
synthesizer (Milligen/Biosearch) and Sequenase (Un. States Biochem. 
corp., Cleveland, OH). For expression purposes, a linker with the optimal 
Kozak-consensus and an NcoI site at the AUG initiation codon was inserted 
into pBlueScript (Stratagene). A 1,200-bp Ncol fragment with the 5'-part 
of human cDNA (starting at the AUG-start codon) was inserted into the 
Ncol site of the linker followed by selection of a clone with the proper orien-
tation for expression. A 2,550-bp BstXI-HindIH fragment with the Y-part 
of the cDNA was added to make the cDNA full-length. The Vaccinia ex-
pression construct, pTM1-Hwt was cloned into pBlueScript (Stratagene). 
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were washed 1 × in dPBS, harvested at room temperature in dPBS containing 5 mM EDTA, pelleted in a Beckman J2-B centrifuge at 1,000 rpm, and resuspended in dPBS containing 1 mM CaCl₂, 1 mM MgCl₂, 5 mM glucose, and 0.2% BSA. For endocytosis assays, aliquots of cells were incubated at 37°C for the indicated times in the presence of BSST (2 μg/ml final concentration). The cells were then returned to ice, pelleted, and the supernatant aspirated. Latency to MesNa and avidin were determined as previously described ( Smythe et al., 1992; Carter et al., 1993). Internalized Tfn was expressed as the percent of total surface-bound Tfn. The level of Tfn internalization was normalized to the amount of internalization for mock-infected cells to account for day to day variation; however, the rate and extent of internalization after 5 h of vaccinia infection in the presence of rifampicin was typically between 60-90% of that observed for control cells which had not been infected.

Assay of ER-to-Golgi Transport

HeLa cells were infected and transfected as described above using 5 μg of dynamin containing plasmid and 5 μg of a plasmid encoding Vescular Stomatitis virus G (VSV-G) protein (pAR-G, Tisdale et al., 1992). After the transfection period, the cells were pulse-labeled for 10 min in methionine-free media containing 20 μCi TRANS 35S-label, and then chased for 1 h in DMEM/10% FCS. The chase was terminated by the addition of lysis buffer (50 mM Tris, pH 8.0, 150 mM NaCl, 1% Triton X-100, 1 mM PMSF) and VSV-G protein was immunoprecipitated using a monoclonal antibody to VSV-G (clone 8(35). Immune precipitates were washed, boiled, and eluted VSV-G protein was digested with endoglycosidase H (endo H) (Boehringer Mannheim, Indianapolis, IN), and analyzed by SDS-PAGE and fluorography (Tisdale, 1992).

Indirect Immunofluorescence

Coverslips containing transfected cells were incubated with 4 μg/ml anti-Tf receptors and monoclonal antibody to dynamin and in the presence of 50 μg/ml of avidin in dPBS/BSA. Excess avidin was removed by washing and the cells were fixed for 20 min at room temperature with 2.5% formaldehyde. Internalized biotinylated-HTR-D65 was detected by permeabilizing the fixed cells with 0.05% saponin and probing with Texas red-conjugated avidin (Leinco Technologies, 2 μg/ml) in permeabilization buffer with 50 μg/ml avidin in dPBS/BSA. Excess antibody was removed by washing at 4°C and the cells were incubated with 4 μg/ml anti-Tf receptor monoclonal antibody (D65) for 30 min at 37°C in dPBS ** (with 1 mM CaCl₂, 1 mM MgCl₂) containing 5 mM glucose and 0.2% BSA. Excess antibody was removed by washing at 4°C and the cells were fixed for 20 min at room temperature with 2.5% formaldehyde. Internalized biotinylated-HTR-D65 was detected by permeabilizing the fixed cells with 0.05% saponin and probing with Texas red-conjugated avidin (Leinco Technologies, 2 μg/ml) in permeabilization buffer with 50 μg/ml avidin in dPBS/BSA. Excess avidin was removed by washing and the cells were fixed for 20 min at room temperature with 2.5% formaldehyde.

