Blood Platelets Are Assembled Principally at the Ends of Proplatelet Processes Produced by Differentiated Megakaryocytes

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Abstract. Megakaryocytes release mature platelets in a complex process. Platelets are known to be released from intermediate structures, designated proplatelets, which are long, tubelike extensions of the megakaryocyte cytoplasm. We have resolved the ultrastructure of the megakaryocyte cytoskeleton at specific stages of proplatelet morphogenesis and correlated these structures with cytoplasmic remodeling events defined by video microscopy. Platelet production begins with the extension of large pseudopodia that use unique cortical bundles of microtubules to elongate and form thin proplatelet processes with bulbous ends; these contain a peripheral bundle of microtubules that loops upon itself and forms a teardrop-shaped structure. Contrary to prior observations and assumptions, time-lapse microscopy reveals proplatelet processes to be extremely dynamic structures that interconvert reversibly between spread and tubular forms. Microtubule coils similar to those observed in blood platelets are detected only at the ends of proplatelets and not within the platelet-sized beads found along the length of proplatelet extensions. Growth and extension of proplatelet processes is associated with repeated bending and bifurcation, which results in considerable amplification of free ends. These aspects are inhibited by cytochalasin B and, therefore, are dependent on actin. We propose that mature platelets are assembled de novo and released only at the ends of proplatelets, and that the complex bending and branching observed during proplatelet morphogenesis represents an elegant mechanism to increase the numbers of proplatelet ends.

Key words: blood platelet • megakaryocyte • microtubules • actin cytoskeleton • thrombopoiesis

Blood platelets are small cells that lack a nucleus, but have a highly organized cytoskeleton, unique receptors, and specialized secretory granules (for review see Zucker-Franklin, 1997). Nearly a trillion platelets circulate in an adult human and respond to blood vessel injury by changing shape, secreting granule contents, and aggregating. These responses are advantageous for hemostasis, but adverse when they cause tissue ischemia or infarction. Thus, the terminal differentiation of mammalian megakaryocytes into platelets represents a unique problem in cell biology with great relevance for human health. Each mature megakaryocyte produces and releases hundreds of platelets into the circulation (Kaufman et al., 1965; Harker and Finch, 1969; Trowbridge et al., 1984), but the mechanisms underlying this remarkable culmination of cell differentiation are poorly understood, in part because of the extreme rarity of megakaryocytes in normal bone marrow (<0.1% of cells).

Over the years, many studies on specific aspects of platelet biogenesis have provided important insights and led to testable hypotheses. Megakaryocytes are polyploid cells whose size and DNA content correlate directly with the circulating platelet mass (Ebbe and Stohlman, 1965; Paulus, 1970). The cytoplasm of mature megakaryocytes is large, replete with platelet-specific granules, and harbors a unique and extensive system of internal membranes. A cording to one model of thrombopoiesis, these internal membranes demarcate fields or territories of prepackaged platelets, and fragmentation of the cytoplasm releases these platelets (Yamada, 1957). A n alternative flow model incorporates the observation that mature megakaryocytes extrude long cytoplasmic extensions, called proplatelets, that contain swellings encompassing platelet organelles (Becker and DeBruyn, 1976). This model makes two dis-
distinct proposals: (1) that platelets assemble de novo within proplatelets, and (2) that the system of internal membranes is not simply a partition, but rather it serves as a reservoir for the plasma membranes of future blood platelets (Radjay and Haller, 1982). Recognizing that circulating platelets have a rich and unique cytoskeletal architecture is at the heart of the distinction between these alternative models because the problem of platelet genesis is not simply that of packaging cytoplasmic organelles. Rather, any model of thrombopoiesis requires elucidation of the mechanism for assembly and organization of a complex cytoskeleton and membrane skeleton. In this light, it is important to note that a developed platelet cytoskeleton is not readily apparent within the cytoplasm of terminally differentiated megakaryocytes.

The ability to expand and culture megakaryocytes in vitro in the presence of the growth factor thrombopoietin has greatly advanced the opportunity to study these processes in detail (reviewed in Kauschansky, 1995). Cultured human and murine megakaryocytes terminate differentiation by producing large numbers of proplatelets (Choi et al., 1995; Cramer et al., 1997; Leclerc et al., 1998). Initial studies that exploited this opportunity established that proplatelets express features of circulating blood platelets and outlined features of the proplatelet cytoskeleton (Choi et al., 1995; Cramer et al., 1997). This work has led investigators to consider proplatelet formation to be the physiologic basis of platelet release, and imparts renewed urgency to obtain a fuller understanding of the underlying mechanisms. Building on these initial studies, we report newly recognized aspects of proplatelet morphogenesis and provide detailed characterization of the structural basis of selected key features.

Materials and Methods

Mega-karyocyte Cultures and Enrichment of Platelet-sized Particles

Livers were recovered from mouse fetuses and single cell suspensions were generated using methods described previously (Leclerc et al., 1998). Between the fourth and sixth day of megakaryocyte culture, cells were placed on a 1.5-3.0% albumin step gradient and sedimented (Drachman et al., 1997) to obtain enriched populations of megakaryocytes. Platelet-sized particles were isolated from culture supernatants by centrifugation at 500 g and, when necessary, were washed using a metrizamide step gradient (Falet et al., 1998).

