Abstract. Differentiation of hypertrophic chondrocytes toward an osteoblast-like phenotype occurs in vitro when cells are transferred to anchorage-dependent culture conditions in the presence of ascorbic acid (Descalzi Cancetta, F., C. Gentili, P. Manduca, and R. Cancetta. 1992. J. Cell Biol. 117:427-435). This process is enhanced by retinoic acid addition to the culture medium. Here we compare the growth of hypertrophic chondrocytes undergoing this differentiation process to the growth of hypertrophic chondrocytes maintained in suspension culture as such. The proliferation rate is significantly higher in the adherent hypertrophic chondrocytes differentiating to osteoblast-like cells. In cultures supplemented with retinoic acid the proliferation rate is further increased. In both cases cells stop proliferating when mineralization of the extracellular matrix begins. We also report on the ultrastructural organization of the osteoblast-like cell cultures and we show virtual identity with cultures of osteoblasts grown from bone chips. Cells are embedded in a dense meshwork of type I collagen fibers and mineral is observed in the extracellular matrix associated with collagen fibrils. Differentiating hypertrophic chondrocytes secrete large amounts of an 82-kD glycoprotein. The protein has been purified from conditioned medium and identified as ovotransferrin. It is transiently expressed during the in vitro differentiation of hypertrophic chondrocytes into osteoblast-like cells. In cultured hypertrophic chondrocytes treated with 500 nM retinoic acid, ovotransferrin is maximally expressed 3 d after retinoic acid addition, when the cartilage–bone-specific collagen shift occurs, and decays between the 5th and the 10th day, when cells have fully acquired the osteoblast-like phenotype. Similar results were obtained when retinoic acid was added to the culture at the 50 nM “physiological” concentration. Cells expressing ovotransferrin also coexpress ovotransferrin receptors. This suggests an autocrine mechanism in the control of chondrocyte differentiation to osteoblast-like cells.

The chick epiphyseal growth plate is a unique site to investigate ontogenic processes leading to the formation of endochondral bone. The sequence of events includes the formation of hypertrophic cartilage, its invasion by blood vessels from the perichondrium, the erosion of the calcified cartilage, and its replacement by bone tissue. It is generally accepted that hypertrophic chondrocytes in the growth plate degenerate at the site of the transition region from cartilage to osteoid and ultimately die. Several authors have reported that in organ cultures hypertrophic chondrocytes start expressing bone markers and they have proposed that hypertrophic chondrocytes may contribute to the formation of a bone matrix (37, 41, 43). We have recently shown that in vitro chick hypertrophic chondrocytes, obtained as single isolated cells after 3 wk in suspension culture, further differentiate to osteoblast-like cells when transferred to anchorage-dependent culture conditions in the presence of ascorbic acid (11). The cells acquire an elongated or star-shaped morphology, start expressing alkaline phosphatase, reorganize their extracellular matrix by discontinuing the production of cartilage-specific proteoglycans and by switching from the synthesis of type II and type X collagens to the synthesis of type I collagen, and express and secrete other specific differentiation marker proteins. Eventually, calcium mineral is deposited on the newly formed matrix. Supplementing the culture medium with 1 mM retinoic acid during the first 5 d in culture results in a dramatic enhancement of the differentiation process. In vitro hypertrophic chondrocytes undergoing further differentiation produce and secrete large amounts of a noncollagenous glycoprotein with an apparent molecular mass of 82 kD in reducing conditions and of 63 kD in unreducing conditions (11).
In the present manuscript, we extend our previous findings, by investigating the cell proliferation rate during the differentiation process and report on the ultrastructural characterization of the osteoblast-like cell cultures. We also report the purification of the 82-kD glycoprotein from conditioned culture medium and its identification as chicken ovotransferrin, by amino acid sequence determination of its NH₂ terminus. This protein is transiently expressed by the differentiating hypertrophic chondrocytes. In cultures supplemented with retinoic acid, both at a high and at a "physiological" concentration, the protein reaches its highest expression level 3 d after retinoic acid addition, when the shift from the cartilage-specific to the bone-specific collagen occurs. Evidence is also given for the expression of ovotransferrin receptors by cultured chondrocytes. The coexpression by the cells of the growth factor and its receptor is suggestive of an autocrine (or self-responsive) mechanism playing a role in the control of chondrocyte proliferation and differentiation during their transition to osteoblast-like cells.

