The UNC-45 chaperone mediates sarcomere assembly through myosin degradation in Caenorhabditis elegans

Megan L. Landsverk,1,2 Shumin Li,1 Alex H. Hutagalung,2 Ayaz Najafov,1 Thorsten Hoppe,3 José M. Barral,1 and Henry F. Epstein1

1Department of Neuroscience and Cell Biology, University of Texas Medical Branch, Galveston, TX 77555
2The Verna and Marrs McLean Department of Biochemistry and Molecular Biology, Baylor College of Medicine, Houston, TX 77030
3Centre for Molecular Neurobiology, University of Hamburg, 20251 Hamburg, Germany

Myosin motors are central to diverse cellular processes in eukaryotes. Homologues of the myosin chaperone UNC-45 have been implicated in the assembly and function of myosin-containing structures in organisms from fungi to humans. In muscle, the assembly of sarcomeric myosin is regulated to produce stable, uniform thick filaments. Loss-of-function mutations in Caenorhabditis elegans UNC-45 lead to decreased muscle myosin accumulation and defective thick filament assembly, resulting in paralyzed animals. We report that transgenic worms overexpressing UNC-45 also display defects in myosin assembly, with decreased myosin content and a mild paralysis phenotype. We find that the reduced myosin accumulation is the result of degradation through the ubiquitin/proteasome system. Partial proteasome inhibition is able to restore myosin protein and worm motility to nearly wild-type levels. These findings suggest a mechanism in which UNC-45-related proteins may contribute to the degradation of myosin in conditions such as heart failure and muscle wasting.

Introduction

Myosins are actin-based motors that play essential roles in a variety of cellular processes, including cytokinesis, cellular trafficking, phagocytosis, maintenance of cell shape, and muscle contraction (Berg et al., 2001). Myosin-based movement results from a precise cycle of the myosin head binding and releasing ATP and actin. During this process, the myosin head transitions through multiple folding conformations.

Molecular chaperones appear necessary for de novo folding and structural maintenance of the myosin head. Expression of the myosin motor domain in bacteria results in misfolding (McNally et al., 1988). In vertebrate systems, the chaperonin containing TCP-1 (CCT), as well as molecular chaperones Hsp90 and Hsc70, are necessary but not sufficient in the folding of striated muscle myosin (Srikakulam and Winkelmann, 1999; Srikakulam and Winkelmann, 2004).

Evidence from a variety of experimental systems indicates that myosins use specialized chaperones during their activity, folding, and assembly. Mutations in UNC-45/Cro1p/She4p(Dim1p) domain (UCS) proteins lead to phenotypes related to defects in myosin folding and assembly (Hutagalung et al., 2002). Decreased UCS domain protein function in fungal mutants produces myosins defective in actin:ATP transduction (Wesche et al., 2003; Lord and Pollard, 2004). In Caenorhabditis elegans, null unc-45 alleles lead to embryonic arrest of body wall muscle development (Venolia and Waterston, 1990), and temperature-sensitive mutations lead to a paralyzed or uncoordinated phenotype at the restrictive temperature with marked disorganization of myofibrils (Epstein and Thomson, 1974; Barral et al., 1998). In vitro, UNC-45 exerts chaperone activity on the myosin head and acts as a cochaperone that specifically binds Hsp90 (Barral et al., 2002).