Video Microscopy

Isolexed megakaryocytes were diluted into semi-solid medium containing 65% Leibovitz L-15 medium (GIBCO BRL) and 35% MethoCult (M-3230; Stem Cell Technologies). Megakaryocytes in suspension were pipetted into chambers formed by mounting a glass coverslip coated with 3% BSA onto a 10-mm petri dish with a 1-cm hole. Preparations were maintained at 37°C. Megakaryocytes in suspension were pipetted into chambers formed by mounting a glass coverslip coated with 3% BSA onto a 10-mm petri dish with a 1-cm hole. Preparations were maintained at 37°C using a BiotomeX microplate shaker (BiotomeX Corp.), and examined on a Zeiss IM-35 inverted microscope equipped with a phase-contrast long working distance condenser. Images were obtained using a Hamamatsu charged coupled device (CCD) camera and a frame grabber. Movies were generated using NIH-I mage 1.61 software.

Electron Microscopy

Samples were prepared for electron microscopy as described (Hartwig, 1992). In brief, megakaryocytes in suspension were placed in wells of a 96-well microtiter plate, each containing a poly-L-lysine-coated coverslip, and the plate was centrifuged at 500 g for 5 min at 37°C. Some cells were fixed with 3% glutaraldehyde in PBS to study their surface topography. Cytoskeletons were isolated by permeabilizing cells with PHEM buffer (60 mM Pipes, 25 mM Hepes, 10 mM EGTA, and 2 mM MgCl₂) containing 0.75% Triton X-100, 2 μM phallicadin, 20 μM taxol, and protease inhibitors for 2 min. Cytoskeletons were washed with PHEM buffer containing 0.1 μM phallicadin and 30 μM taxol, and fixed in PHEM buffer containing 1% glutaraldehyde for 10 min. To examine the membrane skeleton, cells were incubated with permeabilization buffer containing 0.1% glutaraldehyde for 2 min and fixed as described above. Coverslips of fixed cells and cytoskeletons were rapidly frozen, freeze-dried, and coated with tantalum-tungsten and carbon. Replicas were picked up on carbon-formvar-coated copper grids and examined with a J EOL J E M -1200 EX electron microscope at an accelerating voltage of 100 kV.

Immunofluorescence

Antibodies. mAbs specific for β-tubulin were obtained from A. mesham. Secondary antibodies, FITC, and TRITC conjugates of goat anti-rabbit and goat anti mouse IgGs were obtained from Sigma Immunochemical. Primary antibodies were used at 5 μg/ml in PBS + 1% BSA and secondary antibodies at 1:200 dilution in the same buffer.

Confocal Microscopy. Megakaryocytes were fixed with 4% formaldehyde in Hanks’ balanced salt solution (GIBCO BRL) for 20 min, cytocentrifuged at 500 g for 4 min onto coverslips previously coated with poly-L-lysine–coated coverslips previously coated with poly-L-lysine–coated coverslips previously coated with poly-L-lysine–coated coverslips previously coated with poly-L-lysine–coated coverslips previously coated with poly-L-lysine–coated coverslips previously coated with poly-L-lysine–coated coverslips previously coated with poly-L-lysine–coated coverslips previously coated with poly-L-lysine–coated coverslips previously coated with poly-L-lysine. Megakaryocytes were permeabilized with 0.5% Triton X-100 in Hanks’ containing 0.1 mM EGTA, and blocked with 0.5% BSA in PBS. The specimens were incubated in primary antibody for 3-6 h, washed, treated with the appropriate secondary antibody for 1 h, and washed extensively. Controls were processed identically except for omission of the primary antibody. Preparations were examined with a BioRad MRC 1024 laser scanning confocal microscope equipped with Lasersharp 3.1 software. Images were obtained using a Zeiss A xiov 1000 equipped with a 100× differential interference contrast (DIC) oil immersion objective (NA 1.4).

Flow Cytometry Studies

A fluorescein-phalloidin-based flow cytometric assay was employed to quantitate the actin filament content in blood and culture-derived platelets (Howard and Osesoj, 1985). Platelets treated with and without 1 U/ml mouse thrombin (Sigma Chemical Co.) were fixed by the addition of an equal volume of 4% paraformaldehyde in platelet buffer (145 mM NaCl, 10 mM Hepes, 10 mM glucose, 0.5 mM Na₂HPO₄, 5 mM KCl, 2 mM MgCl₂, and 0.3% BSA, pH 7.4) for 30 min at 37°C. Fixed platelets were permeabilized with 0.1% Triton X-100, labeled with 10 μM FITC-phalloidin for 1 h at room temperature, and washed twice with PBS. Samples were analyzed in a Becton Dickinson FACscan, gated for platelets, and a total of 10,000 events were analyzed using Lysys II software (Becton Dickinson). Experiments were repeated in triplicate and the results were averaged.

