Muscle aging is associated with compromised Ca\textsuperscript{2+} spark signaling and segregated intracellular Ca\textsuperscript{2+} release

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Reduced homeostatic capacity for intracellular Ca\textsuperscript{2+} ([Ca\textsuperscript{2+}]\textsubscript{i}) movement may underlie the progression of sarcopenia and contractile dysfunction during muscle aging. We report two alterations to Ca\textsuperscript{2+} homeostasis in skeletal muscle that are associated with aging. Ca\textsuperscript{2+} sparks, which are the elemental units of Ca\textsuperscript{2+} release from sarcoplasmic reticulum, are silent under resting conditions in young muscle, yet activate in a dynamic manner upon deformation of membrane structures. The dynamic nature of Ca\textsuperscript{2+} sparks appears to be lost in aged skeletal muscle. Using repetitive voltage stimulation on isolated muscle preparations, we identify a segregated [Ca\textsuperscript{2+}]\textsubscript{i} reserve that uncouples from the normal excitation–contraction process in aged skeletal muscle. Similar phenotypes are observed in adolescent muscle null for a synaptophysin-family protein named mitsugumin-29 (MG29) that is involved in maintenance of muscle membrane ultrastructure and Ca\textsuperscript{2+} signaling. This finding, coupled with decreased expression of MG29 in aged skeletal muscle, suggests that MG29 expression is important in maintaining skeletal muscle Ca\textsuperscript{2+} homeostasis during aging.

Introduction

Effective muscle contractile performance is contingent upon the maintenance of Ca\textsuperscript{2+} homeostasis and signaling, which requires that intracellular Ca\textsuperscript{2+} ([Ca\textsuperscript{2+}]\textsubscript{i}) be readily available for release. In skeletal muscle, excitation–contraction (E–C) coupling is primarily mediated by conformational coupling between voltage sensors of the sarcolemmal membrane and ryanodine receptor (RyR) Ca\textsuperscript{2+} release channel of the sarcoplasmic reticulum (SR; Ma et al., 1988; Rios et al., 1992; Franzini-Armstrong and Jorgensen, 1994). A secondary process, called Ca\textsuperscript{2+}-induced Ca\textsuperscript{2+} release (CICR), amplifies [Ca\textsuperscript{2+}]\textsubscript{i} release in skeletal muscle, particularly under stress conditions in muscle fatigue and dystrophy (Fong et al., 1990; Takagi et al., 1992; Brotto et al., 2002).

Results and discussion

Aging effects on muscle function have been associated with muscle fiber denervation, loss of motor units, and motor unit remodeling. Because functional alterations occur before significant muscle wasting becomes evident, changes in the E–C coupling machinery and [Ca\textsuperscript{2+}]\textsubscript{i} homeostasis may act as causative factors for, or adaptive responses to, muscle aging (Larsson and Edstrom, 1986; Faulkner et al., 1995; Delbono, 2002). We show that stress-induced Ca\textsuperscript{2+} sparks, which are the elemental events of CICR in striated muscles (Cheng et al., 1993; Klein et al., 1996), are severely compromised in aged skeletal muscle. In addition, we find that muscle aging is associated with the development of a segregated SR Ca\textsuperscript{2+} pool that uncouples from the normal E–C coupling machinery. We present evidence to suggest that mitsugumin-29 (MG29) may act as a sentinel against the effects of age on skeletal muscle Ca\textsuperscript{2+} homeostasis.

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Abbreviations used in this paper: ANOVA, analysis of variance; CICR, Ca\textsuperscript{2+}-induced Ca\textsuperscript{2+} release; E–C, excitation–contraction; EDL, extensor digitorum longus; [Ca\textsuperscript{2+}]\textsubscript{o}, extracellular Ca\textsuperscript{2+}; FDB, flexor digitorum brevis; [Ca\textsuperscript{2+}]\textsubscript{i}, intracellular Ca\textsuperscript{2+}; RyR, ryanodine receptor; SR, sarcoplasmic reticulum; TT, transverse tubule; VICR, voltage-induced Ca\textsuperscript{2+} release; wt, wild-type.

