Accurate segregation of replicated chromosomes during mitosis is essential for the maintenance of genomic integrity. To ensure faithful chromosome segregation, eukaryotic cells have developed a surveillance network called the mitotic checkpoint that delays anaphase onset until sister kinetochores of duplicated chromosomes are properly attached to microtubules emanating from opposite spindle poles (for reviews see Kops et al., 2004; Musacchio and Salmon, 2007). Early in mitosis, various mitotic checkpoint proteins, including Bub1, Bub3, BubR1, Mad1, Mad2, and Mps1, are recruited to unattached kinetochores. These kinetochore-associated checkpoint proteins promote the formation of diffusible Mad2, BubR1, Bub3, and Cdc20 protein complexes that inhibit the anaphase-promoting complex/cyclosome (APC/C), an E3 ubiquitin ligase that drives cells into anaphase by targeting securin and cyclin B for destruction by the 26S proteasome (for reviews see Kops et al., 2005; Peters, 2006; Musacchio and Salmon, 2007). After all chromosome pairs are properly attached to the spindle and aligned in the metaphase plate, mitotic checkpoint proteins dissociate from the APC/C, thus triggering the ubiquitin-mediated destruction of securin and cyclin B. Separase, a protease that is inhibited by securin binding and cyclin B/Cdk1-mediated phosphorylation, then cleaves the kleisin subunit Scc1 of cohesin, thereby allowing sister chromatid disjunction and anaphase onset (for reviews see Nasmyth and Haering, 2005; Peters, 2006).

The discovery of the mitotic checkpoint led to speculation that mutations in mitotic checkpoint genes might play a role in the development of aneuploidy in human cancers (Jallepalli and Lengauer, 2001; Draviam et al., 2004). Over recent years, mutant mitotic checkpoint genes have indeed been identified in various human cancers, although at relatively low frequency (Weaver and Cleveland, 2006; for reviews see Kops et al., 2005; Yuen et al., 2005). The Bub1 kinase is mutated in several cancer types, including colorectal, lung and thyroid cancer, and T cell leukemia (Cahill et al., 1998; Ohshima et al., 2000; Ru et al., 2002; Shichiri et al., 2002). In addition, Bub1 expression is frequently reduced in several human cancers, including colorectal, gastric, and esophageal cancers (Shigeishi et al., 2001; Shichiri et al., 2002; Doak et al., 2004).

Bub1 is a serine/threonine protein kinase that targets to unattached kinetochores at the onset of mitosis (Roberts et al., 1994;
There it is thought to phosphorylate Cdc20, thereby preventing Cdc20 from activating the APC/C (Chung and Chen, 2003; Chen, 2004; Tang et al., 2004a). Additionally, Bub1 is required for Mad1–Mad2 localization to unattached kinetochores. These complexes function to prevent premature APC/C activation by changing the conformation of monomeric Mad2 such that it can efficiently bind to and inhibit the APC/C coactivator Cdc20 (Luo et al., 2002, 2004; Sironi et al., 2002). Besides Mad1 and Mad2, Bub1 also recruits BubR1, Bub3, centromere protein E (CENP-E), and CENP-F to unattached kinetochores (Sharp-Baker and Chen, 2001; Johnson et al., 2004). Several of these proteins are important for microtubule-kinetochore attachment, which may explain why Bub1-depleted cells have chromosome congression defects (Meraldi and Sorger, 2005). In addition, Bub1 contributes to the stability and inner centromere localization of Shugoshin (Sgo1), a protein that functions as an adaptor for phosphatase PP2A (Tang et al., 2004b, 2006; Kitajima et al., 2005). At the inner centromere, PP2A counteracts the Plk1-mediated release of cohesin until anaphase onset, thus preventing the premature separation of sister centromeres (Riedel et al., 2006). Bub1 also controls the stability and correct positioning of the chromosomal passenger complex to the inner centromeric region of sister chromosomes, a function that appears to be critical for the recruitment of Sgo1 to centromeres (Boyarchuk et al., 2007).