Results

Molecular Analysis of Human Dynamin

Human dynamin cDNA was cloned for use in the functional studies in HeLa cells described below, to avoid problems that might have arisen with a heterologous expression system. A human brain cDNA library was probed with radiolabeled Drosophila shibire cDNA, followed by reprobing with the human fragments that were initially obtained. A cDNA spanning the entire coding region of the human shibire homologue was constructed from two overlapping fragments, and then sequenced. The deduced protein is 864 amino acids of which 99% are identical to rat dynamin (Fig. 1). Both human and rat sequences are similar to the Drosophila shibire sequence (69%), and this similarity extends throughout the molecule, strongly suggesting that function is conserved. Similar to rat and Drosophila, human dynamin has an NH₂-terminal GTPase domain that is highly conserved in VPS1p, MX-proteins, and Mgm1p. The remainder of the molecule is divergent compared to these other family members and includes a proline-arginine-rich COOH-terminal domain specific to the dynamin homologues.

Analysis of a large collection of cDNAs revealed alternative splicing at two positions in the coding region (shown in Fig. 1). This has not been reported for rat dynamin, but is comparable to the alternative splicing found in Drosophila (van der Bliek and Meyerowitz, 1991; Chen et al., 1991). Each human variant cDNA was found in multiple copies and the points of divergence correspond to exon-intron boundaries that include splice-site consensus sequences in the corresponding genomic DNA (data not shown). Thus it is unlikely that these represent cloning artifacts or incomplete processing. The first variation is the replacement of a 139-nt segment in the middle exon by an alternative exon of equal length. These two exons were equally represented in the cloned cDNA collection. Both exons encode 47-amino acid segments which differ by 15 amino acids (Fig. 1).

The second splice variation occurs at a position close to the COOH-terminus and involves the insertion of miniexons. Surprisingly, Drosophila RNA is also alternatively spliced.

Figure 1. Deduced amino acid sequence of human dynamin. Sequences that are conserved within the GTPase superfamily are indicated as element I, II, and III. The five amino acids that are different in rat dynamin are indicated above or below the main sequence in bold type. The two points of alternative splicing are shown as alternate translation. In the internal variants a segment of the molecule is divergent compared to these other family members and includes a proline-arginine-rich COOH-terminal domain specific to the dynamin homologues.

C-terminal variants

<table>
<thead>
<tr>
<th>mRNA, has no insert</th>
<th>mRNA has 4-bp insert</th>
<th>mRNA has 37-bp insert</th>
</tr>
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<tbody>
<tr>
<td>GAPVRRPPGAPDPFSCPQPQPVFK</td>
<td>GAPVRRPPGAPDPFSCPQPQPVFK</td>
<td>GAPVRRPPGAPDPFSCPQPQPVFK</td>
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spliced at this position, although it is through replacement of alternative 3'-exons rather than through the insertion of mini-exons (van der Bliek and Meyerowitz, 1991). In a collection of 16 human cDNAs we found four variations at this position: (1) without a mini-exon (in 10 cDNAs), (2) with a 4-bp insertion which results in a frameshift and thus a different COOH-terminus (in 4 cDNAs), (3) a 37-bp insertion which contains a stop codon (in 1 cDNA), and (4) the same insertion encoding an identical COOH-terminus but with additional noncoding sequence (in 1 cDNA). The 37-bp insertion is present in the published sequence of rat dynamin (Obar et al., 1990). It seems likely that the other splice variants will also be found in rat. The 3'-untranslated sequence is common to all cDNAs. The cDNA that we have used for the experiments described below was the most abundant in the collection. It has the second of the two internal variants common to all cDNAs. The size of the dynamin RNAs was determined with an RNA ladder. A comparison between the high level of expression in brain and lower levels in other tissues was made using different exposure time (30 min vs 14 h).

Figure 2. RNA blot analysis of human dynamin expression. An RNA blot containing poly(A)* RNA from human heart, brain, placenta, lung, liver, skeletal muscle, kidney, and pancreas was hybridized with radiolabeled cDNA corresponding to the 5'-half of the dynamin sequence. The size of the dynamin RNAs was determined with an RNA ladder. A comparison between the high level of expression in brain and lower levels in other tissues was made using different exposure time (30 min vs 14 h).

Expression of mutant and wild-type dynamin was monitored by Western blot analysis using a rabbit polyclonal antibody (748), which is monospecific for dynamin. A 100-kD protein, corresponding to dynamin, was detected in nontransfected HeLa cells and was substantially increased in cells transfected with either the wild-type or the mutant dynamin constructs (Fig. 3 b). This band was not detected by preimmune serum (data not shown). Detection of a minor band at ~45 kD was variable and levels of this protein were unaffected by infection or transfection. Dynamin expression was observed 3 h after transfection and continued to accumulate over 12 h. By 5 h after transfection, both wild-type and mutant dynamin accumulated in the cell population to ~14-fold higher than endogenous levels (average ~7.5-fold).