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were treated with osmotic shock to induce Ca\(^{2+}\) sparks. Exposure of the muscle fiber to a hypotonic solution leads to swelling of the fiber. Upon return to isotonic solution, the recovery of cell volume to normal is accompanied by a robust, peripherally localized Ca\(^{2+}\) spark response (Fig. 1 a; Wang et al., 2005). Young muscle fibers display a dynamic Ca\(^{2+}\) spark response to repeated stress cycles, with each round of osmotic shock generating Ca\(^{2+}\) sparks that continue for several minutes (Fig. 1, b and d). This Ca\(^{2+}\) spark response is located in the periphery of both young and aged muscle fibers. The dynamic nature of this Ca\(^{2+}\) spark response is significantly reduced in aged muscle (Fig. 1 c). Relative to young muscle (Fig. 1 d), aged muscle appears to contain a diminished capacity for the generation of dynamic Ca\(^{2+}\) sparks with repeated osmotic stresses (Fig. 1 e).

The diminished Ca\(^{2+}\) spark response in aged muscle could result from changes in resting [Ca\(^{2+}\)]\(_i\), levels or altered Ca\(^{2+}\) storage inside the SR. We measured [Ca\(^{2+}\)]\(_i\), levels and SR Ca\(^{2+}\) storage in FDB muscle fibers obtained from young and aged mice. As shown in Fig. 1 f, the resting [Ca\(^{2+}\)]\(_i\) levels appear to be similar between young and aged skeletal muscle fibers, whereas the caffeine/ryanodine-mobilized SR Ca\(^{2+}\) pool is significantly less in aged skeletal muscle. Thus, the reduced caffeine/ryanodine-mobilized SR Ca\(^{2+}\) store may represent one potential factor for the compromised Ca\(^{2+}\) spark signaling associated with muscle aging.

Additional factors that may contribute to this defective Ca\(^{2+}\) spark signaling include changes in membrane ultrastructure or altered expression of Ca\(^{2+}\) regulatory proteins in skeletal muscle. We conducted a survey of triad junction proteins and found that the expression level of MG29 (Takeshima et al., 1998), a synaptophysin-related membrane protein, is significantly down-regulated during muscle aging (Fig. 2, a and b). To determine the extent that decreased MG29 levels contribute to age-related alterations in muscle Ca\(^{2+}\) homeostasis, muscle fibers obtained from young (3–5 mo) mg29\((-/-)\) mice (Nishi et al., 1999) were stressed by osmotic shock. As with aged wild-type (wt) muscle, there is an initial Ca\(^{2+}\) spark response to the first osmotic shock and subsequent osmotic shocks produce little to no Ca\(^{2+}\) spark response in young mg29\((-/-)\) muscle fibers (Fig. 2 c). Using fura-2 Ca\(^{2+}\) measurements, we found that the resting [Ca\(^{2+}\)]\(_i\) level and SR Ca\(^{2+}\) storage are similar between aged wt and young mg29\((-/-)\) muscle fibers (Fig. 1 f). These results point to a role for MG29 in maintaining normal Ca\(^{2+}\) homeostasis that is lost with its diminished expression during aging.

Our previous studies have shown that the mg29\((-/-)\) mice display contractile alterations and muscle atrophy at ages of 6 mo or younger (Nishi et al., 1999; Nagaraj et al., 2000) that resemble the atrophic phenotype of aged wt mice. Electron microscopy studies reveal similar ultrastructural alterations to triad junction membrane structures of aged wt and young mg29\((-/-)\) skeletal muscle. Although organized alignment of SR and transverse-tubule (TT) membranes is present in young wt muscle, fragmented SR is frequently observed in slow (soleus) and fast (extensor digitorum longus [EDL]) twitch muscles from both young mg29\((-/-)\) and aged wt skeletal muscles (Fig. 3, b–d). Aged wt soleus muscle also displays swelling of the TT that is very similar to that seen in mg29\((-/-)\) muscle (Nishi et al., 1999). The development of these defects appears to be progressive during aging, as EDL fibers display...
a continuum of damage ranging from minor SR fragmentation (Fig. 3 c) to formation of large aggregates of SR (Fig. 3 e and Table S1, available at http://www.jcb.org/cgi/content/full/jcb.200604166/DC1; Agbulut et al., 2000; Chevessier et al., 2004). We suspect that aggregation of SR may result from the accumulation of subtle defects in Ca\(^{2+}\) signaling and membrane recycling, leading to progressive sarcopenia (Fig. 3, f and g).