Materials and Methods

Cell Culture

Cultures of chondrocytes were performed as described (3). Dedifferentiated chondrocytes were obtained by plating on culture dishes cells freshly dissociated from 6-7-old chick embryo tibiae. To reinduce differentiation, dedifferentiated chondrocytes, expanded as adherent cells for 3 wk, were transferred to suspension culture for an additional 3-4 wk until a homogeneous population of single isolated hypertrophic chondrocytes was obtained. Osteoblast-like cells were obtained as described by Descalzi Cancedda et al. (11). Hypertrophic chondrocytes were filtered through a nylon filter, digested with hyaluronidase, and plated at confluence (2 × 10⁵ cells in 30-mm dish) in Coon's modified F12 culture medium containing 10% FCS. After 3 d the medium was supplemented with 100 μg/ml ascorbic acid and 10 mM β-glycerophosphate. The medium was changed every other day without cell passaging. When indicated retinoic acid was added to the culture medium at final concentrations between 10 and 500 nM. Fresh retinoic acid was added and the culture medium was changed every day. Cultures of chondrocytes derived from a single cloned cell were obtained according to Quarto et al. (36). Chick embryo osteoblast cultures were obtained according to the procedure described in Manduca et al. (25) from cells grown out from bone chips.

DNA Measurement

Cell layers were scraped in 0.01% SDS in PBS, and digested overnight at 50°C with proteinase K (50-150 μg/ml in 10 mM Tris HCl, pH 7.8, 5 mM EDTA). DNA content was determined in a DNA fluorometer from Hoefer Scientific Instruments, St. Louis, MO.

Histochemistry

Alkaline phosphatase activity was determined using the histological kit 86 by Sigma Immunochemicals.

Ultrastructural Analysis

Cultures were sacrificed at indicated days by fixation with 4% formaldehyde (freshly made from paraformaldehyde) in 0.1 M phosphate buffer, pH 7.2, postfixed with 1% OsO₄ for 30 min and routinely ethanol dehydrated and Araldite embedded. Thin sections contrasted with U/Pb were examined in a Philips CM10 operated at 80 kV.

Cell Labeling and Protein Analysis

Cells were labeled with [³⁵S]methionine as described by Descalzi Cancedda et al. (9). Aliquots of culture media were run for protein analysis by SDS-PAGE in reducing and unreducing conditions. Immunoprecipitation of specific proteins was performed as previously described (9).

Western Blot Analysis for Transferrin Receptors

Whole cell lysates were prepared by adding to the cells 0.1% SDS in PBS. Aliquots of samples, containing ~300 μg of proteins, were loaded on a 10% SDS-polyacrylamide gel. Electrophoresis was performed in reducing conditions. After electrophoresis the gel was blotted to a BAB5 nitrocellulose membrane (Schleicher & Schuell GmbH, Dassel, Germany) according to the procedure described by Towbin et al. (44). The blot was saturated for 16 h with 2% BSA in TTBS buffer (20 mM Tris HCl, pH 7.5, 500 mM NaCl, 0.05% Tween 20), washed several times with TTBS and incubated with the polyclonal antibody directed against the ovotransferrin receptor (αOTfR) (15) for 2 h at room temperature. After additional washes the detection was performed by a biotin-conjugated anti-rabbit IgG (H + L) (Jackson ImmunoResearch Laboratories Inc., West Grove, PA) and HRP (Jackson ImmunoResearch Laboratories Inc.) using 4-chlor-l-napth (Merck Biochemica, Darmstadt, Germany) as substrate.

RNA Extraction and Northern Analysis

Total RNA was extracted from cells using the guanidinium thiocyanate method (6). For Northern analysis ~10 μg of total RNA were electrophoresed through 1% agarose gel in the presence of formaldehyde and blotted onto Hybond N-membranes. Hybridization and washing conditions were performed as recommended by Amersham Corp. (Arlington Hts, IL). Probes were as follows: BST XI-BST XI fragment of ovotransferrin cDNA (22), pCOL3 for a I collagen (45), pDRS for Ch21 (10), insert of pCP15 for osteopontin (4), and pXCR7 for rRNA (gift from Dr. F. Amaldi, Università di Tor Vergata, Roma, Italy).