Cytoskeletal-disrupting Agents

Taxol, cytochalasin B, and nocodazole were obtained from Sigma Chemical Co.

Image Preparation

All images were imported into either Adobe Photoshop or Claris Draw and printed on a Kodak 8650 PS color printer.

Online Supplemental Material

The manuscript includes three movies supplied in Quick Time™ format.

Figure 1: Proplatelet formation contains 58 frames taken at 10-min intervals using a 40× phase-contrast objective. This movie shows a megakaryocyte undergoing proplatelet formation and demonstrates the dynamic behavior of proplatelets.

Figure 2: (a) Dynamic interconversion contains 30 frames taken at 10-min intervals and shows a higher magnification view of the reversible interconversion between spread lamellar segments and condensed proplatelet forms. (b) Bending/branching contains 10 frames taken at 10-min intervals and shows proplatelet bending and the bifurcation at high magnification. Videos are available at http://www.jcb.org/cgi/content/full/147/6/1299/D.C1.
Results

Proplatelet Intermediates Are Remarkably Dynamic Structures

To date, proplatelets have been studied and described largely through static images, and most conclusions about mechanisms of proplatelet formation are inferred from studies conducted with only limited benefit of time-lapse cinematography. We used video-enhanced light microscopy of cultured mouse megakaryocytes to reveal a spectrum of highly reproducible morphogenetic changes and cellular movements during formation of proplatelets (Fig. 1 and accompanying video). A transformation of the entire megakaryocyte cytoplasm results in condensation of cell material into platelet-sized particles, which have the appearance of beads linked by thin cytoplasmic bridges.

This process unfolds over 4–10 h, and initiates with erosion of one pole of the megakaryocyte cytoplasm. This generates a unique pseudopodial structure that elongates to yield slender tubules of uniform diameter of 2–4 μm; these in turn develop periodic densities along their length that impart the characteristic beaded appearance of proplatelets. Maturation of proplatelets ends in a rapid retraction that separates a variable portion of the proplatelets from the residual cell body, according to a process that presumably mirrors platelet release in vivo.

High resolution time-lapse analysis reveals several striking features that have not been recognized previously, and emphasizes the highly dynamic nature of thrombopoiesis. Remodeling of the megakaryocyte cytoplasm is accompanied by centrifugal spreading that increases the apparent cell surface area and, measured from representative cells

Figure 1. Video-enhanced light microscopy of a terminally differentiated mouse megakaryocyte forming proplatelets in vitro. During the initial stages of proplatelet formation, the megakaryocyte spreads and its cortical cytoplasm begins to unravel at one pole (this zone of erosion is labeled with an asterisk in the first panel). As the cell spreads, the cytoplasm at the erosion site is remodeled into large pseudopodia (white arrow at 2 h) that elongate and become thinner over time, forming narrow tubes of 1–4 μm diam. The proplatelet extensions frequently bend (white arrowhead at 2 h), and it is at sites of bending that the tube bifurcates to generate a new process. In this manner, the entire cytoplasmic space of the megakaryocyte is converted into anastomosing proplatelet extensions. Proplatelets also develop segmented constrictions along their length that impart a beaded appearance. The process of proplatelet elaboration culminates in a rapid retraction that separates the many strands of proplatelets from a residual nuclear mass (asterisk in last panel). Note the crawling of small cytoplasmic fragments (5–10 μm) at the ends of some of the proplatelet extensions (arrowheads at 4–7 h). Crawling begins when the end of the proplatelet adheres and flattens, forms a leading lamellipodia, and migrates away from the cell center, dragging a trail of proplatelets behind. Bar, 20 μm.
Portions of the proplatelet cytoplasm exhibit striking and dynamic interconversion between spread lamellar segments and condensed structures resembling platelets; this oscillation between proplatelet and spread morphologies may occur multiple times (Fig. 2 a and accompanying video). The distal ends of proplatelet processes periodically flatten to form fan-shaped sheets resembling lamellipodia and crawl away from the cell center, dragging a proplatelet trail (Fig. 1, arrowheads at 4–7 h). Interestingly, proplatelet fragments and the megakaryocyte cell body move at similar speeds (0.2–0.4 μm/min), and spreading movements are inhibited by 1–10 μM of cytochalasin B, suggesting that they require actin polymerization; upon removal of this drug, megakaryocytes recover the ability to spread on the surface (data not shown). The stable cytoskeletal architecture of mature blood platelets would seem to preclude the degree of dynamic cell motility revealed by these studies. Hence, platelet assembly must at best be incomplete during this phase of proplatelet morphogenesis, and platelets must be assembled de novo in the course of this transition; structural studies detailed below strongly support this conclusion.

