Commentary

A Cell Biological Perspective on Genome Research

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The genomes of several organisms are currently being sequenced at impressive rates, but some scientists have asked whether the final products will be worth the effort. Here we argue from both the history of structural biology and some encouraging initial results that the answer is a resounding yes.

There have been five overlapping stages in the growth of structural biology. The ancients learned much about animals and plants through comparative anatomy. A systematic study of animal anatomy through dissection began in the 16th century. Microscopic anatomy began in the mid-17th century, and by the early 1900’s there were detailed descriptions of a wide diversity of organs and organisms. 1940–1970 saw a comparable flowering of knowledge about cellular fine structure and macromolecular assembly, thanks in part to the electron microscope. The details of macromolecular structure were, however, largely invisible without crystallography, and the fruits of this technology are still ripening.

Each of the above stages in the development of “anatomy” has depended on a new method or approach whose availability led to rapid growth in structural information at a novel level of size or detail. An analogous situation now pertains to determination of DNA sequence. While the technology for sequencing proteins and nucleic acids has been available for decades, recent improvements have made it possible to learn the primary structure of DNA comparatively quickly. As a result, there has been a huge increase in the number of labs that are sequencing DNA and an almost incredible increase in the rate of sequence acquisition. The availability of computers fast enough to deal expeditiously with all the resulting data has permitted efficient analysis and comparisons of DNA sequences and of the polypeptides predicted from them, so the flow of information from acquisition to utilization for biological comparisons has become both rapid and objective. The resulting catalogues of genes and gene products are facilitating a new version of all the discriminations that have traditionally given context and meaning to sequence data; and genetic and molecular information to give context and meaning to sequence data; and (2) genome work on each organism has begun with the generation of physical maps, so sequence information can be related to genetic maps and studies of mutant phenotype.

One of the first major sequencing efforts has been with the yeast Saccharomyces cerevisiae, whose genome was known to be small (~12.5 megabases [Mb]). Many yeast genes had already been identified by mutation, and several regions of this genome had been analyzed by saturation mutagenesis, which identifies most genetically recognizable loci. The frequency and size of intervening and spacer DNA sequences were known to be small, and physical maps of the genome had already been constructed. There were also methods for manipulating this genome experimentally, e.g., gene replacement by homologous recombination, so one could anticipate that sequence information could be related to the functional consequences of mutation. When systematic sequencing began, the 16 S. cerevisiae chromosomes were undertaken by several yeast labs throughout the world. As of October 1995, over three-quarters of the

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Table I. World Wide Web Addresses for Information about Specific Genome Projects

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<tr>
<th>Genome Project</th>
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<td>S. cerevisiae</td>
<td><a href="http://genome.wustl.edu/gsc/gschmpg.html">http://genome.wustl.edu/gsc/gschmpg.html</a></td>
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<td><a href="http://www.nchgr.nih.gov">http://www.nchgr.nih.gov</a> (National Center for Human Genome Research)</td>
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<td><a href="http://www.ncbi.nlm.nih.gov">http://www.ncbi.nlm.nih.gov</a> (This includes GenBank and the NCBI Newsletter)</td>
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<td><a href="http://gdbwww.gdb.org/omim/docs/omintop.html">http://gdbwww.gdb.org/omim/docs/omintop.html</a></td>
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Genome research on more complex model organisms has again begun with the formation of detailed physical maps and limited genomic sequencing. For the nematode, Caenorhabditis elegans (genome size ~100 Mb), a collection of “cosmid” clones has been constructed, each of which carries tens of kilobases (kb) of exogenous DNA. Many of these sequences are from contiguous loci, yielding an ordered library that covers much of the genome. There is now a C. elegans genomic library in yeast artificial chromosomes (YACS) as well; each of these clones contains hundreds of kb of DNA, so the number needed to cover a larger genome is manageable (8). Meanwhile, DNA sequencing has been going on in two centers, and the rate of new finished sequence (six determinations to minimize the chances of error) now exceeds one cosmid/day (7). Completion of this genome is expected by the end of 1996 (Table 1).