The similar structural alterations seen in aged wt and young mg29\((−/−)\) muscle suggests that further defects in Ca\(^{2+}\) homeostasis beyond their Ca\(^{2+}\) spark response should be present. Previous studies by Kurebayashi et al. (2003) revealed that repetitive Ca\(^{2+}\) release using KCl stimulation on intact mg29\((−/−)\) muscle in the absence of extracellular Ca\(^{2+}\) ([Ca\(^{2+}\)]\(_i\)) did not lead to complete depletion of the SR Ca\(^{2+}\) pool. We now present evidence to show the presence of a segregated SR Ca\(^{2+}\) pool that uncouples from the normal voltage-induced Ca\(^{2+}\) release (VICR) machinery in young mg29\((−/−)\) and aged wt muscle (Fig. 4). When isolated, intact muscle fibers are exposed to 0 [Ca\(^{2+}\)]\(_o\) for 90 min, intermittent nonfatigue voltage stimulation leads to passive depletion of the voltage-sensitive pool of [Ca\(^{2+}\)]\(_i\) releasable, revealing striking differences among the three muscle preparations in their subsequent response to
Segregation of \( [Ca^{2+}] \) release in aged wild-type and young \( mg29(-/-) \) muscle.

(a) Force tracings of soleus muscle bundles after passive depletion of \( [Ca^{2+}] \) stores by single tetanic voltage stimulation contractions once per minute during incubation in 0 \( [Ca^{2+}] \), for \( \sim 90 \) min. Traces are cropped to only include the final six minutes of the experiment. 30 mM caffeine was added at points indicated by the arrows to induce contraction after the force generated by voltage stimulation could no longer be detected. (b) Tetanic voltage stimulation, for a 5-min duration, was applied to isolated soleus muscles in a solution containing 0 \( [Ca^{2+}] \). Arrows indicate the application of 30 mM caffeine. Although young \( wt \) muscles show a minimal response to caffeine, young \( mg29(-/-) \) and aged \( wt \) muscles display a significantly enhanced response to caffeine, indicating the presence of a \( Ca^{2+} \) pool that cannot be mobilized by extended voltage stimulation. (c) Data from multiple experiments are averaged (mean \( \pm \) SEM; \( n = 6-11 \)). Data is provided for both soleus and EDL skeletal muscles. * indicates significant differences, with \( P < 0.05 \) as measured by ANOVA.

Figure 4. Segregation of \( [Ca^{2+}] \) release in aged wild-type and young \( mg29(-/-) \) muscle.
in aged skeletal muscle, as it has been established that DHPR expression and the ratio of DHPR to RyR is decreased in aged skeletal muscle (Renganathan et al., 1997). Therefore, the reduced Ca\(^{2+}\) spark response observed in aged skeletal muscle suggests that changes in other cellular factors, such as MG29 expression, may play a role in regulation of Ca\(^{2+}\) signaling in skeletal muscle at different developmental stages.

MG29 contains a high degree of homology with synaptophysin, a protein thought to be involved with membrane fusion during exocytosis (Alder et al., 1992). Decreased MG29 expression may lead to improper lipid membrane formation or fusion, altering the dynamic process of membrane recycling and SR network formation. In aging wt or young mg29\((-/-)\) mice, the lack of this synaptophysin family member would suppress the efficient maintenance of triad junction structure, while also generating a fragmented SR network. Recent results from our laboratory have begun to shed light on the physiological function of MG29. We have demonstrated that MG29 increases sensitivity of the RyR channel to CICR when expressed in a heterologous cell system and when reconstituted with RyR in lipid bilayer single-channel studies (Pan et al., 2004). The lack of MG29 in aged and mg29\((-/-)\) muscle should decrease the sensitivity of RyR to CICR, in agreement with our current results.