To better understand the dynamic aspects of proplatelet morphogenesis, we examined selected aspects at high resolution. Branching of proplatelets has been recognized previously (Haller and Radley, 1983) but not investigated in detail. Proplatelet bifurcation occurs at a variable distance from the cell body, but only at sites of pronounced bending (Fig. 2 b and accompanying video). The sharp kinks elongate, forming a small loop, and a new proplatelet process formed by this loop extends from the original nexus. Newly generated divisions also develop the periodic constrictions characteristic of proplatelets. Platelet-sized particles and nodules of variable size translocate extensively along megakaryocyte processes (Fig. 2 c), another feature that is incompatible with preassembly of a stable platelet cytoskeleton. Distinct segments move in either direction at an estimated rate of ~1 μm/min, may move in opposite directions on a single extension, and can even reverse direction. The particles typically do not reveal changes in phase density as they move; however, we have observed several platelet-sized particles...
fuse together during translocation (Fig. 2 c, 20-min frame). Additionally, proplatelets undergo continuous cycles of extension and retraction that are most clearly evident in the video accompanying Fig. 1. Thus, these observations reveal proplatelet processes to be inherently unstable structures with a capacity for extensive morphogenetic changes.

The surprisingly dynamic character of proplatelet formation raises many important questions. (1) What is the structure of the intermediate, platelet-like particles detected within developing proplatelets? (2) Which cellular structures drive megakaryocyte spreading, elongation, and thinning of proplatelets? (3) What is the structural basis for the transition between proplatelets and mature platelets? (4) How does branching of proplatelet processes occur, and what is its significance? In the following experiments, we address some of these questions. Previous studies investigating platelet formation by cultured megakaryocytes have only partially assessed the structure and function of the released particles (R adley et al., 1991; Choi et al., 1995; Cramer et al., 1997). Therefore, we first investigated whether the shape changes and movements observed in our studies lead to the release of verifiable platelets.

**Culture-derived Platelets Produced via Proplatelets Manifest Structural and Functional Features of Circulating Blood Platelets**

Platelet-sized particles released by cultured mouse megakaryocytes exhibit each of the hallmark features of blood-derived platelets. They are disc-shaped, 2–3 μm in diameter, lack discernible membrane topology or protrusions (Fig. 3 a), and show periodic surface invaginations (Zucker-Franklin, 1970) that demarcate entrances into the open canalicular system (Fig. 3 a, arrowheads). Removal of the plasma membrane exposes an elaborate membrane skeleton composed of 3–5-nm strands identical in appearance to the spectrin-based network of human platelets (Fig. 3 b), and a prominent marginal band (H aydon and Taylor, 1965; W hite and K rivit, 1967; K enney and L inck, 1985) derived from the apparent coiling of a single microtubule (Fig. 3 c). A complex three-dimensional network of actin filaments, similar to that observed in mature blood platelets (Fox et al., 1988; N achmias and Y oshida, 1988; H artwig and D esisto, 1991; Fox et al., 1996), is also found beneath the membrane skeleton (Fig. 3 d). 200–225 platelet-sized microtubule coils may be seen extending from typical proplatelet-producing cells (see Fig. 5, g and h), a value that is in good agreement with the proposed theoretical range of platelets produced per megakaryocyte (K aulman et al., 1965; H arker and F inch, 1969; T rowbridge et al., 1984; S tenberg and L evin, 1989) and with observations in cultured human cells (Choi et al., 1995). Furthermore, the released particles respond to stimulation with thrombin by extending lamellipodia and filopodia (Fig. 3 e), forming microaggregates (Fig. 3 f), and exhibiting a twofold increase in their proportion of filamentous actin (data not shown), which is similar to observations in activated blood platelets (Carlsson et al., 1979; F ox and P hilips, 1981; F ox et al., 1984; K arlsson et al., 1984). These particles showed increased expression of the activation-dependent antigens P-selectin and the functional fibrinogen receptor (data not shown), as previously reported for human proplatelets (Choi et al., 1995). Hence, platelets released in cell culture display all the morphological and functional criteria that distinguish blood platelets.

**Changes in the Microtubule Cytoskeleton during Proplatelet Morphogenesis**

Microtubule assembly has been previously implicated in platelet formation (H andagama et al., 1987; R adley and H atshorn, 1987; T ablin et al., 1990), and our studies confirm its essential role in the progression of initial pseudopodia into proplatelets. The microtubule disrupting agent nocodazole (1–10 μM) completely inhibits formation of all megakaryocyte projections (Tablin et al., 1990). When nocodazole is added to megakaryocytes after they have formed proplatelet processes, particle transport is completely blocked (data not shown), indicating the requirement for a microtubule-based motile apparatus in this process. To understand the structural basis of the transitions between the megakaryocyte cell body, pseudopodia, proplatelet processes, and nascent platelets, we examined the microtubule cytoskeletons of proplatelet-producing megakaryocytes. Early in the maturation process, before spreading and erosion of the cell body begin, the megakaryocyte cytoplasm is replete with long individual arrays of microtubules that cluster around the nucleus and radiate toward the cell margins (Fig. 4, a and b). A dense large and blunt pseudopodia are formed in the area of erosion, the microtubules consolidate into cortical bundles situated just beneath the membrane surface of the protrusion (Fig. 4 c). As the pseudopodia extend and become thinner, they display a prominent band of microtubules along their edges; in some cases, these bundles are seen to curl or spiral inside the pseudopodia (Fig. 4 d). The next recognizable step is the conversion of the blunt pseudopodia, via elongation, into cytoplasmic extensions that continue to harbor thick bundles of microtubules (Fig. 5 a). These processes always have bulbous ends, and electron microscopy reveals a microtubule bundle that loops just beneath the plasma membrane and reenters the shaft to form a teardrop-shaped structure (Fig. 5, b and c).

