Regulation of Cytoplasmic Dynein Function
In Vivo by the Drosophila Glued Complex

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Abstract. The Drosophila Glued gene product shares sequence homology with the p150 component of vertebrate dynactin. Dynactin is a multiprotein complex that stimulates cytoplasmic dynein-mediated vesicle motility in vitro. In this report, we present biochemical, cytological, and genetic evidence that demonstrates a functional similarity between the Drosophila Glued complex and vertebrate dynactin. We show that, similar to the vertebrate homologues in dynactin, the Glued polypeptides are components of a 20S complex. Our biochemical studies further reveal differential expression of the Glued polypeptides, all of which copurify as microtubule-associated proteins. In our analysis of the Glued polypeptides encoded by the dominant mutation, Glued1, we identify a truncated polypeptide that fails to assemble into the wild-type 20S complex, but retains the ability to copurify with microtubules. The spatial and temporal distribution of the Glued complex during oogenesis is shown by immunocytochemistry methods to be identical to the pattern previously described for cytoplasmic dynein. Significantly, the pattern of Glued distribution in oogenesis is dependent on dynein function, as well as several other gene products known to be required for proper dynein localization. In genetic complementation studies, we find that certain mutations in the cytoplasmic dynein heavy chain gene Dhc64C act as dominant suppressors or enhancers of the rough eye phenotype of the dominant Glued1 mutation. Furthermore, we show that a mutation that was previously isolated as a suppressor of the Glued1 mutation is an allele of Dhc64C. Together with the observed dependency of Glued localization on dynein function, these genetic interactions demonstrate a functional association between the Drosophila dynein motor and Glued complexes.

The minus-end directed microtubule motor cytoplasmic dynein has been implicated in numerous intracellular motile processes including mitotic chromosome movements (Pfarr et al., 1990; Steuer et al., 1990; Hyman and Mitchison, 1991), mitotic spindle morphogenesis (Vaisberg et al., 1993), nuclear positioning (Plamann et al., 1994; Xiang et al., 1994), nuclear and mitotic spindle migration during cell division (Eschel et al., 1993; Li et al., 1993), perinuclear localization of the Golgi complex (Corthéz-Theulaz et al., 1992), and retrograde axonal vesicle transport (Paschal and Vallee, 1987; for review see Holzbaur and Vallee, 1994). While it seems clear that cytoplasmic dynein is likely to participate in the transport of multiple cytoplasmic organelles and/or cargoes, there is much to be learned about the mechanisms that regulate specific functions of the dynein motor within cells. One essential aspect of dynein regulation that is a focus of current research is how the cytoplasmic dynein motor is targeted to particular cellular cargoes. In this regard, recent work has drawn attention to the role of the dynactin complex (Gill et al., 1991; Schroer and Sheetz, 1991) as a potential regulatory component involved in the coupling of the dynein motor complex to membrane-bound vesicles.

Dynactin is a 20S complex that has been biochemically characterized in chicken and bovine brain extracts (Gill et al., 1991; Schroer and Sheetz, 1991; Paschal et al., 1993). The complex includes a doublet of 150/160-kD polypeptides, a 45-kD actin-related protein Arp-1/centraclin, conventional actin, actin-capping protein, and polypeptides of 62, 50, 27, and 24 kD (Gill et al., 1991; Schroer and Sheetz, 1991; Lees-Miller et al., 1992; Paschal et al., 1993; for review see Schroer, 1994). Although the dynactin complex is not required to support dynein-mediated microtubule gliding in vitro, dynactin has been shown to stimulate cytoplasmic dynein-mediated in vitro vesicle motility (Gill et al., 1991; Schroer and Sheetz, 1991). The predicted amino acid sequence of the p150 component also contains a conserved motif that is found in the endosome linker protein CLIP-170 (Pierre et al., 1992), which is postulated to participate in connecting the motor cargo complex to the microtubule lattice. These results have fostered a model in which the dynactin complex acts to regulate dynein function at the level of cargo attachment.

Recent studies in Neurospora crassa (Plamann et al.,...
The Drosophila Glued gene shares significant homology with the gene encoding the p150 subunit of the cytoplasmic dynein heavy chain gene, and/or the Arp-1 homologue ACT3 which results in defects in mitotic spindle orientation and nuclear migration (Eschel et al., 1993; Li et al., 1993; Clark and Meyer, 1982; Muhua et al., 1994). Despite this evidence for similar phenotypes produced by mutations in the dynein heavy chain gene and genes encoding apparent dynactin-related subunits, the nature of the interaction between dynein and dynactin in vivo remains to be determined.

The Drosophila Glued gene shares significant homology with the gene encoding the p150 subunit of the dynactin complex in rat and chicken (Swaroop et al., 1987; Holzbaur et al., 1991). The Glued locus was initially identified by the spontaneous mutation, Glued1 (Gl1), which results in a dominant rough eye phenotype in adult flies (Plough and Ives, 1935). In mosaic clones containing the intact Gl1 mutation, the Gl1 phenotype is recessive lethal, demonstrating that the Glued gene is also essential for normal development in Drosophila. Screens for revertants of the dominant rough eye phenotype of Gl1 recovered additional recessive lethal mutations at the Glued locus (Harte and Kankel, 1982). In addition to late embryonic and larval lethal phenotypes, several of these new Glued alleles displayed a temperature-sensitive maternal effect, suggesting that Glued is required not only for proper nervous system and eye development, but during the early development of the egg or embryo (Harte and Kankel, 1982). In mosaic analyses using a null mutation of Glued, somatic clones of homozygous Glued mutant tissue were not recovered, suggesting that the Glued gene product has an essential function within the cell. Alternatively, Glued might be required for the successful differentiation and incorporation of particular cell types into developing tissues (Harte and Kankel, 1982).

The Drosophila Glued gene is expressed throughout development (Swaroop et al., 1986), consistent with the genetic data suggesting that Glued has an essential role in Drosophila development (Harte and Kankel, 1982). The molecular characterization of the dominant Gl1 mutation revealed the insertion of a transposon into the Glued coding sequence, resulting in the production of a truncated message that could potentially encode a truncated polypeptide (Swaroop et al., 1985). The dominant nature of the Gl1 mutation suggests that the truncated Glued transcript encodes a protein product that acts as a “poison” to disrupt the function of the wild-type polypeptide.

We previously cloned Dhc64C, a gene encoding a Drosophila cytoplasmic dynein heavy chain (Li et al., 1994), and subsequently isolated recessive lethal mutations in the gene (Gepner, J., M.-g. Li, S. Ludmann, C. Kortas, K. Boylan, M. McGrail, and T. S. Hays, manuscript in preparation). In addition to the recessive lethality, we have noticed additional phenotypes in adult flies that are doubly heterozygous for certain combinations of the dynein mutations. These phenotypes include female sterility, male sterility, bristle defects, and a rough eye phenotype similar to that observed in flies that contain the dominant Gl1 mutation (Gepner, J., manuscript in preparation). The Dhc64C mutations provide tools to analyze the regulation and function of the dynein motor. In this report, we use mutations in both the dynein heavy chain and Glued genes to investigate the interaction between the Drosophila cytoplasmic dynein motor and the dynactin-related Glued complex. We identify and characterize the Glued polypeptides as components of a 20S complex similar to vertebrate dynactin and present cell biological and genetic observations that indicate the physical and functional interaction of the Glued and dynein motor complexes.

Materials and Methods

Fly Stocks and Genetic Analyses

The Dhc64C mutations used in this study were isolated by a standard F2 screen for recessive lethal mutations under the deficiency Df(3L)IOH (64B10–12, 64C5–9) which removes the chromosomal region containing the dynein gene. The third chromosome deficiency Df(3L)IOH was provided by J. Garbe (University of California, Berkeley). For the purposes of the studies described here, the identity of the dynein mutations is established by the rescue of the recessive lethal phenotype of the dynein mutations by an X-linked element insert containing a genomic copy of the Dhc64C transcription unit (designated P(Dhc+); see Table III). Molecular clones containing the intact Dhc64C transcription unit were previously reported (Li et al., 1994). A detailed description of the isolation and phenotypic characterization of the dynein mutations will be reported elsewhere (see also McGrail, M., J. Gepner, M.-g. Li, S. Ludmann, K. Boylan, S. Va, durai, and T. S. Hays, 1994. Mol. Biol. Cell Abstracts. S:131a).