UNC-45 has recently been shown to be a substrate of an E3/E4-multiubiquitination complex containing CHN-1 (the C. elegans homologue of CHIP) and UFD-2 (Hoppe et al., 2004). chn-1-null worms are viable and morphologically normal. However, UNC-45 overexpression leads to an uncoordinated phenotype in these worms, suggesting that increased levels of UNC-45 may cause muscle defects. Previous studies have
Figure 1. Overexpression of UNC-45FLAG results in decreased thick filament assembly and a mild paralysis phenotype. (A) Expression levels of UNC-45FLAG protein by two different integrated lines. Two integrated lines overexpressing UNC-45, suIs1, and suIs2 were analyzed for UNC-45FLAG levels by Western blots using a mAb to the C-terminal FLAG tag on transgenic UNC-45. Each lane contains 10 worms total. (B) UNC-45FLAG overexpression results in a mild paralysis phenotype. Motility in liquid was measured at 20°C for N2 and suIs2 grown at 20°C, unc-45(e286) grown at 15 and 20°C, and suIs2; unc-45(e286) grown at 25°C. N2 and suIs2 worms showed a mean of 53 and 21 body bends, respectively, over 15 s. The UNC-45FLAG transgene expressed in suIs2 was able to rescue the motility defect of unc-45(e286) at 25°C from a mean of 2 to 16 bends in 15 s. *, P < 0.0001 versus N2 wild type. (C) suIs2 worms have decreased number of MHC-containing A bands in body wall muscle cells. Ya worms were labeled with an anti-MHC A mAb to reveal the presence of A bands. (D) Quantitation of the myosin assembly defect of suIs2 worms. The number of A bands per cell in regions I and II, posterior to the pharynx (MacKenzie et al., 1978b) of N2 and suIs2 worms, was counted. In region II, N2 worms have a mean of 7.8 A bands per cell, whereas suIs2 worms have a mean of 5.8. In region III, N2 and suIs2 worms have 8.7 and 6.3 A bands per cell, respectively. *, P < 0.0001 versus N2 wild type. Error bars indicate SD.

shown that unassembled components of myofilbrils are degraded through the ubiquitin/proteasome system (UPS) in muscle wasting conditions, including cancer and starvation (Koohmaraie, 1992; Solomon and Goldberg, 1996; Solomon et al., 1998; Acharyya et al., 2004). Our results suggest that enhanced levels of UNC-45 may promote nonnative myosin conformations, rendering them susceptible to degradation by the UPS.

Results and discussion

Wild-type levels of UNC-45 are critical for proper myofilbril assembly

Loss-of-function (Lof) temperature-sensitive UNC-45 mutations result in severely paralyzed worms, with pronounced disorganization of the sarcomere (Epstein and Thomson, 1974; Barral et al., 1998). To investigate the consequences of increased UNC-45 levels in a wild-type background, we used the transgenic line, suIs2 [Punc-54::unc-45FLAG], expressing UNC-45 under control of the strong muscle–specific unc-54 promoter. In contrast to lines overexpressing UNC-45FLAG from an extrachromosomal array, this line stably transmits the transgene from generation to generation. suIs2 worms overexpress UNC-45 at ~10-fold greater amounts than that of the previously studied integrated line suIs1 [Punc-54::unc-45FLAG] (Hoppe et al., 2004; Fig. 1 A).

Although not as pronounced as in Lof alleles, the increased concentrations of UNC-45 in suIs2 also lead to abnormal thick filament assembly and a concomitant defect in movement. We measured body bend rates and found that suIs2 worms were 36% slower than wild type (Fig. 1 B). However, this is not as severe as the temperature-sensitive unc-45(e286) mutants grown at the restrictive temperature, which is the result of diminished, rather than augmented, UNC-45 function. Transgenic UNC-45FLAG was able to rescue the motility defects...
of unc-45(e286) at the restrictive temperature, indicating that it is functional (Fig. 1 B).

*C. elegans* young adult (Ya) wild-type muscle cells are ~120–150 μm in length with a mean of seven to nine myosin thick filament–containing A bands per cell (Fig. 1 C; Mackenzie et al., 1978a). Similarly staged *suIs2* worms demonstrated cells 108–130 μm in length that contained only five to six A bands (Fig. 1, C and D). Thus, UNC-45 overexpression in *suIs2* worms results in a decreased number of myosin-containing thick filaments. This phenotype arises specifically from increased levels of UNC-45\(^{\text{FLAG}}\). Transgenic overexpression of the unrelated protein GFP under control of the *unc-54* promoter does not result in altered muscle cells (Fig. S1 A, available at http://www.jcb.org/cgi/content/full/jcb.200607084/DC1). Furthermore, integration of the UNC-45\(^{\text{FLAG}}\) transgene by itself is not responsible for the muscle phenotype, as the expression of UNC-45\(^{\text{FLAG}}\) as an extrachromosomal array also decreased the number of myosin-containing A bands in cells in which it was expressed (Fig. S1 B). Moreover, in rare instances in which individual muscle cells in *suIs2* worms expressed undetectable levels of UNC-45\(^{\text{FLAG}}\), a wild-type phenotype was observed (Fig. S1 C).