**De Novo Assembly of Platelet Particles at Proplatelet Termi**

Determining when and where platelet units are finally assembled during megakaryocyte differentiation has been controversial (R adley and H atshorn, 1987). At the heart of this question is understanding where and how the marginal microtubule band, a stable feature of terminally differentiated platelets (W hite, 1968; R adley and H atshorn, 1987), is formed. If platelets preassemble within the megakaryocyte cytoplasm into platelet fields or territories, then microtubule coils would be expected to lie within these areas before cytoplasmic fragmentation. The dynamic process of proplatelet formation (Figs. 1 and 2) and the underlying progression of the microtubule cytoskeleton from pseudopodia to proplatelets (Fig. 4) argue strongly against this possibility. In addition, in scanning hundreds of antitubulin immunofluorescence profiles of megakaryocytes, we failed to identify microtubule coils within the cell bodies;
instead, these are consistently and readily detected only in proplatelet extensions (Fig. 3, g and h).

Most released platelet-sized particles remain connected by cytoplasmic bridges, the most abundant being barbell forms composed of two platelet-like particles joined by one cytoplasmic strand. To ask whether the bulbous ends and thickenings along the shaft of proplatelets show mature cytoskeletal features, we examined representative released particles by antitubulin immunofluorescence confocal microscopy (Fig. 5 d) and electron microscopy (Fig. 5, e and f). A microtubule bundle forms the core of the proplatelet shaft, and the ends have microtubule bundles forming teardrop-shaped loops that are similar in both size (1-3 μm) and appearance to those described in mature blood platelets (White, 1968). Distorted microtubule bundles are also visible in platelet-sized swellings along the shaft of the proplatelet; however, in contrast to the terminal rings, those observed along the proplatelet shafts are uniformly less distinct (Fig. 6). Whereas these platelet-sized nodules superficially appear to include microtubule rings, close inspection reveals that they are actually points where microtubule bundles simply diverge for a short distance but fail to form the mature teardrop shape (Fig. 6, c-g). These nodules consistently reveal membrane and other detergent-insoluble materials, presumably destined for final platelet assembly elsewhere (compare Fig. 6, a and b). These results are entirely consistent with the dynamic activity that is observed at the sites of the platelet-sized nodules, including branching (Fig. 2 b), interconversion between condensed and lamellipodial forms (Figs. 1 and 2 a), and rapid translocation of particles (Fig. 2 c), and indicate that they are structurally unstable. More importantly, they establish that the mature cytoskeletal feature of a microtubule coil is only detected at the ends of proplatelets (Fig. 5) and not along their lengths.

**The Microtubule and Actin Cytoskeletons Drive Distinct Aspects of Proplatelet Formation**

The separation of proplatelet morphogenesis into stages...
characterized by specific alterations of the microtubule cytoskeleton and the recognition of distinct dynamic features provide approaches to investigation of the underlying molecular mechanisms. Megakaryocytes cultured in the presence of cytochalasin B, an inhibitor of actin assembly, retain the capacity to extend long, slender proplatelet-like projections (Fig. 7 a) but show specific abnormal features. The same results were observed with cytochalasin D. The cell body fails to spread, proplatelets extend from multiple points around the cell margin instead of the typical single erosion site, branching is completely inhibited, and although proplatelet processes retain bulbous ends, they harbor many fewer intermediate swellings. Thus, actin assembly is not required for megakaryocytes to extend proplatelets but is essential for proplatelet branching. Interestingly, one process associated with proplatelet bending is the attachment of a small region to the substrate over a period of 5-10 min and robust ruffling activity of this portion (data not shown), movements classically thought to be mediated by actin. In the electron microscope, the microtubule bundles composing the shaft routinely reveal small filamentous outpouchings (Fig. 8 a). At sites of bending, meshworks of actin filaments extrude processes that connect the microtubule bundles much like tendons attaching muscle to bone (Fig. 8 b). At more pronounced bends, the apparent sites of proplatelet branching, filamentous aggregates form a cusp between the microtubule bundles (Fig. 8 c). Considered together, these observations demonstrate that actin filaments are enriched at the sites of proplatelet bifurcation and probably required to execute this process.

The elongation of proplatelet extensions could be explained by forces derived from polymerization of tubulin subunits, from sliding of microtubules, or possibly a combination of both processes. Treatment with cytochalasin B results in substantially reduced complexity of proplatelets and permits isolated examination of the elongation process. In the representative sequence of micrographs shown in Fig. 7 b, we followed the growth of a single proplatelet. As this structure elongates, it rolls up upon itself to form a loop. While the distal end subsequently remains stationary (Fig. 7 b, arrowheads), the process continues to elongate and causes the diameter of the loop to increase. This strongly suggests that extension of proplatelet processes can occur without growth at the tip.