The stocks Gl1 Sb, Df(3L)GI1+6, and Su(Gl)h102 were provided by Dr. Douglas Kankel, Yale University, and are described in Harte and Kankel (1982). The alleles egalitarian+, Bicaudal-DR26, cappuccino+, and spin+, were provided by Dr. Trudi Schüpbach, Princeton University, and are described in Manseau and Schüpbach (1989), Suter et al. (1989), and Schüpbach and Weischaus (1991). The second chromosome deficiency Df(2L)TW19, which removes the Bicaudal-D gene, was obtained from Dr. Ruth Steward, Rutgers University, and is described in Steward et al. (1987).

For the analysis of cytoplasmic dynein and Glued localization in the female-sterile combination of dynein alleles Dhc64C+;Dhc64C+, virgin females of the genotype Dhc64C+/TM6B, D HU were crossed with males of the genotype Dhc64C+/TM6B, D HU, and female progeny that were wild type for the dominant marker mutations Dichaete (D, outstretched wings) and Humeral (Hu, extra hairs on the shoulder) were selected. In the analysis of cytoplasmic dynein and Glued localization in the Bicaudal-DR26 mutant, Bicaudal-DR26;Df(2L)TW19 females were recovered from the cross Bicaudal-DR26;CyO×Df(2L)TW19/CyO. Markers and other chromosomes used are described in Lindsley and Zimm (1992).

Analysis of the genetic interactions between the Dhc64C alleles and the Gl1 mutation was performed by crossing heterozygous virgin females from the stock Gl1 Sb/TM6B, D HU with heterozygous males of the following genotypes: Df(3L)IOH/TM6B, D HU; Dhc64C+/TM6B, D HU; Dhc64C+/TM6B, D HU; Dhc64C+/TM6B; D HU; Dhc64C+/TM6B, D HU; Dhc64C+;TM6B, D HU; Dhc64C+;TM6B, D HU. In the progeny, flies

1. Abbreviation used in this paper: Gl1, Glued.
heterozygous for the Gl filament mutation and the dynein mutation were identified by the presence of adults which show the dominant bristle phenotype caused by the Sb (subbrite, short blunt bristles) mutation, but were wild type for the dominant wing and bristles of the D (Dichaete) and Hu (Humeral) mutations. To examine the effect of an additional copy of the wild-type Dhc64C gene in flies heterozygous for the Gl filament mutation and the dynein alleles Dhc64C- or Dhc64C-17, an X chromosome bearing the wild-type Dhc64C transgene was separately crossed into the Dhc64C- and Dhc64C-17 backgrounds and stocked. Males of the genotype P(Dhc+/Y); Dhc64C-17/TM6B, D Hu and P(Dhc+/Y); Dhc64C-17/TM6B, D Hu were crossed to virgin Gl Dhc64C/TM6B, D Hu females. All of the female progeny from this cross carry one X chromosome bearing the Dhc64C transgene P(Dhc+).

Complementation analyses between the Su(G2)102 mutation and the Dhc64C mutations were performed by crossing heterozygous Su(G2)102/TM6B, D Hu virgin females with males heterozygous for the deficiency Df(3L)10H or the appropriate Dhc64C allele, and TM6B, D Hu. The critical class was scored by the absence of the TM6B, D Hu third chromosome balancer dominant marker mutations Dichaete (D) and Humeral (Hu) in adult progeny.

To test the ability of the dynein transgene to rescue the lethality of Su(G2)102 in combination with Df(3L)10H and the Dhc64C alleles Dhc64C-6, Dhc64C-17, and Dhc64C-3; the following crosses were performed: heterozygous w, P(Dhc+/Y); Su(G2)102/TM6B males were crossed with heterozygous, virgin w/v, Df(3L)10H/TM6B, w/v; Dhc64C-6/ TM6B, w/v; Dhc64C-17/TM6B or w/v; Dhc64C-3/TM6B females. Rescue of the critical class (Su(G2)102/mutant) was scored by the presence of w+ Dichaete Dichaete Humeral adults in the progeny from each cross. Because the wild-type dynein transgene is carried on the X chromosome and is contributed by the male, only female progeny inherit the wild-type dynein transgene and should be rescued. The absence of male progeny that carry the dynein mutations but lack the paternally derived X-linked wild-type dynein transgene demonstrates that the rescue of lethality in the female progeny is due to the presence of the dynein transgene.

**RNA Blot Analyses**

Total RNA was isolated from ovaries and testes, staged collections of embryos, larvae, pupae, and heads, as described previously (Li et al., 1994). 25 μg total RNA was run on a 7.5% agarose-formaldehyde denaturing gel, and blotting, hybridization, and washing conditions were as described previously (Li et al., 1994). A 1.8-kb BamHI fragment from the Glued cDNA, provided by Dr. Alan Garen, which encodes the ~600 COOH-terminal residues of the Glued open reading frame (Swaroop et al., 1987), was cloned in frame into the expression vector pGEX1 (Smith and Johnson, 1988) and expressed in the bacterial strain HB101. Inclusion bodies containing the glutathione S-transferase-Glued fusion protein were prepared as described (Li et al., 1994) and fractionated by SDS-PAGE. Gel slices corresponding to the glutathione-S-transferase-Glued fusion protein were excised and used to elicit a polyclonal serum in rat as described by Towbin et al. (1979). Proteins were electrophoresed on 0.75 mm, 7.5% polyacrylamide slab gels prepared with a 1:100 ratio of bis-acrylamide/total monomer. To better resolve the migration of the truncated Glued polypeptide from the wild-type, samples shown in Fig. 3 were run on 6.5% polyacrylamide gels. Gels were stained with Coomassie brilliant blue or electroblotted to PVDF membrane (Millipore Corp., Bedford, MA). Tubules were probed with the affinity-purified anti-Glued antibody diluted 1:100, or the affinity-purified anti-dynein heavy chain antibody PEP1 (Li et al., 1994) diluted 1:500, in PBS/0.05% Tween-20 containing 0.2% BSA. Hardcopy image prints were produced on a dye sublimation printer (Phaser IISDX; Tektronix, Inc., Beaverton, OR).

**Microtubule-associated Protein Preparations**

Ovaries from well-fed 3-d-old wild-type OregonR females and heterozygous Gl/TM6B females were dissected in EBR buffer (130 mM NaCl, 5 mM KCl, 2 mM CaCl2, 10 mM Hepes, pH 6.9), placed on ice until 100 μl of ovaries were obtained, quick frozen in liquid nitrogen, and stored at -80°C until use. Heads and embryos were collected from the wild-type OregonR and the Gl/TM6B stocks, quick frozen in liquid nitrogen, and stored at -80°C until use. Microtubule-associated proteins from each tissue were prepared as described previously (Hays et al., 1994). Briefly, ~1 ml packed ovaries, 6 ml packed heads, or 15 ml packed embryos, were rinsed three times in PMEG buffer (100 mM Pipes, pH 6.9, 5 mM MgOAc, 5 mM EGTA, 0.1 mM EDTA, 0.5 mM DTT, 0.9 M glycerol) plus protease inhibitors (10 μg/ml aprotinin, 1 μg/ml leupeptin, 1 μg/ml pepstatin, 0.1 μg/ml each of soybean trypsin inhibitor, n-tosyl-l-arginine methyl-ester, and benzamidine). Tissues were homogenized on ice in a glass homogenizer in 1.5 vols PMEG plus protease inhibitors. The homogenate was spun at 125,000 g for 40 min at 4°C, and the supernatant was pulled from between the top lipid layer and the insoluble pellet at the bottom of the tube. All subsequent steps were carried out at 4°C. Microtubules were assembled in the high-speed supernatant by the addition of GTP to 1.0 mM and taxol to 20 μM, plus 20 μM cytochalasins B and D to inhibit assembly of filamentous actin. After 15 min incubation with gentle rocking, endogenous ATP was depleted by the extract by the addition of hexokinase and glucose to final concentrations of 10 U/ml and 100 mM, respectively, and the extract incubated further for 45 min. The extract was underlaid with a 1/4 vol 15% sucrose cushion and centrifuged at 37,000 g for 30 min in a rotor (SW50.1; Beckman Instruments, Inc., Fullerton, CA) to pellet microtubules and microtubule-associated proteins. The microtubule pellet was washed once by resuspension in PMEG plus taxol and recentrifuged. The microtubule pellet was extracted with PMEG containing 10 mM Mg-ATP and 20 μM taxol, and recentrifuged at 37,000 g for 15 min in an SW50.1 rotor plus adapters for 5 × 41 mm open-topped ultraclear tubes (Beckman Instruments, Inc.).