Subcellular fractionation of HeLa cells in isotonic sucrose indicated that dynamin was approximately equally distrib-
uted between the particulate and the soluble fraction (Fig. 3 c, lanes a) as determined by densitometric analysis (see Materials and Methods). This distribution was similar to that reported for brain tissue and PC12 cells (Scaife and Margolis, 1990). Tubulin served as a control for the almost quantitative release of soluble proteins. The distribution of dynamin was unaltered in virally infected and mock-transfected cells (Fig. 3 c, lanes b). At high levels of expression (>13-fold over endogenous), the amount of sedimentable wild-type dynamin increased ~4-fold (Fig. 3 c, lanes c), while most of the excess dynamin was present in the soluble pool. Similar results were obtained with mutant dynamin except that the amount of sedimentable dynamin was approximately eightfold over endogenous (Fig. 3 c, lanes d). Lysis under higher salt conditions results in complete redistribution of dynamin into the soluble pool (data not shown). These results suggest that dynamin interacts peripherally with a saturable component on membranes and that this interaction might be modulated by GTP binding.

Expression of Mutant Dynamin Inhibits Receptor-mediated Endocytosis of Transferrin

To follow receptor-mediated endocytosis in transfected HeLa cells, Tfn was biotinylated via a cleavable disulphide bond (the complex is referred to as BSST) for use as a ligand. Internalization was measured by following the latency of transferrin to the low molecular weight reducing agent, MesNa (β-mercaptoethane sulfonic acid) as previously described (Smythe et al., 1992; Carter et al., 1993). The experiment presented in Fig. 4 demonstrates that both the rate and extent of endocytosis of transferrin were inhibited by >40% in cells overexpressing mutant dynamin as compared to mock-transfected cells (i.e., vaccinia-infected cells treated with TransfectAce). HeLa cells overexpressing wild-type dynamin showed neither inhibition nor stimulation of endocytosis over mock-infected cells, indicating that the level of dynamin expression was not rate-limiting. In general, mock transfection or transfection with wild-type dynamin resulted in dynamic family members are boxed. References to the mutations are as follows: SRP54, Bernstein, H., and Walter, P., personal communication; SRP-Ro, Rapiejko and Gilmore, 1992; Ha-ras, Der et al., 1988; Sigal et al., 1986. (B) Western blot analysis of wild-type and mutant dynamin overexpression. Vaccinia-infected cells transfected with a plasmid encoding wt or mutant constructs of dynamin were allowed to express protein for 5 h, harvested and processed for Western blot analysis as described in Materials and Methods. Lane a, a' untreated cells; lane b, b', mock-transfected cells (which were infected with vaccinia virus and transfected without plasmid); lane c, c', cells transfected with wild-type dynamin; and lanes d, d' cells transfected with the element I mutation. a-d, 10 μg of cell lysate protein; a'-d', 50 μg of cell lysate protein. (C) Subcellular distribution of endogenous and overexpressed dynamin. Control, mock-transfected, and transfected cells were lysed by a freeze-thaw cycle and lysates were resolved into particulate and soluble fractions as described in Materials and Methods. Equal amounts of soluble (S) and particulate (P) fractions were analyzed by SDS-PAGE and Western blotting. Tubulin, detected with a polyclonal anti-tubulin antibody (ICN, Irvine, CA) was used as a control to monitor release of soluble contents. Lanes a, untreated cells; lanes b, mock-transfected cells; lanes c, cells transfected with wild-type dynamin; and lanes d, cells transfected with the element I mutation.

Figure 3. Transient expression of wild-type and mutant dynamin constructs in HeLa cells. (A) Point mutations in the nucleotide-binding region of dynamin family members which interfere with function. Shown are the domain structure of dynamin and the three GTP-binding elements conserved among all members of the GTPase superfamily. The invariant lysines in element I and element III interact directly with bound nucleotides (Pai et al., 1990). Two dynamin mutations, K44-→A in element I and K706-→D, predicted to have impaired guanine-nucleotide-binding affinities were generated by site-directed mutagenesis. Analogous point mutations shown to result in a severely impaired guanine-nucleotide binding.
The most likely explanation for the partial block in endocytosis was always observed in cells overexpressing mutant dynamin. In this experiment, the level of overexpression of transfected dynamin was six-fold over endogenous.