For model organisms with larger genomes, like fruit flies (genome 1.5 × that of C. elegans) and mice (30 × the nematode), the above strategy is being employed in an expanded form. Both YAC and cosmid libraries are now available to help map genes to specific pieces of DNA. Indeed, much of the recent work on the human genome has been focused not on DNA sequencing, but on detailed chromosomal mapping. Microsatellites (short segments of repetitive DNA) are being used to identify unique molecular markers with better than 1 centimorgan resolution, and physical maps are being generated with sequence-tagged sites (STSs) on YAC and cosmid clones. This work has significantly refined results from the classic methods of cytogenetics, like Q or G banding, as well as older molecular mapping methods, like restriction fragment length polymorphisms. The result is both a useful map of genes related to human diseases and tools that will be necessary for molecular studies of particular chromosomal regions. Systematic sequencing of the human genome is just now getting underway (Table 1).

Sequence Data Are Telling Us Surprising Things about Genome Structures and Functions

Information from S. cerevisiae and C. elegans is already telling us interesting things about genome structure and organization. In addition to the obvious features, like the number of genes in a given amount of DNA, the statistics of gene size, and the distribution of genes among major families defined by sequence, things are coming to light that few would have predicted. The sequences from regions subjected to saturation mutagenesis have revealed many more genes than were found in searches for mutants with phenotypes. For example, genetic estimates predicted ~3,000 essential genes in C. elegans (3), but estimates from genome sequence suggest ~13,000 (10). Presumably, the inactivation of the additional genes by mutation is either irrelevant for growth as we assess it in the laboratory or is “covered” by the functions of other, unmutated genes. The work has also revealed novel features of genome organization: there are genes within introns, genes that appear to encode giant proteins, and clusters of genes with unknown functions (12). While more than half the genes so far discovered by sequencing are of unknown function, ~40% of the genes from C. elegans (7), ~55% from S. cerevisiae (11), and 58% from the prokaryote, H. influenzae (6) encode proteins whose sequences are similar to the products of genes already identified in other organisms. Clearly, there is much still to do in tracking down and analyzing unknown genes, but it is encouraging that so much sequence information can already be understood in some functional context and related to processes that are under study elsewhere.

Large scale studies of gene sequences have also opened up a new approach to genetic comparisons. Many genes from humans and other organisms have been partially characterized by sequencing only a small part of a cDNA to yield an “expressed sequence tag” (EST) (1). These have

1. Abbreviations used in this paper: EST, expressed sequence tag; Mb, megabase; YAC, yeast artificial chromosome.
been organized in data bases of ESTs (dbEST), and the catalogues of these sequences are expanding by several hundred sequences/day (Table I). When the sequences of genes identified by mutation are compared with those in the dbESTs, it has sometimes been possible to find a significant similarity, thereby identifying a mammalian gene that corresponds to a gene already characterized in a simpler organism, like a yeast (9). Homologous sequences have now been identified in model organisms for most of the ~50 human diseases genes that have been mapped by positional cloning (1, 2, 4, and Table I). There are two important consequences of these developments: (a) the detailed analysis of gene structure and function, which is comparatively cheap and efficient in model organisms, is likely to be of direct benefit in understanding complex processes and pathologies in higher eukaryotes; and (b) it will be possible to study many human diseases in organisms whose experimental manipulation is not ethically offensive.

The Conservation of Some Genes That Control Complex Functions Is Strong Enough That Currently Mysterious Aspects of Organismic Biology Are Likely to Profit from Genome Work

The homeotic loci of Drosophila are widely known to play key roles in the hierarchical regulation of gene expression. Studies of homologous sequences in organisms with apparently different body plans have more recently suggested that genome structure is related to the processes that regulate development. For example, the genomic order of the homeotic "HOX" genes in several vertebrates is correlated with both the temporal and the positional order of their expression (for a review see reference 5). This case suggests that there may be important properties of genomic organization that are still beyond our grasp. While sequence information is unlikely to be sufficient for elucidating these properties, it is almost certainly a prerequisite. There may be a parallel between the way information about specific genes is currently linking yeast and mammalian cell biology, and the way analogous information about higher order genome structure will illuminate developmental biology.