**Immunoblotting**

SDS-PAGE and immunoblotting were carried out as described by Laemmli (1970) and Towbin et al. (1979). Proteins were electroblotted to a 0.75 mm, 7.5% polyacrylamide slab gels prepared with a 1:100 ratio of bis-acrylamide/total monomer. To better resolve the migration of the truncated Glued polypeptide from the wild-type, samples shown in Fig. 3 were run on 6.5% polyacrylamide gels. Gels were stained with Coomassie brilliant blue or electroblotted to PVDF membrane (Millipore Corp., Burlington, MA). Blots were probed with the affinity-purified anti-Glued antibody diluted 1:100, or the affinity-purified anti-dynein heavy chain antibody PEP1 (Li et al., 1994) diluted 1:500, in PBS/0.05% Tween-20 containing 0.2% BSA and 0.2% Triton X-100 (Tropix, Bedford, MA). Alkaline phosphate-conjugated secondary antibodies were diluted in the same. Blots were developed with NBT and BCIP (Sigma Chemical Co.) in alkaline phosphate buffer (100 mM NaCl, 5 mM MgCl2, 100 mM Tris, pH 9.5), or with a nonradioactive chemiluminescence detection system (Trotix) according to the supplier's instructions. Images of gels were captured on an ISRS Indigo Video board installed on a Silicon Graphics R3000 Indigo workstation (Silicon Graphics Computer Systems, Mountain View, CA). Images of blots were obtained from blots and x-ray films scanned into a Macintosh Power PC 8100 (Apple Computer Corp., Cupertino, CA) with a flat bed scanner (ScanJet IIc; Hewlett Packard, Corvallis, OR) using Adobe Photoshop 3.0 software (Adobe System, Inc., Mountain View, CA). Hardcopy image prints were produced on a dye sublimation printer (Phaser IISDX; Tektronix, Inc., Beaverton, OR).

**Immunolocalization**

Ovaries were dissected, fixed, and prepared for immunocytochemistry as described previously (Li et al., 1994). Briefly, ovaries were dissected from 2-3-d-old females in EBR and fixed for 5 min in 100 μl devitellinizing buffer/600 μl heptane using the method of Cooley et al. (1992). Devitellinizing buffer = 1 vol buffer B1 vol 36% formaldehyde/4 vol H2O. Buffer B contains 100 mM KH2PO4/K2HPO4, pH 6.8, 450 mM KCl, 150 mM NaCl, and 20 mM MgCl2.H2O. After fixation, ovaries were rinsed three times in PBS, rinsed three times in PBS/0.1% Triton X-100 (BFT), and washed for
1–2 h in PBT with gentle rocking. The ovariololes were then teased apart, washed for 1–2 h on a rotating wheel (Cole-Parmer Instrument Co., Chicago, IL), and blocked in PBT containing 1% BSA for 1–2 h at room temperature. The ovariololes were double labeled with the affinity-purified rabbit anti-dynein antibody PEP1 (Li et al., 1994) diluted 1:5 or 1:50, and the affinity-purified rat anti-Glued antibody diluted 1:5. The anti-dynein and anti-Glued antibodies were detected with anti-rabbit FITC-conjugated (Boehringer Mannheim Biochemicals, Indianapolis, IN) and anti-rat Texas red-conjugated (Jackson Immunoresearch Laboratories, West Grove, PA) secondary antibodies, respectively. Antibody incubations were performed overnight at 4°C. The secondary antibodies were used at a final dilution of 1:100 after preabsorbing against fixed embryos at a dilution of 1:10. All antibodies were diluted in PBS/0.1% Triton X-100 containing 1% BSA. Ovariololes were mounted in a solution of PBS/90% glycerol containing 1 mg/ml p-phenylenediamine (Sigma Chem. Co.), and examined on a diaphot (Nikon Inc. Instrument Group, Melville, NY) microscope with a confocal imaging system (MRC-600; Bio-Rad Laboratories, Hercules, CA), using a 60×/1.4 planapochromat lens.

**SEM Analysis**

Drosophila heads were dehydrated in an ethanol series as described (Carthew and Rubin, 1990) and prepared for scanning electron microscopy by critical point drying and coating with gold-palladium. Images were recorded on film (type 55; Polaroid Corp., Technical Imaging Products, Cambridge, MA).

**Results**

**Drosophila Glued Gene Product Is a Component of a 20S Complex**

To initiate our analysis of the *Drosophila Glued* gene, we characterized the expression profile of *Glued* during development. Our results, like those reported by Swaroop et al. (1986), show that the *Glued* transcript is expressed throughout embryonic development. Our analysis also shows that *Glued* is expressed in heads, testes, and ovariololes of adult flies (Fig. 1 a). The developmental profile of the *Glued* gene is highly similar to that of the cytoplasmic dynein heavy chain gene, *Dhc64C* (Li et al., 1994). Both the *Glued* and *Dhc64C* transcripts are abundantly expressed in the ovariololes of adults, and are present at high levels in 0–2-h embryos (Fig. 1 a, and Li et al., 1994). The high level of *Glued* transcript present in 0–2 h embryos indicates that, like *Dhc64C*, there is a substantial maternal contribution of *Glued* in the early embryo.

The polypeptides encoded by the *Glued* gene exhibit biochemical characteristics similar to those of the homologous vertebrate gene products. As shown in Fig. 1 b, we used an affinity-purified rat polyclonal serum specific for the *Drosophila* Glued polypeptides (see Materials and Methods) to determine whether the polypeptides were components of a multiprotein complex. Analysis of *Drosophila* ovary cytosol by sucrose density centrifugation revealed that a doublet of polypeptides of ~145/160-kD are recognized by the Glued antibody, and sediment in the ~20S fraction of the sucrose gradient (Fig. 1 b, bottom). The apparent molecular weights of the Glued polypeptides and their sedimentation as a 20S particle are similar to the characteristics of the homologous p150 subunits in vertebrate dynactin (Gill et al., 1991; Schroer and Sheetz, 1991; Paschal et al., 1993). In comparison, the cytoplasmic dynein complex in ovary extracts sediments at a lower S-value as detected by antibodies that recognize the heavy chain polypeptide (Fig. 1 b, top). The *Drosophila* Glued polypeptides, like the homologous vertebrate poly-

**Glued and Cytoplasmic Dynein Colocalize Throughout Drosophila Oogenesis**

The *Drosophila* ovary consists of ~15 ovariololes, each of which contains a series of egg chambers of increasing developmental age (for reviews on *Drosophila* oogenesis, see King, 1970; Spradling, 1993). The egg chamber is formed at the tip of the ovariololes in the germarium, where the stem cells reside. Oogenesis begins with a stem cell division, which produces a cytoklast that will then undergo four rounds of cell division with incomplete cytokinesis to produce a cyst of 16 cells that remain connected by intercellular bridges called ring canals. As the 16-cell cluster moves posteriorly through the germarium, one of the 16 cells is specified to become the oocyte, while the other 15 adopt the fate of nurse cells. Somaically derived follicle cells migrate around the 16-cell cluster, eventually enveloping the cluster as it buds off the germarium to form the stage 1 follicle, or egg chamber. As the egg chamber matures, the nurse cells supply the oocyte with the materials required for growth. Ultimately, during stages 10B–11 the nurse cells transfer their entire cytoplasmic content into the oocyte and subsequently degenerate as formation of the mature egg is completed.