**Altered UNC-45 levels lead to decreased myosin accumulation**

We next investigated whether the overall myosin content was decreased in these worms. Four distinct myosin heavy chain (MHC) isoforms are expressed in *C. elegans* as major components of the sarcomere (Miller et al., 1983; Dibb et al., 1989). MHC A and B are present in the body wall muscles, whereas MHC C and D are located in the minor pharyngeal muscles (Ardizzi and Epstein, 1987). To assess myosin levels, individual worms were analyzed for MHC A, B, and D. In contrast to MHC D, which was not affected because UNC-45\(^{\text{FLAG}}\) is specifically expressed in body wall muscle, both body wall MHC A and B were decreased in *suIs2* to ~70% of wild type (Fig. 2 A). Worms homozygous for the e286 allele grown at the restrictive temperature resulted in decreased accumulation of all myosin isoforms examined (Fig. 2 A). This is expected, as endogenous UNC-45 is involved in pharyngeal as well as body wall muscle myosin assembly (Venolia and Waterston, 1990). Previous results showing that MHC A content was not substantially affected in e286 mutant worms may be explained by the fact that the protein paramyosin was used for normalization. Paramyosin is also a major component of thick filaments, and e286 worms at the restrictive temperature have a 45% mean decrease in number of thick filaments compared with worms at the permissive temperature (Barral et al., 1998). Paramyosin levels are also decreased in *suIs2* worms, indicating a loss of total thick filaments (Fig. S2, available at http://www.jcb.org/cgi/content/full/jcb.200607084/DC1).

Because our transgenic line carries multiple copies of the *unc-54* promoter, the possibility existed that the observed muscle phenotype was caused by its excessive binding of transcription factors. However, real-time RT-PCR verified that mRNA levels of both *myo-3* and *unc-54*, which encode MHC A and B, respectively, were not significantly different between N2 wild-type and *suIs2* worms (Fig. 2 C) with GAPDH mRNA as a control. These results indicated that the decrease in body wall myosin is not a result of diminished transcription.

**The UPS is responsible for decreased myosin accumulation in worms overexpressing UNC-45**

To examine a role for the UPS in the degradation of endogenous myosin in *C. elegans*, we first tested whether we could detect ubiquitinated myosin species in whole worm lysates. We performed pull downs from wild-type worms with antibi ubiquitin and antiamyosin antibodies and blotted them subsequently with
a myosin-specific mAb (Fig. 3A). The ubiquitin mAb was able to pull down full-length myosin as well as species of slower mobility, consistent with polyubiquitinated myosin species. Several smaller myosin-immunoreactive bands were also detected in both pull downs, which may represent ubiquitinated proteolytic fragments of myosin, consistent with previous studies (Zdinak et al., 1997; Ikemoto et al., 2001; Acharyya et al., 2004; Du et al., 2004). Sarcomeric myosin can thus be ubiquitinated in C. elegans. Elevated ubiquitination would lead to accelerated degradation, which in fact was observed in lysates of suIs2 and unc-45(e286) worms at 25°C when compared with N2 and unc-45(e286) worms at 15°C (Fig. S3, available at http://www.jcb.org/cgi/content/full/jcb.200607084/DC1).

Proteasome-dependent proteolytic degradation requires ATP and is inhibited by MG132 (Rock et al., 1994). We thus tested whether degradation of endogenous C. elegans myosin exhibited these properties. We incubated separate reactions containing crude wild-type worm extracts with no additions, ATP supplementation, MG132 supplementation, and a combination of both, and examined the levels of full-length myosin and its degradation products by immunoblotting (Fig. 3B). The un-supplemented reaction exhibited constant levels of full-length myosin, whereas the reaction supplemented with ATP showed a clear reduction of full-length myosin over time. MG132 by itself had no effects on full-length myosin levels, but the observed ATP-dependent degradation was effectively blocked by addition of MG132. The proteasome is therefore capable of degrading myosin in C. elegans.