The Investment in Genome Structure by Labs That Are Set Up to Sequence Efficiently May Expedite Cell Biological Research and Save Money

Many cell biologists who began their careers as microscopists or biochemists are now engaged in "molecular biology," meaning that they are cloning and sequencing the genes that encode proteins essential for the cellular processes they study. This makes sense because we have all witnessed the power of recombinant DNA technology for expediting research on proteins that are present in amounts too small for practical biochemistry. Moreover, the use of antisense RNA, gene disruptions, and transfection with mutant DNA encoding proteins with dominant negative phenotypes are now important companions to antibody perturbations and pharmacology for the study of complex processes in organisms without powerful genetics. The result is that "cell biology" labs all over the world are now spending significant time and resources cloning, mapping, sequencing, and mutating DNA, when what they want to be doing (and do best) is studying the behavior of gene products. This work is often called "reagent construction," and it is now common to spend years assembling the DNA clones, mutants, protein fragments, and antibodies necessary to do a key cell biological experiment. For students of cell biology, it is almost a modern Rite of Passage to get a clone and sequence it, etc., in preparation for the originally intended experiment. Lamentably, this is mostly mundane work in which daily tasks bear little relation to the biological problems selected for study. When one sees bright and energetic cell biologists dulling their enthusiasm on kilobases of DNA sequence in order to get the groundwork information necessary to make the tools they need, one can't help but think that there must be a better way.

Taken together these observations have made us supporters of genome research. The investment of a reasonable fraction of our community's resources in accumulating information about DNA sequence and organization is likely to pay big dividends, not only in knowledge of genome structure but in the saving of countless hours of molecular drudge work. A well set up sequencing lab can produce ~2 kb of finished sequence per worker per day, while labs that are sequencing by hand are doing well if they get 1/10th that amount. Moreover, the cost per base drops by ~10-fold when the work is done in a lab with proper equipment and experience. The current rate of sequence collection will probably increase as technology improves, implying that we should continue with significant investments in the study of genome structure for the near future. While those of us doing experiments may resent the expenditure of money that might have helped us to stay funded, the resulting information will almost certainly be worth the investment, and the approach will probably save money.

Even with All This Success, There Are Problems That Must be Addressed by Our Scientific Community

Enthusiasm for genome research does, however, lead to three problems that our community must address: (1) since resources are limited, the dollars spent on genome research must be consistent with our current funding constraints. The 1995 investment in extramural research by the National Center for Human Genome Research (NCHGR) was ~$114 million, while the Departments of Energy and of Agriculture spent ~$69.5 and $3.8 million, respectively, on genome work. The Small Business Administration spent ~$3 million, and comparatively small sums that are hard to determine came from other Institutes at the National Institutes of Health (NIH) and from the National Science Foundation. These investments in aggregate are small relative to the whole NIH budget (~$12 billion for 1995), but they are significant compared with the budget of the National Institute for General Medical Sciences (just over $900 million), the agency upon which many of us rely. Given the importance of genome work and given the investments by other institutes and agencies in basic biomedical research, broadly defined, the current balance seems to us reasonable, but a greater investment in genome work would tip the balance.

(2) The results from sequencing are first available to the sequencers themselves. With genome labs funded by in-

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The free and rapid flow of sequence information into the public domain is critical to avoid wasting or improperly using sequencing data. The NCHGR has stipulated that sequence data from the labs they fund must be made public within 6 months, but some labs in foreign countries are slower. Efforts should be made to minimize the time to public access to maximize the rate of good science. Profits from sequences and restrictions on their use by individuals and companies is a problematic trend that will take years to sort out in the courts. In the interim, we must minimize the chances of profit motives retarding scientific progress.

In the future, sequencing labs will certainly provide important information, such as the genome structures for model organisms, humans, and important pathogens, as well as the mapping of human disease genes. However, sequencing labs may become entrenched, so their lifetimes exceed their utility. It is impossible to say when enough genome work will have been done, but there is an obvious analogy with a learning curve. Sequencing centers may want to go from human to aardvark, caiman, and dodo, expanding our knowledge of evolution, phylogeny, and extinction. The community of biologists will have to recognize when new knowledge from new sequences has begun to plateau and limit their spending. Sequencing programs must be structured so they can shrink as their value diminishes, giving way to more experimentally oriented biology.

That said, it is clear that future biologists will be working in an environment defined by a wondrous wealth of information about genome structure. It is mind-boggling to think of how our experimental lives will be changed as a result. No field of biology will be untouched, and a whole new generation of experimental approaches will likely emerge to justify the investments that are now being made in sequence acquisition and analysis.

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References