We have previously reported that during *Drosophila* oogenesis cytoplasmic dynein accumulates in the presumptive oocyte in region 2b of the germarium, and is asymmetrically localized to the posterior pole of the oocyte during stage 9 of oogenesis (Li et al., 1994). The pattern of Glued localization is indistinguishable from that of cytoplasmic dynein (Fig. 2, a–d). Glued is initially evenly distributed throughout all cells in the anterior of the germarium (Fig. 2 c, arrowhead), then accumulates in the single cell destined to become the oocyte in region 2b of the germarium (Fig. 2 c, arrow). Similar to the cytoplasmic dynein heavy chain, the Glued polypeptides remain enriched in the oocyte as the egg chamber matures and later during stage 9 become localized to the posterior pole of the oocyte (Fig. 2 b and d, arrowheads).

**Glued Localization in Oogenesis Requires Cytoplasmic Dynein**

To address whether the colocalization of dynein and Glued...
Figure 1. Developmental profile of the Glued mRNA and behavior of the Glued gene product. (a) Top: RNA blot containing total RNA isolated from adult ovaries; 0-2, 2-4, 4-8, 8-12, and 12-24-h embryos; larvae (mixed instars); pupae; and adult testes and heads. The blot was hybridized with a 32P-labeled DNA probe produced from the Glued cDNA. Bottom: as a control to monitor equal loading of the RNA, the blot was subsequently rehybridized with a probe derived from the Drosophila RP49 gene (Vaslet et al., 1980). (b) Immunoblot demonstrating the behavior of the dynein and Glued polypeptides on a sucrose density gradient. 1 mg total protein from a high-speed ovary extract was fractioned on a 5-20% sucrose density gradient, then collected into 0.5 ml fractions. 15-µl samples from each fraction of the gradient were analyzed in duplicate on separate gels by SDS-PAGE and immunoblotting with the affinity-purified anti-dynein heavy chain antibody PEP1 (Li et al., 1994) or the affinity-purified anti-Glued antibody. Sedimentation of the cytoplasmic dynein heavy chain peaks at ~18S (top, fractions 9-12). The Glued antibody detects a ~145/160-kD doublet present in a ~20S complex (bottom, fractions 7-8). Numbers above the panels indicate the gradient fractions; the positions of the 19S and 11S sedimentation standards are indicated below. Left, bottom of gradient. (c) The Glued polypeptides sediment with microtubules and are enriched with cytoplasmic dynein after ATP extraction of the microtubule pellet. Samples containing 1 µg total protein from each fraction of a microtubule-associated protein preparation from ovary extracts were run, in duplicate, on separate 7.5% polyacrylamide gels. Lanes 1-8, Coomassie blue-stained gel. Lanes 2'-8', immunoblot of duplicate gel probed with the affinity-purified anti-Glued antibody. (Lane 1) molecular weight markers; (lanes 2 and 2') homogenate; (lanes 3 and 3') high speed pellet; (lanes 4 and 4') high speed supernatant; (lanes 5 and 5') taxol-stabilized microtubule pellet; (lanes 6 and 6') supernatant of microtubule pellet; (lanes 7 and 7') microtubule pellet after ATP extraction; (lanes 8 and 8') ATP-extracted microtubule-associated proteins. The high molecular weight band in lane 8 is the cytoplasmic dynein heavy chain polypeptide (arrow, DHC; Li et al., 1994). The affinity-purified anti-Glued antibody detects a doublet of polypeptides that sediment with the microtubule pellet (lane 5') and are enriched in the ATP elution fraction (lane 8') with cytoplasmic dynein. The relative molecular mass of each marker is indicated between the panels.

in oogenesis reflects an association of the dynein and Glued complexes, we asked whether Glued localization is dependent on the function of cytoplasmic dynein. To do this, we made use of mutations in the cytoplasmic dynein heavy chain gene that disrupt the localization of dynein during oogenesis. Flies that are doubly heterozygous for the mutations Dhc64C^66 and Dhc64C^6a^2 are female sterile (data not shown). In ovaries derived from these sterile females, the dynein heavy chain does not accumulate at the posterior pole of the oocyte in stages 9 and 10 egg chambers, but is mislocalized in a punctate pattern at the anterior margin of the oocyte (arrows, DHC; Li et al., 1994). The affinity-purified anti-Glued antibody detects a doublet of polypeptides that sediment with the microtubule pellet (lane 5') and are enriched in the ATP elution fraction (lane 8') with cytoplasmic dynein. The relative molecular mass of each marker is indicated between the panels.
Figure 2. The Glued polypeptides colocalize with cytoplasmic dynein during oogenesis and have the same genetic requirements for localization. (a-d) Double-label immunolocalization of cytoplasmic dynein (a and b) and Glued (c and d) in ovaries from wild-type Oregon R females. The Glued polypeptides are evenly distributed in the mitotically active cells in the anterior of the germarium (arrowhead, c) and are differentially localized to the pro-oocyte (arrow, c) coincident with cytoplasmic dynein (arrow, a; Li et al., 1994). The Glued polypeptides remain enriched in the oocyte as the egg chamber matures during its passage down the ovariole. Later in oogenesis, like cytoplasmic dynein (arrowhead, b; Li et al., 1994), the Glued polypeptides are asymmetrically localized to the posterior pole of the oocyte in the stage 9 egg chamber (arrowhead, d). (e-h) Double-label immunolocalization of cytoplasmic dynein (e and f) and Glued (g and h) in stages 9 and 10 egg chambers from heteroallelic Dhc64C6-6/Dhc64C6-12 females. (e and f) In egg chambers from Dhc64C6-6/ Dhc64C6-12 females, dynein is not localized to the posterior pole of the stage 9 and stage 10 oocyte. Instead, dynein is mislocalized in a punctate pattern at the anterior margin of the oocyte (arrows, e and f), and in bright patches within the nurse cell cytoplasm (arrowheads, e and f). (g and h) Like cytoplasmic dynein, localization of the Glued polypeptides to the posterior pole of the oocyte is disrupted in the Dhc64C6-6/Dhc64C6-12 mutant egg chambers. Glued is mislocalized with dynein to the anterior margin of the oocyte (arrows, g and
ruptured, and Glued is colocalized with the mislocalized dynein at the anterior margin of the oocyte (arrows, Fig. 2, g and h) and to the patches within the nurse cells (arrowheads, Fig. 2, g and h). Earlier in oogenesis both dynein and Glued accumulate in the developing oocyte as in wild-type egg chambers (data not shown). The coincident mislocalization of the dynein and Glued polypeptides due to mutations in the cytoplasmic dynein heavy chain gene provides strong evidence that the localization of Glued is dependent on its association with the cytoplasmic dynein motor complex.

**Glued and Cytoplasmic Dynein Localization Exhibit Similar Genetic Requirements in Oogenesis**

To examine further the association of dynein and Glued in vivo, we asked whether mutations in other genes that disrupt the localization of dynein during oogenesis also affect the distribution of the Glued polypeptides. We have previously shown that mutations in the genes *Bicaudal-D* (*Bic-D*) and *ega* (*eggl*) disrupt the accumulation of dynein to the presumptive oocyte (Li et al., 1994). Loss-of-function mutations in either of these genes prevent oocyte differentiation and result in egg chambers containing 16 cells that develop as nurse cells (Suter et al., 1989; Schüpbach and Weischaus, 1991). Like the cytoplasmic dynein heavy chain polypeptide, the Glued polypeptides fail to accumulate in a single cell in egg chambers derived from females homozygous mutant for the *egglwU50* mutation, or hemizygous for the *Bic-DK06* mutation (Fig. 2, i and k).