Segregation of Ca\(^{2+}\) pools in aged muscle may have a physiological role in maintaining muscle integrity in the face of decreasing homeostatic capabilities. The resulting dampened Ca\(^{2+}\) mobilization in aged muscle may be a compensatory mechanism that protects aged fibers from Ca\(^{2+}\)-induced injury. It is also possible that the presence of a segregated Ca\(^{2+}\) reserve isolated from VICR responses contributes to cellular stress and decreased homeostatic capacity. Although the mechanism of active shuffling of Ca\(^{2+}\) from the VICR-responsive to the VICR-nonresponsive pool is not known, selective regulation of this Ca\(^{2+}\) shuffling to modulate the VICR-responsive pool would allow for the enhancement of aging skeletal muscle performance and/or protect skeletal muscle during aging. The mg29\((-/-)\) mouse represents a model system in which these mechanisms can be examined and these hypotheses can be tested.

### Materials and methods

#### Ca\(^{2+}\) imaging and spark analysis

Studies were conducted with wt C57Bl6/J male mice (Aged Rodent Colony, maintained by the National Institute on Aging), which were either 2–4 or 26–27 mo old. Male mg29\((-/-)\) mice (2–5 mo old), and wt animals of the same genetic background (129Sv/J backcrossed to C57Bl6/J) were maintained in local facilities and handled in a manner approved by local regulations. FDB fibers were isolated by enzymatic disassociation in 0.2% type I collagenase (Sigma-Aldrich) for 55 min at 37°C and loaded with 10 μM Fluo-4-AM for 60 min at room temperature. Mean FDB fiber size was 1 mm × 20 μm. Measurements of Ca\(^{2+}\) release were performed on a confocal microscope [Radiance 2100; Bio-Rad Laboratories] equipped with an argon laser (488 nm) and a 60×, 1.3 NA, oil immersion objective. For Ca\(^{2+}\) spark measurements, fibers were perfused with a 170-mosM hypertonic solution containing (in mM) 70 NaCl, 5 KCl, 10 Hepes, 2.5 CaCl\(_2\), 2 MgCl\(_2\), pH 7.2, for 60–180 s to induce swelling before perfusion was switched back to the initial Tyrode solution (in mM) 140 NaCl, 5 KCl, 10 Hepes, 2.5 CaCl\(_2\), 2 MgCl\(_2\), pH 7.2, with an osmolarity of 290 mosM as measured by a Micro Osmometer 3300 (Advanced Instruments). Image analysis was performed using custom routines on IDL software (Research Systems, Inc.; Cheng et al., 1999; Wang et al., 2005).

For determination of resting cytosolic Ca\(^{2+}\) levels and total SR Ca\(^{2+}\) store, individual FDB fibers were loaded with 10 μM fura-2 AM for 45 min at room temperature in Tyrode solution. 20 μM N-benzylp-toluene sulfonamide, a myosin II inhibitor, was applied for 15 min to prevent motion artifact from muscle contraction (Cheung et al., 2002; Pinninger et al., 2005). Fibers were also embedded into silicone grease to maintain their position in the culture dish (Jacquemond, 1997). The ratio of fura-2 fluorescence at excitation wavelength of 350 and 380 nm was measured on a P1 spectrofluorometer (Photon Technology International) to assess the resting [Ca\(^{2+}\)]\(_i\). The SR Ca\(^{2+}\) store was measured by addition of 20 mM caffeine plus 5 μM ryanodine in the presence of 0 [Ca\(^{2+}\)]\(_i\). Aged Rodent Colony, maintained by the National Institute on Aging), which were either 2–4 or 26–27 mo old. Male mg29\((-/-)\) mice (2–5 mo old), and wt animals of the same genetic background (129Sv/J backcrossed to C57Bl6/J) were maintained in local facilities and handled in a manner approved by local regulations. FDB fibers were isolated by enzymatic disassociation in 0.2% type I collagenase (Sigma-Aldrich) for 55 min at 37°C and loaded with 10 μM Fluo-4-AM for 60 min at room temperature. Mean FDB fiber size was 1 mm × 20 μm. Measurements of Ca\(^{2+}\) release were performed on a confocal microscope [Radiance 2100; Bio-Rad Laboratories] equipped with an argon laser (488 nm) and a 60×, 1.3 NA, oil immersion objective. For Ca\(^{2+}\) spark measurements, fibers were perfused with a 170-mosM hypertonic solution containing (in mM) 70 NaCl, 5 KCl, 10 Hepes, 2.5 CaCl\(_2\), 2 MgCl\(_2\), pH 7.2, for 60–180 s to induce swelling before perfusion was switched back to the initial Tyrode solution (in mM) 140 NaCl, 5 KCl, 10 Hepes, 2.5 CaCl\(_2\), 2 MgCl\(_2\), pH 7.2, with an osmolarity of 290 mosM as measured by a Micro Osmometer 3300 (Advanced Instruments). Image analysis was performed using custom routines on IDL software (Research Systems, Inc.; Cheng et al., 1999; Wang et al., 2005).