Purification of Ovotransferrin

Conditioned culture medium was supplemented with tracer amounts of radioactive medium obtained by metabolic labeling of stage III chondrocytes with [³⁵S]methionine and containing a highly radioactive 69-kD glycoprotein. Medium was freed of collagen by 30% ammonium sulfate precipitation and then concentrated by precipitation with saturated ammonium sulfate. The precipitated proteins were dissolved and dialyzed in 20 mM phosphate buffer, pH 7.6, and loaded on a DE52 column equilibrated with the same buffer. The radioactive peak was collected and an aliquot was analyzed by SDS-PAGE followed by silver staining.

Amino-terminal Sequence Determination

The amino-terminal sequence of the purified protein (0.3 mg) was determined with a pulse liquid sequencer from Applied Biosystems Inc. (Foster City, CA) according to the manufacturer's instructions.

Results

Cell Growth during In Vitro Differentiation

To determine whether chondrocyte differentiation to osteoblast-like cells was accompanied by cell proliferation, we directly compared the growth rates, measured as an increase in the culture DNA content, of two hypertrophic chondrocyte cultures, maintained in the presence of ascorbic acid and grown adherent or in suspension (Fig. 1 A). Cells were plated sparse (4 × 10⁶ cells per 3-cm dish). In the adherent culture, the cell doubling time was ~7 d. In the control, hypertrophic chondrocyte suspension culture, the cell doubling time was >14 d. Fig. 1 B shows results from an independent experiment in which growth rates of hypertrophic
chondrocytes grown adherent in the presence and absence of retinoic acid were compared. Cells were plated at confluence (2 x 10^6 cells per 3-cm dish). In the culture not supplemented with retinoic acid the cell doubling time was ~4 days. The addition of 500 nM retinoic acid to the culture medium enhanced the cell doubling time by approximately twofold. Both in the presence and absence of retinoic acid, the growth curves reached a plateau at the time that mineralization of the extracellular matrix started (Fig. 1, arrows). It should be noted that cell proliferation was observed also when cells were plated at confluence (Fig. 1 B), since the cells have a tendency to grow in multilayers.


cell Ultrastructure, Matrix Formation, and Mineralization

The morphology, organization and mineralization of the extracellular matrix produced by osteoblast-like cells were assessed by EM (Fig. 2). Obvious differences were noted compared with the morphology of the cartilage-like matrix produced by tibial hypertrophic chondrocytes in suspension (unpublished results) and cultured vertebral chondrocytes (16). Fibrils exhibiting a 64-70-nm periodicity, consistent with type I collagen, represented the dominant feature observed both in the absence (Fig. 2, a and b) and presence of retinoic acid (Fig. 2 c). No proteoglycan "granules," typical of cartilage and cartilage-like matrix, were ever observed. Fibrils were coagulated and assembled in coherent bundles in older cultures. Early foci of mineralization consisted of clusters of thin mineral crystals, exhibiting a filament or plate-like habit. At the edges of such clusters spreading of mineralization along collagen fibrils was obvious. Individual collagen fibrils with evidence of discontinuous mineralization along their axis were observed in cultures treated with retinoic acid (14 and 16 d), and in control cultures of embryonic osteoblasts grown out from bone chips (Fig. 2 d). The mineralization highlighted the fibril periodicity, as is commonly seen in early mineralization of bone matrix in vivo. Such individually mineralized fibrils were not detected in the absence of retinoic acid, in spite of the virtual identity of the morphology of the collagenous matrix produced in such cultures with that seen in retinoic acid-treated cultures.

Ovotransferrin Is Transiently Expressed by Hypertrophic Chondrocytes Undergoing Differentiation to Osteoblast-like Cells

The 63-kD glycoprotein secreted by differentiating hypertrophic chondrocytes was purified from conditioned culture medium by ammonium sulfate precipitation, DEAE cellulose chromatography, and HPLC reverse phase chromatography (Fig. 3). The amino acid sequence determination of its NH2 terminus was performed by automated Edman degradation. The sequence of the first 25 amino acids was identical to the published sequence of the NH2 terminus of the chicken ovotransferrin (7) (Fig. 4).

Ovotransferrin mRNA levels in hypertrophic chondrocytes replated as adherent cells and maintained in the presence of ascorbic acid for 0, 20, and 40 days were also determined (Fig. 5). Ovotransferrin mRNA was not detected in the starting hypertrophic chondrocyte population, reached a maximum level after 20 days in adherent culture in the presence of ascorbic acid and was barely detectable at later times. In the same cells, the level of type I collagen mRNA progressively increased while mRNA for Ch21, a carrier protein highly expressed by hypertrophic chondrocytes (9, 10), decreased. Interestingly osteopontin was highly expressed only at late culture times, when mineralization occurred. When the relative amount of the ovotransferrin secreted was investigated in a parallel labeled culture dish, it was found that, after induction, the protein decrease was somewhat slower than expected, based on the mRNA level (not shown). This suggests a possible heterogeneity in the differentiation timing of different culture dishes. Transient expression of the ovotransferrin protein was more clearly observed when chondrocytes were treated with retinoic acid, because this condition gives rise to a synchronous and homogeneous population of differentiating cells (see below).