We also examined the effect of mutations in *cappuccino* (*capu*) and *spire* (Schüpbach and Weischaus, 1991) on the accumulation of Glued during oogenesis. The *capu* and *spire* genes are required for both dorsoventral and anteroposterior axis formation during oogenesis. Mutations in these two maternal effect loci cause a disruption in the accumulation of the posterior group gene products, *staufen* protein and *oskar* mRNA, to the posterior pole of the oocyte (Manseau and Schüpbach, 1989). We have previously shown that both the *capu* and *spire* gene products are also required for the enrichment of cytoplasmic dynein at the posterior pole of stage 9 oocytes (Li et al., 1994). As shown in Fig. 2, we observe a similar disruption of Glued accumulation in the posterior pole of oocytes derived from females that are homozygous mutant for either *capuK12* or *spireRP48* (Fig. 2, j and l). The observations that the Glued and cytoplasmic dynein heavy chain polypeptides exhibit similar genetic requirements for their proper localization during oogenesis is consistent with the physical association of the two complexes.

**A Dominant Mutation in the Glued Gene, GIl, Encodes a Truncated Polypeptide that Associates with Microtubules**

The dominant mutation in the Glued locus, GIl, was previously shown to be caused by the insertion of a transposable element in the 3' coding sequence of the Glued gene (Swaroop et al., 1985). Analysis of Glued expression in heterozygous GIl/+ flies revealed, in addition to the wild-type Glued transcript, a truncated message presumably caused by premature termination of transcription due to the inserted transposon (Swaroop et al., 1985). Like other Glued alleles, the GIl mutation is a recessive lethal mutation and homozygous animals die in late embryogenesis or early larval stages of development (Plough and Ives, 1935; Harte and Kankel, 1982). In addition, the GIl mutation produces a dominant rough eye phenotype in heterozygous adults (Plough and Ives, 1935). This dominant phenotype was shown to be dosage dependent. The addition of small chromosomal duplications carrying extra copies of the wild-type Glued gene into heterozygous GIl/+ flies rendered the severity of the eye phenotype less extreme in a dosage-sensitive manner (Harte and Kankel, 1982). The dosage dependence of the eye phenotype, together with the molecular nature of the lesion, suggests that the GIl mutation encodes a truncated polypeptide that acts as a poison product. In support of this model, we have identified the truncated protein product encoded by the GIl mutation and examined the association of the truncated polypeptide with microtubules and the Glued complex.

To identify the product of the GIl mutation, we compared the Glued polypeptides in microtubule-associated proteins prepared from wild-type versus heterozygous GIl/+ tissues. These studies provided a number of new findings. First, the pattern of Glued polypeptide species encoded by the wild-type Glued gene is tissue dependent. The simplest pattern is observed in extracts from adult heads, in which the predominant species of Glued polypeptide is a single band which migrates at ~160 kD on SDS-PAGE (upper arrow, Fig. 3 a, lane marked *head, +/+*). This species is larger than the predicted molecular mass of 148 kD based on the cDNA sequence (Swaroop et al., 1987). An additional lower molecular mass species of ~145 kD is present in minor amounts (lower arrow, Fig. 3 a, lane marked *head, +/+*). In contrast, the 145-kD species in ovaries appears more abundant than the 160-kD form (Fig. 3 a, lane marked *ovary, +/+*). In embryos, the 160-kD and 145-kD species are present in relatively equal amounts (Fig. 3 a, lane marked *embryo, +/+*). In all three tissues, a third species that migrates at ~135 kD is present in minor amounts (arrowhead, Fig. 3 a). The presence of a single Glued transcript in each of these tissues suggests that the multiple Glued polypeptides may arise from posttranslational modification of a single wild-type product, as opposed to different products resulting from alternative splicing. Second, each of the wild-type Glued species sediments with microtubules (Fig. 3 b, lanes marked *+*/+; see also Fig. 1 c). Lastly, a substantial portion of the Glued polypeptides in each tissue re-
main unbound in the supernatant after microtubule sedimentation (Fig. 3 a, lanes marked +/+), similar to the behavior described for the dynactin complex from chicken (Gill et al., 1991).

The analysis of microtubule-associated protein preparations from heterozygous Glu/+ tissues reveals a truncated Glued polypeptide of ~130 kD in addition to the wild-type polypeptides. The truncated polypeptide is most easily distinguished in the samples derived from head extracts, in which the truncated polypeptide is most abundant (asterisks, Fig. 3 a-c, lanes marked Glu/+). In comparison to wild type, the additional polypeptide present in Glu/+ extracts migrates slightly faster than the smallest wild-type species of ~135 kD. The presence of this additional polypeptide in extracts from heterozygous Glu/+ tissues indicates that the ~130-kD polypeptide represents the truncated protein encoded by the Glu mutation. The variation in the amount of the mutant Glued polypeptide between ovaries, embryos, and heads may reflect variability in the expression of the Glued gene, or in the stability of the truncated Glued product, in these different tissues. Like the wild-type Glued polypeptides, the ~130-kD Glued polypeptide sediments with microtubules (asterisks, Fig. 3 b, lanes marked Glu/+). This result suggests that the COOH-terminal ~20 kD of the wild-type Glued polypeptide is not strictly required for the ability of Glued to sediment with microtubules.

To examine further the behavior of the mutant Glued polypeptide, we analyzed the sedimentation of the truncated polypeptide on sucrose density gradients. Extracts from wild-type and heterozygous Glu/+ tissues were fractionated and analyzed by SDS-PAGE and immunoblotting with the affinity-purified Glued antibody. In wild-type head extracts the predominant 160-kD Glued polypeptide sediments in the 20S fraction of the gradient, similar to the sedimentation profile of the major 145/160-kD doublet of Glued polypeptides present in wild-type ovaries (see Fig. 1 b) and embryos (data not shown). In head extracts from Glu/+ flies, the truncated Glued polypeptide of ~130 kD sediments at ~6–7S (asterisk, Fig. 3 d), while the 160-kD wild-type Glued polypeptide remains in the 20S fraction of the gradient (arrow, Fig. 3 d). A similar result was obtained from the fractionation of extracts from Glu/+ embryos (data not shown). The distinct sedimentation values of the wild-type and mutant Glued polypeptides indicate that the truncated Glued product does not associate with...
the wild-type polypeptide in an ~20S complex. Based on the predicted molecular mass of ~130 kD for the truncated polypeptide, its sedimentation behavior suggests that the mutant Glued polypeptide may be present in the cytoplasm as a monomer. However, our experiments do not exclude the possible association of the truncated Glued protein with other component polypeptides of the Glued complex.

**Mutations in Dhc64C Display a Rough Eye Phenotype Similar to the Gl Mutations**

The *Drosophila* adult eye is composed of ~800 ommatidia arranged in a precise hexagonal pattern (Fig. 4, a and a'; for a review on *Drosophila* eye development, see Dickson and Hafen, 1993). Each of the ommatidia, or unit eyes, is made up of ~20 cells, including eight photoreceptor cells, four cone cells which secrete the lens material, several pigment and associated cells, and a single hair cell. The ordered array of ommatidia in the adult eye arises from a precise spatial and temporal sequence of cell divisions, cell shape changes, and cell signaling events during the differentiation of the eye imaginal disc epithelium in the third larval instar.