In partial reduction of endocytosis as compared to untreated cells. While significant inhibition of receptor-mediated endocytosis was always observed in cells overexpressing mutant dynamin molecules, the extent of inhibition varied. In all experiments (n = 12), the extent of inhibition as compared to cells expressing equivalent levels of wild-type dynamin was 38 ± 13% (range 19–59%).

A correlation between the extent of inhibition of receptor-mediated endocytosis and the levels of expression of mutant dynamin molecules in the transfected population is shown in Fig. 5. Overexpression from the transfected dynamin constructs could be detected at 3 h after transfection and dynamin continued to accumulate in the transfected cell population over the 5-h time course shown (Fig. 5 B). At the indicated times, cells expressing either wild-type or mutant dynamin were harvested and incubated for 15 min with BSST. As can be seen in Fig. 5 A (solid bars), the inhibition of TfN internalization relative to mock-transfected controls increased with increasing levels of expression of mutant dynamin. The degree of inhibition of receptor-mediated endocytosis in cells overexpressing mutant dynamin was 31, 59, and 65% at 3, 4, and 5 h after transfection, respectively. At 5 h after transfection, cells expressing wt dynamin exhibited partially reduced endocytosis, possibly due to prolonged viral infection. Inhibition due to expression of mutant dynamin is therefore measured relative to cells expressing wt dynamin. These results demonstrate that expression of mutant dynamin greatly reduces receptor-mediated endocytosis in mammalian cells.

**Immunofluorescence Assay Reveals a Subpopulation of Cells with Severely Inhibited Endocytosis**

The most likely explanation for the partial block in endocytosis in the cell population, given that transfection efficiencies are 25–60%, is cell-to-cell variability in the levels of dynamin expression. For this reason, an immunofluorescence assay was developed to demonstrate that receptor-mediated endocytosis is strongly inhibited in a subpopulation of cells. In this single cell assay, transfected HeLa cells expressing mutant and wild-type dynamin were incubated for 30 min at 37°C with a biotinylated-monoclonal antibody (HTR-D65) which recognizes the ectodomain of the TfN receptor (White et al., 1990). Unlike BSST, this antibody ligand will not be recycled and will accumulate in cells capable of efficient endocytosis (Weissman et al., 1986). Subsequently, the cells were washed and incubated for 30 min at 4°C with avidin to mask accessible biotin residues. The cells were washed, fixed, permeabilized, and incubated with Texas-red-conjugated-avidin to detect the internalized, biotinylated-anti-TfR antibody. These results illustrated in Fig. 6, demonstrate that mock-transfected cells (Fig. 6; A) or transfected cells overexpressing wild-type dynamin (Fig. 6; B) showed efficient and uniform internalization of receptor-bound antibody. In marked contrast, a subpopulation of transfected cells expressing the mutant dynamin failed to internalize any detectable anti-TfR mAbs (Fig. 6, C and D, arrowheads).

The extent of inhibition determined biochemically corresponded to the fraction of cells unable to internalize HTR-D65 as detected by immunofluorescence. For example, in two experiments in which endocytosis was inhibited in cells expressing mutant dynamin by 29 and 37% as compared to cells expressing wild-type dynamin, 23 and 32% of ~400 cells counted for each condition showed inhibited endocytosis by immunofluorescence. Taken together, these assays provide strong evidence that functional dynamin is required for endocytosis of the TfN receptor.

**Mutant Dynamin Blocks an Intermediate Stage in Coated Vesicle Formation**

We have previously shown that early and late stages in coated vesicle formation can be distinguished by determining the inaccessibility of bound ligand either to bulky probes such as
Figure 6. Complete inhibition of Tfn-receptor internalization in a subpopulation of cells expressing mutant dynamin. Cells grown, infected, and transfected on coverslips as described in Materials and Methods were allowed to internalize surface bound biotinylated anti-Tfn-R antibody, HTR-D65, for 30 min at 37°C. Residual surface bound biotinylated antibody was masked with avidin, cells were fixed, permeabilized, and internalized HTR-D65 was subsequently detected with a Texas-red-conjugated ultra-avidin as described in Materials and Methods. Mock-transfected cells (A) and cells expressing wild-type dynamin (B) showed uniformly high levels of intracellular HTR-D65. In contrast, a subpopulation of cells transfected with ele I GTP-binding domain mutant dynamin (C and D) were incapable of internalizing detectable HTR-D65 (open arrowheads). The level of inhibition determined biochemically in this experiment was 38% over wild-type.