We next tested whether proteasome inhibition could restore the diminished myosin levels observed in the suIs2 worms that overexpress UNC-45FLAG. RNAi against the proteasomal subunit RPT-2 has been previously used to inhibit proteasome function in living C. elegans worms (Hoppe et al., 2004). Importantly, at the concentration used, RPT-2 RNAi did not result in complete inhibition of the proteasome, which allowed the nematodes to survive. When RPT-2 RNAi was fed to suIs2 worms, we observed clear restoration of body wall myosin (Fig. 3, C and D). Furthermore, RPT-2 RNAi also significantly improved the mobility of suIs2 nematodes (Fig. 3E). Because UNC-45 itself is subject to degradation by the UPS, the previous observation that unc-45 Lof mutants are rescued by RPT-2 RNAi (Hoppe et al., 2004) can be explained by increased levels of UNC-45, myosin, or both. Our current results show that endogenous myosin can be ubiquitinated in C. elegans and that proteasome inhibition can prevent its degradation, in both a wild-type background as well as that resulting from increased UNC-45 function.

Implications for mechanism and muscle disease

We have found that UNC-45 overexpression results in diminished myosin accumulation and assembly because of its increased degradation via the UPS. Lof mutants also show similar but enhanced defects. These results allow us to propose an idealized model in which precise levels of UNC-45 are critical for supporting adequate myosin folding and assembly into thick filaments (Fig. 4). As a myosin chaperone (Barral et al., 2002), UNC-45 binds to newly synthesized myosin motor domains that have not yet attained their full native structure. UNC-45 assists in folding of the motor domain so that it becomes competent for assembly. We hypothesize that myosin produced in the context of UNC-45 Lof mutations is less capable of attaining its native structure and, as a result, is degraded by the UPS, thus explaining its drastically reduced levels (Fig. 2A) and aberrant assembly into thick filaments (Barral et al., 1998). Myosin synthesized in the context of excess UNC-45 may also be prevented from reaching the assembled state because of mass action and is then susceptible to degradation via the UPS. Because both endogenous and transgenic UNC-45 can associate with assembled myosin in vivo (Ao and Pilgrim, 2000; unpublished data), excess UNC-45 may also be capable of shifting the myosin equilibrium from the assembled state into a degradation-susceptible unassembled state. The interplay between UNC-45, myosin, and the UPS may be of relevance to similar concentration-dependent phenomena between chaperones and client proteins.
capable of polymerization, such as those reported for Hsp104 and yeast prions (Chernoff et al., 1995).

Vertebrates contain two distinct genes encoding different UNC-45 isoforms. General cell UNC-45 is expressed in multiple tissues and appears necessary for various cytoskeletal functions, whereas striated muscle (SM) UNC-45 is specifically expressed in heart and skeletal muscle and may be necessary for sarcomere organization (Price et al., 2002). SM UNC-45 may function during sarcomere assembly in a mechanism similar to the one described here for C. elegans UNC-45. Alterations in human SM UNC-45 function or concentrations may be significant in hypertrophy, dilation, and failure of the heart, as well as in skeletal muscle wasting in a variety of human disorders.

**Materials and methods**

**General C. elegans methods**

Nematode strains were grown under standard conditions (Brenner, 1974). N. andrewsi strains were grown under standard conditions (Brenner, 1974). Stage worms were lysed in 50 mM NaPO4, pH 7.4, 200 mM NaCl, 5 mM MgCl2, 1% Triton X-100, 5 mM DTT, and 5 mM ATP. Samples were resolved by SDS-PAGE and transferred to Hybond nitrocellulose (GE Healthcare) for 2 h at 4°C, and the supernatant was divided into 5,000 g at 4°C, and the supernatant was divided into aliquots to which no supplementation, 2 mM ATP, and/or 100 μM proteasome inhibitor MG132 were added. Samples were kept at room temperature, and aliquots were removed at each time point, mixed with SDS-PAGE sample buffer, and immediately heated at 95°C for 10 min. Samples were resolved by SDS-PAGE followed by Western blots using 5 μg/ml mAb 28.2.