We have identified several heteroallelic combinations of *Dhc64C* mutations that produce viable adults displaying a rough eye phenotype. For example, as shown in Fig. 4 (b and b'), the heteroallelic combination of *Dhc64C* alleles *Dhc64C*<sup>32</sup>/*Dhc64C*<sup>6</sup> disrupts the shape, size, and organization of the ommatidia within the eye. This eye phenotype is similar to that observed for the dominant *Gl* mutation (Plough and Ives, 1935; Meyerowitz and Kankel, 1978; Harte and Kankel, 1982) (Fig. 4, c and c'). The *Gl* rough eye phenotype is apparently due to the failure to complete and/or maintain the differentiation of clusters of neuronal precursor cells within the developing retinal epithelium during the third larval instar (Renfranz and Benzer, 1989). The similarity between the rough eye phenotypes of heteroallelic combinations of *Dhc64C* mutations and the dominant *Gl* mutant suggests that the dynein and Glued complexes are components of a common cellular pathway involved in the development of the adult eye.

**Mutations in Dhc64C Act To Suppress or Enhance the Rough Eye Phenotype of Gl**

To obtain more direct evidence that dynein and Glued interact in vivo, we asked whether mutations in the *Dhc64C* gene could act to modify the rough eye phenotype of the dominant mutation *Gl*. In the analysis of flies doubly heterozygous for a *Dhc64C* allele and the *Gl* mutation, we have found that certain *Dhc64C* alleles act as dominant suppressors of the *Gl* rough eye phenotype, while other *Dhc64C* alleles act as dominant enhancers of the *Gl* phenotype (Table I and Fig. 4). For example, in flies doubly heterozygous for the *Dhc64C*<sup>6</sup> allele and the *Gl* mutation, the rough eye phenotype is largely suppressed except in the most posterior portion of the eye (Fig. 4 e), and the shape and packing of ommatidia is very similar to that observed in wild-type flies (Fig. 4 e'). In contrast, the *Dhc64C*<sup>6</sup> and *Dhc64C*<sup>6</sup> alleles enhance the rough eye phenotype of *Gl*; the eyes are significantly smaller (Fig. 4 g) and the remaining ommatidia are more severely disrupted than those observed in flies carrying only the *Gl* mutation (Fig. 4 g'). The specificity of these genetic interactions is demonstrated by the observation that a deficiency that removes the *Dhc64C* gene, *Df(3L)10H*, in heterozygous combination with *Gl* does not alter the *Gl* rough eye phenotype (Fig. 4, f and f'; Table I). Moreover, the *Dhc64C* alleles do not display a phenotype in combination with a deletion that entirely removes the *Glued* gene, *Gl<sup>42</sup>* (Table I; Harte and Kankel, 1982), or with other recessive lethal alleles of *Glued* (data not shown). The observed allele specificity indicates that the dominant genetic interactions depend on the presence of the mutant dynein and *Glued* gene products.

To gain insight into the nature of the interaction between the enhancer alleles of *Dhc64C* and *Gl*, we examined the effect of an additional copy of the wild-type *Dhc64C* gene on the enhancement of the *Gl* rough eye phenotype by the *Dhc64C*<sup>6</sup> and *Dhc64C*<sup>6</sup> alleles. The appropriate crosses were conducted to introduce a wild-type *Dhc64C* transgene carried on the X chromosome into flies that were also doubly heterozygous for the *Gl* mutation and the *Dhc64C*<sup>6</sup> or *Dhc64C*<sup>6</sup> allele. The increased dosage of wild-type dynein heavy chain resulted in the reversal of the enhanced rough eye phenotype (Table I and Fig. 4 h). The shape, size, and alignment of the ommatidia were less disrupted (Fig. 4 h') and more nearly resembled the original *Gl* rough-eyed phenotype. This result demonstrates that the activity of the wild-type *Dhc64C* gene product can functionally compete with the product of the *Dhc64C* enhancer allele. We conclude that the genetic interaction observed between the *Dhc64C* and *Glued* mutations reflects the normal interaction of the dynein and Glued complexes in vivo, rather than an aberrant (neomorphic) activity associated with the mutant polypeptides.

**A Previously Isolated Suppressor of Gl, Su(GI)102, Is a Dhc64C Allele**

Our observations that certain known *Dhc64C* mutations interact with *Gl* suggested that mutations originally isolated as suppressors of *Gl* might identify additional *Dhc64C* alleles. One candidate was the *Su(GI)102* mutation (Harte and Kankel, 1982), which was previously recovered in a screen for EMS-induced revertants of the *Gl* rough-eyed phenotype (Fig. 4, d and d'). The *Su(GI)102* mutation is recessive lethal and maps within the interval between *roughoid* and *hairy* on the left arm of chromosome three (Harte and Kankel, 1982). Significantly, this interval includes the deficiency *Df(3L)10H* which removes the *Dhc64C* gene. To examine whether the *Su(GI)102* mutation is an allele of *Dhc64C*, we performed a number of complementation tests. We found that *Su(GI)102* is lethal in combination with the deficiency *Df(3L)10H*, as shown by the absence of progeny of the genotype *Su(GI)102/Df(3L)10H* in the cross between heterozygous *Su(GI)102/TM3* and *Df(3L)10H/TM6* flies (Table II, column labeled Critical Class). This result shows that the *Su(GI)102* mutation lies within the region uncovered by the deficiency. Similar crosses were performed between the *Su(GI)102* mutation and each of the *Dhc64C* alleles, and show that *Su(GI)102* is lethal in combination with all but two of the *Dhc64C* alleles that we had isolated (Table II, column la-
Figure 4. Low (a–h) and high (a′–h′) magnification SEM micrographs documenting the genetic interaction between Dhc64C alleles and the Gl1 mutation. (a and a′) An eye from a wild-type OregonR adult fly. (b and b′) The heteroallelic combination of Dhc64C alleles Dhc64C2–3/Dhc64C5–10 produces a rough eye phenotype. (c and c′) The phenotype of the dominant mutation Gl1, in which the ommatidia appear disorganized, misshapen, and the alignment of bristles is disrupted. (d and d′) The SuGl102 mutation suppresses the Gl1 rough eye phenotype; the bristles and ommatidia in the eye are normally aligned. (e and e′) The Dhc64C5–10 allele suppresses the Gl1 rough eye phenotype, except in the most posterior region of the eye, similar to the SuGl102 mutation (d and d′). (f and f′) The third chromosome deficiency, Df(3L)10H, in combination with Gl1 does not alter the rough eye phenotype. (g and g′) The Dhc64C5–10 allele enhances the Gl1 rough eye phenotype in that the eye is smaller, has fewer ommatidia of abnormal shape and size, and duplicated or missing bristles. A similar result is obtained with the Dhc64C5–8 allele (data not shown). (h and h′) Addition of a wild-type copy of the Dhc64C transgene can rescue the enhancement of Gl1 by the Dhc64C5–10 allele. The alignment of ommatidia and bristles is less disrupted than in the Dhc64C5–10/Gl1 combination (g and g′). The same result is obtained when the Dhc64C transgene is present in Dhc64C5–8/Gl1 flies (data not shown). The genotypes of the flies represented in each panel are (a and a′) wild-type OregonR; (b and b′) Dhc64C2–3/Dhc64C5–10; (c and c′) Gl1; (d and d′) SuGl102/Gl1; (e and e′) Dhc64C5–8/Gl1; (f and f′) Df(3L)10H/Gl1; (g and g′) Dhc64C5–10/Gl1; (h and h′) P(Dhc64C5–10); Dhc64C5–8/Gl1. Crosses to generate flies of the above genotypes are described in Materials and Methods.
Table I. Summary of Eye Phenotypes in Flies Doubly Heterozygous for Glued and Dhc64C Mutations