Differentiation of Osteoblast-like Cells in a Cloned Cell Population

In some experiments, the starting hypertrophic chondrocyte population was derived from a single cloned dedifferentiated cell. The change in the pattern of proteins secreted by these hypertrophic chondrocytes grown adherent in vivo. Such individually mineralized fibrils were not detected in the absence of retinoic acid, in spite of the virtual identity of the morphology of the collagenous matrix produced in such cultures with that seen in retinoic acid-treated cultures.

Retinoic Acid Enhances Differentiation of Hypertrophic Chondrocytes both at High and Physiological Concentration

We have previously shown that retinoic acid supplemented to the culture medium for 5 days at a relatively high concentration (1 #M) highly accelerated the maturation of hypertrophic chondrocytes to osteoblast-like cells (11). It has been suggested that the addition of retinoic acid at nonphysiological concentrations may cause artefacts and that the observed phenotypic changes may not reflect a true cell differentiation (34). In addition the pulse treatment of the cells may cause
Reversion of the phenotype when retinoic acid is removed. Therefore, we performed the experiments by keeping retinoic acid constant, at different concentrations, during culture.

The pattern of labeled proteins secreted by chondrocytes continuously treated with 50 nM retinoic acid is shown in Fig. 7. The pattern of proteins secreted by chondrocytes continuously treated with 500 nM retinoic acid is substantially identical, although the transition to the osteoblast-like pattern is slightly accelerated in the latter culture (not shown). Ovotransferrin was clearly transiently expressed in both cell cultures. Expression of ovotransferrin occurred at the switch from the synthesis of type II collagen to the synthesis of type I collagen (Fig. 8). The maximal expression was observed at day 3, when the pro a1 (II) band disappeared, the a1 (II) band was highly diminished and the type I procollagen was already detectable.

In some experiments retinoic acid was added at a 10-nM concentration. In this condition the differentiation of hypertrophic chondrocytes into osteoblast-like cells was not as rapid as in culture treated with 50 or 500 nM retinoic acid. After 15 d type X collagen was still expressed and the persistence of a large number of polygonal cells in the culture dishes was observed (not shown).

Ovotransferrin expression preceded the expression of alkaline phosphatase. In the retinoic acid–treated culture, alkaline phosphatase activity began to be detectable after ~1 wk and reached its maximal values after ~8–10 d (Fig. 9 B), 1–2 d before the onset of mineralization. At that time the culture presented a distinct pattern of cell growth with the appearance of a reproducible circular swirling pattern previously described in calvaria-derived osteoblast cultures (17). At the same time in the control untreated cultures the large majority of the chondrocytes presented a polygonal morphology and were alkaline phosphatase negative; the small percentage of cells positive for alkaline phosphatase activity presented a more elongated morphology and had a tendency to group in clusters (Fig. 9 A). It is to be noted that, when retinoic acid was present throughout the whole culture, mineralization was already observed after 9–11 d, well before the 3 wk required in the pulse treated cultures (11).

Expression of Ovotransferrin Receptors by Hypertrophic Chondrocytes Undergoing Differentiation to Osteoblast-like Cells

Cell lysates were prepared from hypertrophic chondrocytes plated as adherent cells and maintained in culture for 5 and 14 d in the presence of ascorbic acid. At the same time a lysate was also prepared from a culture maintained for 9 d in the presence of ascorbic acid and retinoic acid. The presence of the tissue (oviduct) specific transferrin receptor was inves-

Figure 2. Ultrastructure of extracellular matrix. (a) Overview of the collagenous matrix produced by adherent osteoblast-like cells grown in the absence of retinoic acid at 47-d culture after mineralization occurred. A patch of early mineralization is obvious (arrow). (b) Matrix produced by osteoblast-like cells in the absence of retinoic acid. Detail of a bundle of collagen fibrils with 64–70-nm periodicity. (c) Matrix produced and mineralized by osteoblast-like cells grown in the presence of 500 nM retinoic acid, 14 d. A patch of mineralization is shown (arrows), together with evidence of mineralization of individual collagen fibrils (arrowheads). (d) Demonstrates the pattern of matrix mineralization observed in cultures of chick embryo osteoblasts outgrown from bone chips. Note the identity of the patterns demonstrated in c and d. Bars: (a) 0.2 μm; (b–d) 0.1 μm.
tigated by Western blot analysis (Fig. 10). The presence of a protein recognized by the anti-transferrin receptor and comigrating with the receptor from the oviduct (not shown) was clearly detectable in all three extracts.