**Immunoblotting**

In Fig. 2 A, Fig. 3 C, and Fig. S2, Ya worms were hand-picked, placed in SDS sample buffer, and heated at 95°C for 10 min. Amounts corresponded in Fig. 2 A, Fig. 3 C, and Fig. S2, Ya worms were hand-picked, placed in SDS sample buffer, and heated at 95°C for 10 min. Amounts corresponded in Fig. 2 A, Fig. 3 C, and Fig. S2, Ya worms were hand-picked, placed in SDS sample buffer, and heated at 95°C for 10 min. Amounts corresponded in Fig. 2 A, Fig. 3 C, and Fig. S2, Ya worms were hand-picked, placed in SDS sample buffer, and heated at 95°C for 10 min. Amounts corresponded in Fig. 2 A, Fig. 3 C, and Fig. S2, Ya worms were hand-picked, placed in SDS sample buffer, and heated at 95°C for 10 min. Amounts corresponded in Fig. 2 A, Fig. 3 C, and Fig. S2, Ya worms were hand-picked, placed in SDS sample buffer, and heated at 95°C for 10 min. Amounts corresponded in Fig. 2 A, Fig. 3 C, and Fig. S2, Ya worms were hand-picked, placed in SDS sample buffer, and heated at 95°C for 10 min. Amounts corresponded.

**Microscopy**

Immunofluorescence microscopy on whole mounts of Ya worms was performed as described previously (Finney and Ruvkun, 1990). Worms were reacted with rhodamine-conjugated mAb 5-6 (anti-MHC A), FITC-conjugated mAb 5-8 (anti-MHC B), or Cy3-conjugated mAb 5-23 (anti-paramyosin; Miller et al., 1983), or anti-HDA-1 (Santa Cruz Biotechnology, Inc.) at 2, 5, 25, 5, and 2 μg/ml, respectively, for 1 h, followed by HRP-conjugated anti–mouse IgG or anti-rabbit IgG at 1:10,000 dilutions. Densitometry was performed using AlphaEaseFC software (Alpha Innotech).

**Motility assays**

Individual Ya worms were placed in M9 buffer, and body bends were counted over 15 s (Epstein and Thomson, 1974).

**RNAs**

RNAi RNAi was performed by feeding nematodes the HT115 bacteria containing pRGT2 double-stranded RNA in plasmid pD129.36 (Hoppe et al., 2004) as described previously (Kamath et al., 2001) except that RNA transcripts were induced with 10 nM IPTG to ensure survival.

**Myosin degradation assay**

N2 worms were equilibrated and lysed in 20 mM Tris-HCl, pH 8.0, 200 mM NaCl, 5 mM MgCl2, and 5 mM DTT. The homogenate was centrifuged at 5,000 g for 10 min at 4°C, and the supernatant was divided into four aliquots to which no supplementation, 2 mM ATP, and/or 100 μM proteasome inhibitor MG132 were added. Samples were kept at room temperature, and aliquots were removed at each time point, mixed with SDS-PAGE sample buffer, and immediately heated at 95°C for 10 min. Samples were resolved by SDS-PAGE followed by Western blots using 5 μg/ml mAb 28.2.

**Pull-down assays**

N2 worms were lysed and incubated with 100 μg mAb 28.2 or 200 μg anti-ubiquitin mAb (Santa Cruz Biotechnology, Inc.) bound to protein G Sepharose (GE Healthcare) for 2 h at 4°C. Pull-downs were washed five times with lysis buffer, mixed with SDS sample buffer, and heated at 95°C for 10 min. Samples were separated by SDS-PAGE followed by Western blotting using 5 μg/ml mAb 28.2. Fivefold less volume of the MHC B pull down compared with the ubiquitin pull down was loaded to avoid saturation.

**Online supplemental material**

Fig. S1 shows that the suIs2 phenotype arises specifically from UNC-45FLAG overexpression. Fig. S2 shows that paramyosin levels are also decreased in suIs2 worms. Fig. S3 shows that myosin accumulation is also decreased in whole worm lysates from suIs2 and unc-45(e286) worms at the restrictive temperature. Online supplemental material is available at http://www.jcb.org/cgi/content/full/jcb.200607084/DC1.

We thank Dr. N. Dharajia for assistance with real-time RT-PCR, Dr. David Baillie for critically reading the manuscript.

This work was supported by grants from the National Institutes of Health (ROIAR500551), the Muscular Dystrophy Association, and the Cecil H. and Ida M. Green Endowment to H.F. Epstein.

Submitted: 17 July 2006
Accepted: 5 March 2007

**References**