avidin or antibodies, or to small probes such as MesNa. Internalization of biotinylated ligands into coated vesicles renders them inaccessible to both large and small probes; however, ligands sequestered into deeply invaginated coated pits become inaccessible to avidin while remaining accessible to MesNa (Schmid and Carter, 1990; Schmid and Smythe, 1991). Under normal conditions, the rate and extent of endocytosis as measured using the two probes is identical. However, in cells depleted of ATP, membrane fission is selectively blocked resulting in fivefold greater sequestration from avidin as compared to MesNa (Schmid and Carter, 1990). These two probes were used to determine the stage at which coated vesicle formation was blocked in cells expressing mutant dynamin. The results, illustrated in Fig. 7, demonstrate that the sequestration of Tfn into deeply invaginated coated pits was as effectively inhibited as Tfn internalization in HeLa cells expressing mutant dynamin. These experiments suggest that dynamin activity is required for a relatively early stage of endocytosis, preceding both coated pit invagination and the membrane fission event leading to coated vesicle formation.

The effect of overexpression of dynamin mutants on coat assembly, an earlier stage of endocytosis, was examined by indirect immunofluorescence of the AP2 complex, a major plasma membrane coat protein. The results are shown in Fig. 8. The punctate staining pattern characteristic of cell surface-coated pits was uniformly observed in untreated cells (Fig. 8; A) and was unaltered in transfected cells expressing either (○, ●) wild-type dynamin or ele I GTP-binding domain mutant dynamin (Δ, △) were harvested after an 8-h expression and incubated with BSST for the indicated times. The extent of Tfn internalization was assessed either by MesNa-resistance (●, △) or avidin accessibility (○, Δ) as described in Materials and Methods.
either wild-type (Fig. 8, B) or mutant dynamin (Fig. 8, C). This distribution was uniformly observed throughout the population even though endocytosis of Tfn in these cells was 40% inhibited. The distribution of clathrin, as detected by indirect immunofluorescence was similarly unaffected (data not shown). These results are consistent with the accumulation of coated pits seen by EM of Drosophila shibire mutants (Kosaka and Ikeda, 1983) and suggest that the initiation of coat assembly is unaltered by mutant dynamin.

**Mutant Dynamin Does Not Inhibit Transport from ER to the Golgi Complex**

Proteins involved in vesicular transport could be specific to a single step or could participate in multiple membrane trafficking events (Rothman and Orci, 1992). To determine whether mutant dynamin had a specific effect on the endocytic pathway, we investigated the consequence of its overexpression on the secretory pathway. Transport along the early part of the secretory pathway was monitored by cotransfecting vaccinia-infected HeLa cells with T7-expression constructs encoding VSV-G and constructs encoding wild-type or mutant dynamin. VSV-G which serves as a marker for transport, acquires two endoglycosidase H(endoH)-sensitive N-linked oligosaccharide chains in the ER that are processed to endoH-resistant forms upon transport to the cis/medial Golgi compartments. The results presented in Fig. 9 demonstrate that pulse-labeled VSV-G was quantitatively (>90%) converted to the endoH resistant form after a 60-min chase. The extent of transport to the Golgi was unaffected in cells cotransfected with either wild-type or mutant dynamin constructs. In parallel experiments, endocytosis was inhibited by 30–40% (data not shown). Preliminary immunofluorescence showed an unaltered distribution of VSV-G protein in cells expressing mutant and wild-type dynamin suggesting that export from the Golgi and delivery to the cell surface was also unaffected (data not shown). More thorough examination of the effects of dynamin on post-Golgi trafficking to the cell surface or to lysosomes awaits the generation of stable transformants. These results rule out the possibility of nonspecific cytotoxic effects and indicate that mutant dynamin expression causes a specific reduction in the early stages of receptor-mediated endocytosis.

**Discussion**

**Dynamin Is Required for Receptor-mediated Endocytosis**

We have shown that receptor-mediated endocytosis is inhibited in mammalian cells that overexpress mutant dynamin. Using assays which enable detection of intermediates in coated vesicle formation, the dynamin mutants were found to block endocytosis at a stage preceding the sequestration of ligands into deeply invaginated coated pits. The distribution of clathrin and APs, the major coat constituents, was unaffected in cells overexpressing these dynamin mutants, suggesting that the block follows coat assembly. These results demonstrate that mutations in the GTP-binding domain of dynamin block Tfn-endocytosis in mammalian cells, suggesting that a functional dynamin GTPase is required for receptor-mediated endocytosis via clathrin-coated pits.