**Discussion**

In the present study, we have extended our previous observation that hypertrophic chondrocytes undergo differentiation to osteoblast-like cells in vitro, when they are transferred to substrate-dependent culture conditions in the presence of ascorbic acid. Here we focused on the modulation of cell proliferation and on the extracellular matrix mineralization. In addition we described the ovotransferrin transient expression during the process.

After their plating on the adhesion permissive substratum, the cell doubling time was enhanced to $\sim$4-7 d depending upon initial cell density. Addition of retinoic acid to the culture medium further enhanced cell proliferation. The measured doubling time was $\sim$2 d. The dependence of cell division on cell shape and anchorage is a well known fact. In experiments where cells of the 3T3 cell line were either held in suspension or allowed to settle on patches of an adhesive material on nonadhesive substratum, it was observed that the patch diameter determines the extent of spreading of individual cells and the probability that they will divide (30).

Differentiating osteoblast-like cells stop proliferating when mineralization occurs. Type I collagen is the only collagen secreted in large amounts at that time. The data presented here demonstrate that the texture and organization of the matrix assembled by the osteoblast-like cells derived...
Figure 8. Proteins secreted by cultured chondrocytes treated with retinoic acid. Hypertrophic chondrocytes were replated as adherent cells and maintained in medium supplemented with ascorbic acid and β-glycerophosphate (control) or in the supplemented medium with the further addition of 500 nM retinoic acid. In the lower panel aliquots of [35S]methionine-labeled culture media were immuno-precipitated with specific anti-collagen antibodies and run for protein analysis on SDS-PAGE. Numbers refer to days in culture. OTF indicates ovotransferrin. Polyacrylamide gel concentration was 7.5%. Electrophoresis was performed in reducing conditions.

Figure 9. Expression of alkaline phosphatase in a retinoic acid-treated culture. Histochemical staining was performed 9 d after plating on chondrocytes maintained in the presence of 500 nM (B) or on control chondrocytes not supplemented with retinoic acid (A). Bar, 100 µm.

In culture, a major expression of alkaline phosphatase activity is observed before mineralization. In addition, osteopontin is highly expressed at the time mineralization occurs. The same sequence of events, i.e., alkaline phosphatase activation, arrest in cell proliferation, osteopontin expression and deposition of mineral in the extracellular matrix on type I collagen fibers was observed also in cultures of differentiating osteoblasts (17, 31).

The finding that the hypertrophic chondrocyte to osteoblast-like cell transition is detectable in cultures derived from cloned cells, together with the rapid (2–3 d) phenotypic change observed in retinoic acid treated cultures, are in agreement with the existence of an additional maturation step of hypertrophic chondrocytes and strongly against the possibility of a selection during culture of existing cell subpopulations.

Our results demonstrate that hypertrophic chondrocytes undergoing differentiation to osteoblast-like cells transiently express high levels of transferrin. Transferrins are a group of glycosylated, iron binding, 80-kD proteins (7, 8).
is preceded by active cell proliferation (39, 40). Trans-
ferin is produced by liver and yolk sac and is present in the
organ cultures of both developing kidneys and teeth, transfer-
nation of different cell populations and overt cell differenti-
ation rate of different cell populations and overt cell differenti-
ation has not been demonstrated yet.

We have evidence that a transition of hypertrophic chon-
drocyte to osteoblast-like cell may occur also in vivo at the
chondro-osseous junction of long bones (Galotto, M., G.
Campagnile, G. Robino, F. Descalzi Cancedda, P. Bianco,
and R. Cancedda, manuscript in preparation). Given the di-
rect effect of transferrin on the proliferation of induced
mesenchymes and in tissue morphogenesis, it may be postu-
lated that, by paracrine mechanisms, hypertrophic chondro-
cytes at the chondro-osseous junction induce or enhance
proliferation and osteoblastic differentiation of the em-
byronic periosteum mesenchyme and/or undifferentiated
mesenchymal cells accompanying the ingrowth of meta-
physseal blood vessels.