<table>
<thead>
<tr>
<th>Genotype*</th>
<th>Eye phenotype</th>
</tr>
</thead>
<tbody>
<tr>
<td>Dhc64C*6/Gl±</td>
<td>Rough eyes, missshapen ommatidia, abnormal bristle distribution</td>
</tr>
<tr>
<td>Dhc64C*6/Gl±</td>
<td>GL rough eye is suppressed, normal hexagonally shaped ommatidia except in the most posterior of the eye</td>
</tr>
<tr>
<td>Dhc64C*6/Gl±</td>
<td>GL rough eye is enhanced, eyes are reduced in size, frequently fused ommatidia</td>
</tr>
<tr>
<td>Dhc64C*6/Gl±</td>
<td>GL rough eye is enhanced, eyes are reduced in size, frequently fused ommatidia</td>
</tr>
<tr>
<td>Dhc64C*6/Gl±</td>
<td>GL rough eye phenotype similar to the original GL rough eye phenotype</td>
</tr>
<tr>
<td>Dhc64C*6/Gl±</td>
<td>GL rough eye phenotype is neither suppressed nor enhanced</td>
</tr>
<tr>
<td>Dhc64C*6/Gl±</td>
<td>Wild-type eyes</td>
</tr>
</tbody>
</table>

*Flies doubly heterozygous for the Glued1 (Gl1) mutation and a Dhc64C mutation (Dhc64C*6, Dhc64C*8, or Dhc64C*10) were generated by crossing heterozygous virgin females of the genotype Gl1/+ with heterozygous Dhc64C*6/+ , Dhc64C*8/+ , or Dhc64C*10/+ . The small third chromosome deficiency, Df(3L)10H, removes the Dhc64C gene (Gepner, J., M.-g. Li, S. Ludmann, C. Kortas, K. Boylan, M. McGrail, and T. S. Hays, manuscript in preparation).

Table II. Suppressor of Glued Mutation, Su(Gl)102, Is Lethal in Combination with Dhc64C Mutations

<table>
<thead>
<tr>
<th>Genotypes of mated flies*</th>
<th>Critical class2</th>
<th>Sibling classes</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Su(Gl)102/TM6</td>
<td>m/TM3</td>
</tr>
<tr>
<td>mfSu(Gl)102</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Df(3L)10H/TM6</td>
<td>0</td>
<td>107</td>
</tr>
<tr>
<td>Dhc64C*6/TM6</td>
<td>0</td>
<td>104</td>
</tr>
<tr>
<td>Dhc64C*8/TM6</td>
<td>0</td>
<td>28</td>
</tr>
<tr>
<td>Dhc64C*10/TM6</td>
<td>0</td>
<td>183</td>
</tr>
<tr>
<td>Dhc64C*6/TM6</td>
<td>57a</td>
<td>75</td>
</tr>
<tr>
<td>Dhc64C*8/TM6</td>
<td>148a</td>
<td>133</td>
</tr>
</tbody>
</table>

*Males (♂) heterozygous for the deficiency Df(3L)10H or a Dhc64C mutation were mated with virgin females (♀) heterozygous for the Su(Gl)102 mutation. TM6 and TM3 refer to multiply marked balancer chromosomes which are wild type at the Dhc64C locus.

The presence of the critical class in these two crosses demonstrates that the Su(Gl)102 mutation is viable in combination with the Dhc*6 and Dhc*8 alleles.
expression of a dominant phenotype in normal development (Harte and Kankel, 1982), the ex-

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from head tissue of wild-type flies, a single 160-kD species shown that the mutation behaves as an antimorphic muta-

tion, most likely as the result of the truncated Glued gene product. This "poison product" or dominant-

completes that partial complexes containing less than a full 

function of the wild-type Glued polypeptide. Con-

st on the X chromosome; Y chromosome; +/+, maternal X chromosome pair.

At the top of each column the genotype of the progeny class is indicated. Note that female and male progeny are shown in separate columns. Because male progeny inherit their 

In our characterization of the Drosophila Glued polypep-

did not distinguish by their biochemical properties, the 

a Glue polypeptide to incorporate into the 20S Glued 

rifles with microtubules and is present in the 20S Glued 

In extracts derived from head tissue of wild-type flies, a single 160-kD species is the predominant Glued polypeptide. In GlU/+ heterozy-

flies, the truncated Glued polypeptide is more abund-

suggested by the apparent restriction of the 

of the GlU dominant phenotype to the eye. In extracts derived from head tissue of wild-type flies, a single 160-kD species is the predominant Glued polypeptide. In GlU/+ heterozy-

The dominant GlU mutation was previously shown to re-

result from the insertion of a transposable element that causes the premature termination of the Glued transcript (Swaroop et al., 1985). In GlU/+ heterozygous flies, both the wild-type transcript of 6.0 kb and a shorter transcript of 5.1 kb are detected. The shorter transcript has been pro-

in the domain of the Glued polypeptide. Since the Glued gene is required for normal development (Harte and Kankel, 1982), the expres-

functionally competes 


Although multiple Glued polypeptides are not distinguished by their biochemical properties, the functional significance of the variation in expression of each species is suggested by the apparent restriction of the 


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of the Glued polypeptide to the eye. In extracts derived from head tissue of wild-type flies, a single 160-kD species is the predominant Glued polypeptide. In GlU/+ heterozy-

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result from the insertion of a transposable element that causes the premature termination of the Glued transcript (Swaroop et al., 1985). In GlU/+ heterozygous flies, both the wild-type transcript of 6.0 kb and a shorter transcript of 5.1 kb are detected. The shorter transcript has been pro-

Table III. Su(Gl)102 Mutation is a Dhc64C Allele

<table>
<thead>
<tr>
<th>Genotypes of mated flies</th>
<th>P(Dhc+/+; m/Su(Gl)102)</th>
<th>+/Y; m/Su(Gl)102</th>
<th>P(Dhc+/+; Su(Gl)102/TM6)</th>
<th>P(Dhc+/+; Su(Gl)102/TM6)</th>
<th>P(Dhc+/+; m/TM6)</th>
<th>+/Y; m/TM6</th>
</tr>
</thead>
<tbody>
<tr>
<td>+/+; Dhc64C4-16/TM6</td>
<td>0</td>
<td>101</td>
<td>99</td>
<td>76</td>
<td>80</td>
<td></td>
</tr>
<tr>
<td>+/+; Dhc64C4-16/TM6</td>
<td>89</td>
<td>0</td>
<td>78</td>
<td>92</td>
<td>67</td>
<td>88</td>
</tr>
<tr>
<td>+/+; Dhc64C4-16/TM6</td>
<td>97</td>
<td>0</td>
<td>64</td>
<td>99</td>
<td>76</td>
<td>98</td>
</tr>
<tr>
<td>+/+; Dhc64C4-16/TM6</td>
<td>76</td>
<td>0</td>
<td>98</td>
<td>85</td>
<td>61</td>
<td>44</td>
</tr>
</tbody>
</table>

*Virgin females (♂) heterozygous for the Dhc4-16 mutation or the deficiency Df(3L)IOH were mated with males (♂) heterozygous for the Su(Gl)102 mutation. The wild-type Dhc4 transgene, P(Dhc+), is on the paternal X chromosome; Y chromosome; +/+, maternal X chromosome pair.

Downloaded from jcb.rupress.org on October 20, 2017
polypeptide to compete with the wild-type 20S Glued complex for binding sites on the dynein complex and/or microtubules. In the context of previously proposed models in which the dynactin complex acts as a cargo adaptor (Gill et al., 1991; Paschal et al., 1993; Holzbaur et al., 1994), the truncated Glued polypeptide may interfere with the native Glued complex in the docking of cellular cargoes to cytoplasmic dynein and/or microtubules.

The sedimentation behavior of the mutant Glued product demonstrates that the COOH-terminal ~20-kD domain missing from the truncated protein is required for the association of the wild-type Glued polypeptide with itself and/or the native 20S Glued complex. The transposon insertion in the GI gene is located in a region close to or within the conserved COOH-terminal stretch of ~130 residues which exhibit α-helical coiled-coil characteristics (Swaroop et al., 1987). The interruption, or absence, of this potential coiled-coil domain may prevent the mutant polypeptide from dimerizing with the wild-type Glued product, and could account for its distinct sedimentation on the gradient. Alternatively, the inability of the truncated polypeptide to incorporate into a particle with a higher sedimentation value may reflect the absence of the conserved cluster of charged residues KKEK, which lies COOH-terminal to the site of transposon insertion and would be missing in the mutant polypeptide. In vitro studies of the p150 component of dynactin from rat suggest that the homologous sequence, KKEK, mediates the binding of p150 to the centrin/ARP-1 subunit of the dynactin complex (Waterman-Storer et al., 1995).