The microtubule stabilizing agent taxol completely blocks bending and branching along the length of the proplatelet tube and increases both the diameter of individual tubes and the thickness of microtubule bundles within them. Megakaryocytes treated with taxol remain capable of projecting short, thickened extensions with large bulbous tips. Fig. 7 c shows representative video-enhanced light microscopy designed to investigate the mechanics of proplatelet extension after taxol treatment, and illustrates that the processes formed are thickened compared with normal. As processes elongate in the presence of taxol,
Figure 5.
in released platelets (b) in these samples. (c) Low magnification electron micrograph showing the microtubule bundles lining the shaft of a typical proplatelet. Although proplatelet ends (arrowhead) have distinct microtubule coils, microtubules observed in swellings along the shaft (arrow) are not coiled. (d and g) Proplatelets stained for tubulin by immunofluorescence and photographed on a confocal microscope. Proplatelets show periodic segments connected by thin cytoplasmic bridges, but only the ends (arrowheads) have microtubule bundles arranged into teardrop-shaped loops. (e and f) Higher magnification electron micrograph of a swelling along a proplatelet shaft showing that these are points where the microtubule bundle separates for a short distance (arrows) but does not form a loop. Bars: (b and f) 0.5 μm; (c and e) 1 μm.

Figure 5. Organization of microtubules at the ends of proplatelet extensions (a–c) and in released proplatelet forms (d–g). (a) Anti-tubulin staining of a megakaryocyte and its early proplatelet extension. Microtubules concentrate along the edges of the megakaryocyte and enter into the proplatelet extensions. Bundles enter the projections from both sides and separate periodically within the extensions. The distal end of each proplatelet has a teardrop-shaped enlargement that contains a microtubule loop (arrowhead) have distinct microtubule coils, microtubules observed in swellings along the shaft (arrow) are not coiled. (d and g) Released platelet-sized particles are connected by thin cytoplasmic bridges, the most abundant being barbell shapes composed of two platelet-like particles connected by a single cytoplasmic strand. Microtubules line the shaft and rim the bulbous ends. (e) Low magnification electron micrograph showing the microtubule-based cytoskeleton of a representative released proplatelet form. A microtubule bundle lines the shaft of the proplatelet. Bar, 2 μm. (f) Higher magnification electron micrograph of one end of the proplatelet shown in e. Ends have microtubule bundles arranged into teardrop-shaped loops, similar to those at the ends of proplatelets extending from megakaryocytes. Within these loops are microtubule coils structurally similar to those observed in mature platelets isolated from the blood. Bar, 0.5 μm.

Figure 6. Organization of microtubules along the shaft of proplatelets. Swellings along the proplatelet (a) contain detergent-insoluble membrane and vesicles that are similar to those found in released platelets (b) in these samples. (c) Low magnification electron micrograph showing the microtubule bundles lining the shaft of a typical proplatelet. Although proplatelet ends (arrowhead) have distinct microtubule coils, microtubules observed in swellings along the shaft (arrow) are not coiled. (d and g) Proplatelets stained for tubulin by immunofluorescence and photographed on a confocal microscope. Proplatelets show periodic segments connected by thin cytoplasmic bridges, but only the ends (arrowheads) have microtubule bundles arranged into teardrop-shaped loops. (e and f) Higher magnification electron micrograph of a swelling along a proplatelet shaft showing that these are points where the microtubule bundle separates for a short distance (arrows) but does not form a loop. Bars: (b and f) 0.5 μm; (c and e) 1 μm.

their distal tip bends and curves back on itself until it makes contact with the shaft and appears to fuse with it to form a teardrop-shaped structure. In the presence of taxol, microtubules still align and compress in the cell cortex from which they enter pseudopodia (Fig. 7 d). In many cases, they do so from two sides and separate within the extension. Immunofluorescence reveals that microtubules either coil at the tips of projections or fold back on themselves to generate teardrop forms.

Discussion

The notion that mammalian platelets assemble through intermediate proplatelet structures was originally hypothesized by Becker and DeBruyn (1976), developed into a plausible model by Radley and co-workers (Radley and Scurfield, 1980; Radley and Haller, 1982), is strongly supported by the sum of many previous investigations, and is inconsistent with a competing model of thrombopoiesis that states that platelets are first formed as platelet territories within megakaryocytes. If the platelet territory model is true, as has been suggested by freeze-fracture and thin-section electron microscopy on the internal membranes of megakaryocytes (Y amada, 1957; Zucker-Franklin and Peterson, 1984), one would expect to find microtubule rings within such platelet fields (Radley and Hatshorn, 1987). This simple model finds little support here, or historically, as microtubule coils have not been observed in the megakaryocyte body. In contrast, many observers have documented the elaboration of proplatelets in megakaryocyte cultures (Handagama et al., 1987; Leven and Yee, 1987; Radley et al., 1987; Topp et al., 1990; A n et al., 1994; Choi et al., 1995; Cramer et al., 1997), and occasional micrographs have captured human megakaryocytes extruding proplatelet-like processes into bone marrow sinusoids (Behnke, 1968, 1969; Lichtman et al., 1978; Scurfield and Radley, 1981). The infrequent sighting of this morphology in situ most likely results from geometric limitations of thin-section transmission electron microscopy. Proplatelets have been recognized in a wide range of mammalian species, including mice, rats, guinea pigs, dogs, cows, and humans; platelet territories are not observed in megakaryocytes from some of these species. Moreover, the early characterization of proplatelets revealed many features that are consistent with known aspects of thrombopoiesis, including the critical requirement for microtubule integrity (Handagama et al., 1987; Tablin et al., 1990; Choi et al., 1995; Cramer et al., 1997). Finally, mice lacking two distinct hematopoietic transcription factors have severe thrombocytopenia and fail to produce proplatelets in culture (Lecine et al., 1998). Taken together, these findings establish the central importance of proplatelets in mammalian thrombopoiesis.