The role of retinoic acid in cartilage differentiation has
been reviewed in a variety of articles including ref 11. Here
we have confirmed and extended our previous report that at
later stages of chondrocyte differentiation retinoic acid pro-
motes maturation of chondrocytes to stage II (hypertrophic)
and stage III (osteoblast-like) (11). This is in agreement with
the observation made by other authors that retinoic acid in-
duces differentiation in cultured chondrocytes (29, 34). In
particular we observed that the continuous presence of
retinoic acid at a physiological concentration (i.e., a concen-
tration similar to the concentrations determined in the
posterior chick limb bud whereas lower concentration was
present in the anterior limb bud) (42) in the culture medium
accelerates the chondrocyte differentiation process toward
the organization of a mineralized bone-like matrix in a man-
ner comparable with the acceleration observed when high
concentrations of retinoic acid are present.

In our culture system, retinoic acid has an inductive effect
on ovotransferrin expression. Levels of transferrin and trans-
ferin mRNA are responsive to added retinoids also in Ser-
toli cells (21). Retinoic acid responsive elements can be
identified by computer assisted search on transferrin pro-
moters (19). It is interesting to note that the Ch21 protein,
a newly described marker of hypertrophic chondrocytes be-
longing to the lipocalin family (extracellular transport pro-
teins for small hydrophobic ligands, such as retinoids and
some of its metabolites) (9, 10), is maximally expressed by
chondrocytes before the highest level of ovotransferrin ex-
pression is reached.

Although it is generally believed that hypertrophic cells
are terminally differentiated cells unable to undergo any fur-
ther differentiation, the alternate view that hypertrophic
chondrocytes can transdifferentiate to either osteoblasts or
marrow stromal cells has long been held. In vitro cultures

Figure 10. Immunoblot analy-
asis of transferrin receptor. Cell lysate from hypertrophic
chondrocytes plated as adher-
ent cells and maintained in the
presence of ascorbic acid for 5
(lane J) and 14 (lane 2) d were
run on 10% polyacrylamide gel, blotted to nitrocel-
lulose filter and subjected to
immunoblot analysis using
\(\alpha\)OV-TfR polyclonal antibod-
ies. In the blot the lysate from
the same cells grown adherent
in the presence of ascorbic
acid and with the further sup-
mlement of 500 nM retinoic
acid was also analyzed (lane
J). Arrow refers to the char-
acteristic 95-kD oviduct
transferrin receptor recog-
nized by the antibodies.
cartilage pieces of different origin have shown that hypertrophic chondrocytes may have osteogenic potential (23, 37, 41, 43, 46). In different species, hypertrophic chondrocytes express in vivo proteins characteristic of the “osteoblast phenotype” (I, 14, 26, 27, 32, 33, 38). A peculiar form of ossification suggestive of a deregulated maturation of chondrocytes into osteoblast-like cells has been described in Thanatophoric Dysplasia, the most common of the lethal neonatal bone dysplasias in man (20).

We have presented the first evidence that hypertrophic chondrocytes in culture undergo differentiation to osteoblast-like cells; however, to prove that the phenotype switch from hypertrophic chondrocytes to osteoblasts occurs in vivo additional experiments are required. The hypothesis can be made that cellular cross-talk between preosteoblasts and hypertrophic chondrocytes, mediated by extracellular matrix and/or secreted factors as ovotransferrin and Ch21, triggers the deposition of the first bone, and qualifies it as a joint endeavor of hypertrophic chondrocytes and osteoblasts.

We thank Dr. Severino Ronchi and Dr. Armando Negri for automated Edman degradation of the purified ovotransferrin; Dr. John Lucas for the cOV-TR antibodies; Drs. Pierre Chambon, Enrico Avvedimento, and Francesca Amaldi for the DNA probes; Prof. Ermanno Bonucci for advice and support; Dr. Ralph Bradshaw for helpful discussions; and Barbara Minuto and Daniela Giacoppo for editorial and secretarial help.

This work was supported by grants from Progetti Finalizzati: “Ingegneria Genetica” and “Applicazioni cliniche della ricerca oncologica,” CNR, Rome and by funds from the Associazione Italiana per la Ricerca sul Cancro, Italy.

Received for publication 20 November 1992 and in revised form 29 April 1993.

References