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![Figure 8](image_url). The distribution of α-adaptins is unaffected by overexpression of mutant dynamin. The distribution of α-adaptins was determined by indirect immunofluorescence using the AP.6 mAb as described in Materials and Methods. (A) mock-transfected cells; (B) cells expressing wild-type dynamin; (C) cells expressing mutant dynamin. The level of inhibition determined biochemically in this experiment was 40% over wild-type.
GTP-binding domain of VPS10, a dynamin-related protein in yeast, including some which do not affect GTP-binding, have dominant negative effects on vacuolar sorting (Vater et al., 1992). Conversion of pulse-labeled VSV-G protein from an EndoH sensitive form found in the ER to an EndoH resistant form associated with the Golgi complex was determined as described in Materials and Methods.

Dynamin was first linked to endocytosis through the discov-
ery that the Drosophila homologue of dynamin is encoded by the shibire gene (van der Bliek and Meyerowitz, 1991; Chen et al., 1991). Shibire was identified earlier as a mutation that causes temperature sensitive paralysis in adult flies (Grigliatti et al., 1973) and was shown to affect development if the temperature was shifted at a critical time (Poodry et al., 1973). Evidence suggesting that shibire affects endocytosis has come largely through morphological studies which have shown that conditional paralysis correlates with a depletion of synaptic vesicles, and a concomitant increase in coated pits at nerve endings (Koenig and Ikeda, 1989). In other cell types, for example, nephrocytes (Kosaka and Ikeda, 1983; Koenig and Ikeda, 1990) or oocytes (Kessel et al., 1989), shift to the nonpermissive temperature also leads to the apparent disappearance of the endosomal compartment and an increase in cell surface area, reflected by the formation of large membrane invaginations. Coated pits, which appear normal, are often seen at the bottom of these membrane invaginations.

Overexpression of mutant dynamin appeared to spe-
cifically inhibit receptor-mediated endocytosis: transport of VSV-G protein from the ER to the Golgi was unaffected and preliminary results suggested that delivery to the cell surface was also unaffected. Given the incomplete inhibition in these transient transfection assays, more thorough analysis of the effects of overexpression of these dynamin mutants on other vesicular trafficking pathways, including fluid-phase endocy-
tosis, receptor-recycling, post-Golgi trafficking, and lysosomal targeting will await the generation of inducible stably transformed cell lines.

The Shibire Phenotype and the Role of Dynamin in Endocytosis

Dynamin was first linked to endocytosis through the discov-
cortex, the hippocampus, and the cerebral cortex (Nakata et al., 1991; Faire et al., 1992) similar to the distribution of AP subunit transcripts (Robinson, 1989). Recent studies have demonstrated that coated vesicles isolated from synaptic termini are almost exclusively involved in recycling the constituents of synaptic vesicles (Maycox et al., 1992). Thus, high levels of dynamin expression in nerve cells are consistent with its role in expediting synaptic vesicle recycling.

Conclusion

What role might dynamin play in receptor-mediated endocytosis and coated vesicle formation? Our results show that mutant dynamin blocks endocytosis after the initiation of coat assembly but before coated pit invagination. This specifically assigns a role for dynamin in the intermediate stages on the temporal sequence of events that can be distinguished biochemically: coat assembly, receptor recruitment, coated pit invagination and vesicle budding (Schmid, 1993). Consistent with this assignment and with the known affinity of dynamin for GTP, coated pit invagination in vitro requires \( \sim 10 \mu M \) GTP (Carter et al., 1993). Members of the GTPase superfamily, in general, act as monitors and molecular switches which either trigger an event or ensure the fidelity of vectorial reactions (Bourne et al., 1990, 1991). During coated pit invagination dynamin might monitor receptor recruitment, necessary coat protein rearrangements, or the assembly of a molecular complex required for invagination or budding. In response to completion of these requisites to vesicle formation, dynamin might trigger invagination. While we cannot rule out that dynamin plays an active, mechanochemical role in the invagination and/or pinching process, this seems less likely given its membership in the GTPase superfamily. Our observation that dynamin is equally distributed in partuculate and soluble fractions suggests cycling on and off the membrane as part of its function. Further characterization of the role of dynamin will follow from identification of interacting proteins.

Genbank Accession Numbers

The human dynamin cDNA sequences are accessible under Genbank accession numbers L07807-L07810.

References


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