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12 mm and a mean mass of 80 mg, whereas soleus muscles had a mean length of 10 mm and a mean mass of 10 mg. Muscles were mounted vertically on a glass-stimulating apparatus (Radnoti) with platinum electrodes and attached to a movable isometric force transducer and to a stationary anchor, which allowed muscles to be stretched until both maximal forces for a given frequency and the frequency producing $T_{\text{max}}$ were obtained.

**Force measurements during passive depletion and fatigue**

After $T_{\text{max}}$ was determined, the intact muscles were allowed to equilibrate for 20 min in the Ringer’s solution. During equilibration, muscle strips were stimulated with $\sim 100–120$ Hz (EDJ) or $\sim 60–80$ Hz (SOL), 330 mA, 500-ms electrical pulse-trains administered with a periodicity of 1 min to generate $T_{\text{max}}$. After equilibration, the muscles from the passive-depletion group were washed five times in Ringer’s solution with the same composition as described in the previous section, except that no CaCl$_2$ was added while 0.1 mM EGTA was added, to create a nominal 0 [Ca$^{2+}$]$_i$ solution. Muscles were stimulated with one $T_{\text{max}}$ every minute in 0 [Ca$^{2+}$]$_i$ solution until force declined to nondetectable levels; the muscles were then exposed to 30 mM caffeine. Force produced in response to caffeine application was recorded until a stable plateau was obtained. After the passive depletion protocol, muscles were subjected to extensive washes in normal Ringer’s solution containing 2.5 Ca$^{2+}$ and then electrically stimulated until force returned to initial equilibration values. In 80% of our preparations, this was achieved. After forces were stable and comparable to the initial levels before the onset of the passive depletion, muscles were returned to the 0 [Ca$^{2+}$]$_i$ solution for 5 min and subsequently subjected to a 1–5 min fatiguing protocol consisting of the same stimulatory pattern administered at a 1-s periodicity (i.e., 50% duty cycle). In between fatigue runs, muscles were washed in 2.5 Ca$^{2+}$ solution and force was allowed to recover to prefatigue levels before the onset of the next fatigue run. At the end of each fatiguing protocol, muscles were treated with 30 mM caffeine and maximal response to caffeine was recorded. Caffeine was mixed in a small volume of the Ringer’s solution and added to the chambers to produce a final concentration of 30 mM in the bathing chamber. Whenever possible, paired experiments were performed with young wild-type animals and aged mg29(wt) mice. Paired experiments followed the protocols as previously described (Brotto and Nosek, 1996; Brotto et al., 2004).

**Statistical analysis**

All statistical analysis in this study was conducted using ANOVA, and data is presented as the mean ± SEM.

**Online supplemental material**

Table S1 describes the parallel disruption of triad junctions in aged wt and young mg29(-/-) skeletal muscle. Table S2 is an assessment of muscle contractile function in young wt, aged wt, and young mg29(-/-) mice. Online supplemental material is available at http://www.jcb.org/cgi/content/full/jcb.200604166/DC1.

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