Here, we report detailed studies aimed at investigating
discrete steps in platelet formation by terminally differentiated mouse megakaryocytes, and our analysis represents the first effort to dissect dynamic aspects of this process. Proplatelets extend by a microtubule-based system, bifurcate repeatedly to increase the number of ends, and deliver packets of platelet material to these ends. Proplatelets and their contents are highly dynamic structures that continuously engage in alternate extension and retraction, reversible spreading, and bidirectional transport of particles. This dynamic behavior is inconsistent with the notion of proplatelets as static, linear arrays of nascent blood platelets, and strongly suggests that platelets mature along and at the ends of proplatelets in the final stages of an elaborate process. Our analysis suggests the model for platelet formation shown in Fig. 9.

Significance of the Dynamic Behavior of Proplatelet Morphogenesis

Segments of proplatelets clearly demonstrate the capacity to spread reversibly on the substratum and lose their characteristic tubular, beaded appearances. These changes occur both at the ends of tubes and along the shafts and are transient, with rapid reversion to the original, beads-on-a-string appearance (e.g., Figs. 1 and 2a). This remarkable plasticity needs to be reconciled with the presence in blood platelets of an elaborate membrane skeleton, where molecules of filamin A (ABP-280) extend through pores in the spectrin network, connecting actin filaments to the cytoplasmic tail of the \( \alpha \) chain of GPIb of the von Willebrand factor receptor, and create an arrangement that restricts the mobility of the membrane adherent spectrin lattice (Andrews and Fox, 1991; Hartwig and DeSisto, 1991; Kovacsovics and Hartwig, 1996). Although not discussed here, the plasma membranes of proplatelet extensions have a spectrin lattice identical in appearance to that of the mature platelet (Italiano, J.E., R.A. Shivdasani, and J.H. Hartwig, unpublished observations). Our observations on the dynamic nature of proplatelet morphogenesis suggest that the linkages holding the membrane skeleton in compression between the plasma membrane and actin cytoskeleton must be formed only during the final stages
of platelet maturation. Because lamellar spreading of proplatelets involves the dynamic assembly and disassembly of actin filaments, the filamin A–von Willebrand factor receptor linkage could only be established once the underlying actin cytoskeletal network is stabilized. This reinforces the notion that intermediate swellings do not correspond to assembled platelets, and suggests that platelet assembly completes just before release.
Figure 9. Model of platelet production suggested by these experiments and previous studies. After commitment to the megakaryocyte lineage, cells undergo polyploidization and cytoplasmic maturation (Stage 1). During the initial stages of proplatelet formation, megakaryocytes remodel their cytoplasm into thick pseudopodia (Stage 2), which contain bundles of microtubules situated just beneath the protruding membrane. Blunt pseudopodia elongate into proplatelet processes, which harbor thick bundles of microtubules in their core and contain a teardrop-shaped loop at their distal tip (Stage 3). Proplatelets frequently bend and form a branched structure from which new processes extend (Stage 4). These proplatelet processes form constrictions along their length giving the beaded appearance to proplatelets (Stage 5). The swellings along proplatelets are unstable structures presumed to contain packets of platelet material in the process of being delivered to the ends. Proplatelets are released from the megakaryocyte body after a retraction (Stage 6), and may undergo further fragmentation to yield individual platelets.

An additional dynamic feature of intermediate proplatelet segments is their rapid internal translocation. Viewed in the light microscope, nodular segments move bidirectionally along the proplatelet shaft, collide into each other, and fuse. These observations suggest that they are packets of material destined for assembly into nascent platelets, and the linear arrays of microtubules present throughout proplatelet processes probably serve as tracks for the transport of membrane, organelles, and granules into developing platelets. Therefore, we interpret the observed movement of particles (e.g., Fig. 2 c) to result from a combination of translocation along the microtubule tracks and proplatelet extension by microtubule growth.