In contrast to the potential role in dimerization of the Glued polypeptide and/or its incorporation into the Glued complex, the COOH-terminal ~20-kD domain is not strictly required for association of the Glued polypeptide with microtubules. We have shown that the truncated Glued product pellets with microtubules and elutes from microtubules with ATP-dependent, microtubule-associated proteins. These observations are consistent with studies by Waterman-Storer et al. (1995) reporting the existence of a microtubule-binding motif within the NH2-terminal 39–150 residues of the p150 subunit of dynactin from rat. However, our study does not distinguish between the direct binding of the truncated polypeptide to microtubules and an indirect association mediated by an unknown component present in the cytoplasmic extract.

Although we have not yet determined the specific molecular lesions in the dynein alleles that enhance or suppress the dominant GI eye phenotype, the genetic interactions exhibit several characteristics that provide insight into the nature of the mutations. We have shown that certain Dhc64C alleles act to suppress the rough eye phenotype, while other alleles act to enhance the rough eye phenotype of the GI mutation. The observed dominant interactions are likely not due to mutations that up-regulate Dhc64C expression, as additional copies of a wild-type Dhc64C transgene in heterozygous GI flies have no effect on the severity of the GI rough eye phenotype (data not shown). Moreover, since a deficiency that entirely removes the Dhc64C gene does not modify the severity of the GI rough eye phenotype, the suppressor and enhancer alleles do not represent complete loss-of-function alleles. In other words, the specificity of the genetic interactions between certain dynein alleles and the GI mutation suggests that the mutant dynein heavy chains retain some wild-type activity. In addition, the suppressor and enhancer Dhc64C alleles fail to produce eye phenotypes when present in combination with a deletion of the Glued gene (GI-Δ2), or in combination with other recessive lethal Glued alleles. This allele specificity indicates that the dominant interactions require the presence of both the mutant Dhc64C gene product and the truncated Glued polypeptide. The analysis of the lesions in the interacting Dhc64C alleles may help to identify domains in the dynein heavy chain involved in the association of the Glued and dynein motor complexes.

What molecular mechanism can account for the ability of certain dynein alleles to enhance the GI rough eye phenotype, while other alleles suppress the phenotype? Given the allele-specific characteristics of the genetic interactions and the truncated product encoded by the GI mutation, we favor a model in which the truncated Glued product retains the ability to associate with the dynein complex, thereby competing with the wild-type Glued complex and its attached cargo for association with dynein. The dynein alleles that suppress the dominant effect of the GI mutation would do so by acting to eliminate or reduce the defective Glued product from interfering in dynein motor function. One possibility would be that the suppressor alleles encode dynein heavy chains that incorporate into a dynein complex that can associate with the defective Glued polypeptide as well as the wild-type Glued complex, but fails to associate with cellular cargoes. The remaining wild-type dynein heavy chain and Glued polypeptides in the transheterozygote would provide sufficient levels of functional complexes to ensure proper cargo transport within the cell. In contrast, the enhancer alleles may encode dynein heavy chains that incorporate into the motor complex and retain the ability to attach with cellular cargoes, but cannot support cargo transport. The enhanced rough eye phenotype would result from the combined effects of the truncated Glued product and the defective dynein motor competing with wild-type dynein and Glued complexes for the proper association and transport of cargoes.

Regardless of the detailed mechanism of interaction, the allele specificity of the observed suppression and enhancement implies that the truncated Glued product is capable of associating with the dynein complex. Additional evidence of the physical association between the Glued complex and cytoplasmic dynein is provided by our observation that the pattern of localization of the Glued polypeptides is dependent on dynein function. We show that mutations in the Dhc64C gene that disrupt the localization of dynein result in the coincident mislocalization of the Glued polypeptides. Furthermore, other genes required for dynein localization during oogenesis, including the Bicoidal-D, egalitarian, cappucino, and spire genes, are also required for proper Glued localization. Whether the association of the dynein and Glued complexes in vivo involves the direct binding of the Glued polypeptide to the dynein heavy chain, or to another subunit of the dynein complex, is not known. However, recent in vitro evidence indicates that the 74-kD intermediate chain subunit of cytoplasmic dynein can mediate the association of dynein...

Examples of genetic interactions between genes whose products interact in the formation of complex structures or in common cellular processes have been reported in a number of organisms. In Chlamydomonas, extragenic suppressors of paralyzed flagellar mutations have identified loci that alter the outer and inner dynein arm structures (Huang et al., 1982; Porter et al., 1992; 1994), as well as regulatory components (Huang et al., 1982; Piperno et al., 1992, 1994; Gardner et al., 1994) of the flagellar axoneme. Similarly, in Aspergillus nidulans, extragenic suppressors of mutations in the β tubulin gene, benA, have identified α tubulin (Morris et al., 1979) and γ tubulin genes (Oakley and Oakley, 1989). In other examples, dominant enhancer, or second-site noncomplementing, mutations have revealed structural interactions between gene products. For example, mutations in the Caenorhabditis elegans collagen gene sqt-1 (Kramer et al., 1988) show unusual dominant interactions with mutations in other loci, causing defects in cuticle structure and body shape (Kuschk and Edgar, 1986). Like sqt-1, these loci might encode other collagens or components of the worm cuticle. In addition, mutations in the α tubulin genes of S. cerevisiae (Stearns and Botstein, 1988) and Drosophila (Hays et al., 1989) have been identified by their failure to complement mutant alleles of β tubulin genes. As in the examples cited above, the dominant allele-specific interactions between the Drosophila Dhc64C and Glued mutants reflect a functional interaction between the Glued and dynein complexes.

The eye phenotypes observed for the dominant Gl1 mutation and certain intragenic combinations of Dhc64C alleles, as well as the enhancement and suppression of the Gl1 rough-eyed phenotype by specific Dhc64C alleles, make clear the necessity for dynein and Glued function during Drosophila eye development. Previous studies have described the effect of Gl1 on the projection of axons from the retina to the optic tectum in adult flies (Meyerowitz and Kankel, 1978). This defect could reflect the disruption of dynein function in retrograde axonal transport. Perhaps dynein transport is required for the delivery of a signal from the optic tectum back to the retinal cell body and nucleus that indicates formation of the proper synaptic connection. Alternatively, the defective projection of retinal axons could result from an earlier defect in dynein or Glued function during the development of the retinal epithelium. Differentiation of the eye disc epithelium is accompanied by a highly orchestrated series of events including the synchronization of cell cycles and divisions, cell shape changes, cell-specific patterns of nuclear migration, and the formation of the founder cell clusters that give rise to the ommatidia. Renfranz and Benzer (1989) have described an apparent failure to complete and/or maintain the ommatidial clusters of cells in Gl1 eye imaginal discs in the third larval instar. Additional analysis will be required to reveal the underlying basis of the rough eye phenotype caused by the dynein and Glued mutants.

In addition to the eye phenotypes described above, certain alleles of both the Glued (Harte and Kankel, 1982) and Dhc64C (Gepner, J., M.-g. Li, S. Ludmann, C. Kortas, K. Boylan, M. McGrail, and T. S. Hays, manuscript in preparation) genes exhibit female-sterile phenotypes. Together with our observations that dynein and Glued colocalize during oogenesis and share similar genetic requirements for localization, the female-sterile phenotypes indicate a role for dynein and Glued in the development of the egg. There is growing evidence implicating microtubule motors and cytoskeletal components in the specification and differentiation of the oocyte (Cooley and Theurkauf, 1994). We are currently conducting genetic screens for additional second-site modifiers of Dhc64C and Glued mutations to identify other gene products that may provide a link between dynein motor function and the mechanisms that regulate specification and differentiation of cell fate.

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