Structural Insights into Proplatelet Formation

Proplatelet processes are generated by the elongation and thinning of blunt pseudopodia, which first appear at one pole of the cell and exhibit a unique organization of microtubule bundles parallel to the cell margins. Pseudopodial growth is associated with extension and apparent compression of these cortical microtubule bundles, and it is likely that the final orientation of microtubule bundles in the shafts of thinner proplatelet processes result from a continuation of this compression reaction. These structural details highlight earlier work (Radley and Haller, 1982; Handagama et al., 1987; Tablin et al., 1990) demonstrating the central role of microtubules in proplatelet formation; indeed, disassembly of cytoplasmic microtubules with nocodazole blocks formation of pseudopodia, whereas taxol treatment results in the formation of very few abnormal protrusions. Proplatelets terminate in bulbous structures invariably lined by a bundle of microtubules that originate in the shaft, form a teardrop-shaped loop within the bulb, and reenter the shaft. The loop itself reveals at least two discrete bundles, which must have opposing polarity, within the distal portion of the proplatelet.

Whether loop formation is part of the mechanism that drives tube extension or a structural consequence of this process is unclear. In the simplest model, assembly of microtubules at their plus ends, either at the tip or tube base, could generate the forces leading to proplatelet extension. The invariable finding of microtubule loops at the termini, however, argues against a model of plus end–driven tip growth, such as that which occurs in axonal extension. Moreover, extension of the proplatelet per se does not require tip elongation because time-lapse microscopy reveals stationary tips during periods of intense shaft growth (e.g., Fig. 7 b). Alternatively, tubulin subunits could add selectively to microtubules at the base and thereby elongate the tube; this model makes the testable prediction that the plus ends (the preferred sites of microtubule assembly) of proplatelet microtubules lie near the cell body, an unlikely possibility. Therefore, we favor the third possibility that tubulin subunits add to the plus ends of microtubules dispersed throughout the proplatelet shaft, and that microtubule motor proteins aid in tube elongation by sliding individual microtubules past one another. Such sliding would both lengthen the tube and reduce its diameter, consistent with the present observations, while microtubule-based motor activity might also contribute to the observed bends in proplatelet extensions and deliver platelet components to the ends.

Our examination of the sequence of events leading to proplatelet bending and bifurcation reveals critical details and again suggests mechanistic possibilities. For a proplatelet extension to bifurcate into two processes, three unique events must occur. First, the proplatelet shaft must bend sharply until it folds over on itself to form a loop that in turn elongates until it is structurally indistinguishable from its parent form. Second, microtubules within the tube must fragment, branch, or bend into loops, as depicted in Fig. 5, which elongate to form new processes. Third, either two sections of the shaft must fuse or looping must occur within the confines of the proplatelet shaft. The transient cell spreading and actin-based movements observed at sites of bifurcation may be necessary to accomplish this feat. The mechanical forces leading to tube branching require the activity of the actin cytoskeleton because treatment with cytochalasins prevents all branch formation. Regions with focal actin filament assembly may, thus, transmit contractile forces to the microtubule bundle to help compress it during loop formation.

In the last step, proplatelets separate from the residual cell mass and are released into the culture medium by a retractive mechanism. The tension generated by this retraction can be observed as a snapping of the strands between individual proplatelet extensions (data not shown). Although individual platelet-sized particles frequently appear in the culture, most of the released material consists...
of chains containing two or more platelet-sized particles. Release of individual platelet-sized particles in vivo may be accelerated by a fragmentation mechanism that is not expressed in cell culture or by shear forces normally encountered in the circulation. The process of proplatelet formation may also be modified by factors present in vivo. The bone marrow environment is composed of a complex adherent cell population containing endothelial cells, macrophages, and preadipocytes, which could play a role in proplatelet formation by direct cell contact or secretion of cytokines.

**Toward a Modified Proplatelet Model of Thrombopoiesis**

Our observations support the view that proplatelets are essential intermediates in platelet assembly, but refute the widely assumed concept that proplatelets are simply chains of fully and equally mature blood platelets. While platelet-sized units found along the shaft of proplatelets may represent intermediate stages in platelet matura-

tion, these segments are surprisingly motile and unstable. Therefore, we suggest a conservative reinterpretation of previous models that imply that each platelet-sized swelling observed along proplatelet shafts is a nascent blood platelet. Only the particles at the ends of proplatelets consistently display the mature feature of a single microtubule rolled into a coil. This finding posits the ends of proplatelets as the best sites of platelet assembly. One implication of these results is that proplatelet branching represents an elegant mechanism designed to increase the number of termini for productive thrombopoiesis. The process of proplatelet branching, therefore, appears to play an essential role in platelet biogenesis and provides a mechanism for increasing the final number of released platelets. By looping microtubules, the mechanism of proplatelet bifurcation may also initiate the process of rolling individual microtubules into the marginal ring destined to be incorporated into each mature blood platelet.

Reconstitution of platelet formation from murine megakaryocytes in vitro provides a powerful system for identifying the cytoskeletal components involved in the remarkable mechanism of platelet formation and release. More detailed structural analysis, coupled with the ability to study knockout mice lacking specific proteins will allow us to probe the contribution of individual components in thrombopoiesis. The principles learned from studying cytoskeletal dynamics in megakaryocytes are likely to provide insights into mechanisms of shape changes and movement of other cells.

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