Although the molecular mechanisms of Bub1 action are beginning to emerge, the physiological role of Bub1 in higher eukaryotes is still unknown. The most definitive way to address this role would be to generate Bub1 knockout mice by homologous recombination in embryonic stem (ES) cells. However, previous gene knockout studies for Mad1, Mad2, BubR1, and Bub3 revealed that these mitotic checkpoint proteins are essential for cell proliferation, causing mice to die during the early
stages of embryonic development (Dobles et al., 2000; Kalitsis et al., 2000; Babu et al., 2003; Baker et al., 2004; Wang et al., 2004; Iwanaga et al., 2007). Anticipating that Bub1-null mice would be embryonically lethal as well, we generated a series of mice in which the expression of Bub1 protein is reduced in a graded fashion from normal to zero. We find that Bub1-null mice are indeed embryonically lethal but that mice with very low levels of Bub1 protein are viable. Here, we show that Bub1 deficiency is associated with aneuploidy and spontaneous tumorigenesis in a dose-dependent fashion. Furthermore, we provide evidence for a novel role of Bub1 in eliminating cells that have undergone chromosome missegregation.

**Results**

**Generation of mutant mice with graded reduction in Bub1 levels**

By homologous recombination, we inserted a neomycin-resistance (Neo) gene flanked by a 
\textit{loxP} site into intron 8 and a 
\textit{loxP} site into intron 9 of the mouse Bub1 gene (Fig. 1 A). This created a hypomorphic allele (called Bub1N) because the Neo gene harbors a cryptic exon that is known to reduce the level of normally spliced messenger RNA (Jacks et al., 1994; Meyers et al., 1998; Baker et al., 2004). Correctly targeted ES clones were used to produce Bub1H/H mice. Intercrosses of Bub1H/H mice yielded viable Bub1–/H offspring with normal Mendelian frequency. Furthermore, interbreeding of Bub1H/H and Bub1+/- mice yielded viable Bub1–/– offsprings at the expected Mendelian frequency. Additionally, mice with Bub1+/-, Bub1H/H, and Bub1–/– mice exhibited no changes in development or appearance when compared with wild-type mice. We performed Western blotting to measure the level of wild-type Bub1 protein in mouse embryonic fibroblasts (MEFs) derived from Bub1+/-, Bub1+/-, Bub1H/H, and Bub1–/– mice (Fig. 1, D and E). We assessed that Bub1 signals from Bub1+/-, Bub1+/-, Bub1H/H, and Bub1–/– MEFs were ~75%, 50%, 30%, and 20% of those from Bub1+/- MEFs.
Figure 2. Proper targeting of Mad1 to kinetochores is highly sensitive to Bub1 down-regulation. MEFs with various levels of Bub1 expression were analyzed for the proper localization of proteins whose association with kinetochores or centromeres is known to be Bub1 dependent. (A) Fluorescent images of Bub1+/+, Bub1+/−, and Bub1−/− prophase cells stained for kinetochores, Bub1, and DNA showing that the gradual reduction of cellular Bub1 protein levels corresponds with a gradual decline in kinetochore-associated Bub1 protein. (B) Images of Bub1+/+, Bub1+/−, and Bub1−/− prometaphase cells stained for kinetochores, Mad1, and DNA demonstrating that kinetochore targeting of Mad1 is highly sensitive to Bub1 down-regulation. (C) Fluorescent images of prometaphase cells of the indicated genotypes stained for kinetochores, BubR1, and DNA showing that BubR1 localization to kinetochores is severely perturbed in Bub1−/− MEFs but not in Bub1+/− MEFs. (D) Fluorescent images of prometaphase cells of the indicated genotypes stained for kinetochores, CENP-E, and DNA demonstrating that CENP-E localization to kinetochores is impaired in Bub1−/− MEFs but not in Bub1+/− MEFs. (E) Fluorescent images of
respectively. Truncated forms of Bub1 encoded by the – and H alleles were undetectable even after overexpression of the Western blots (Fig. S1, available at http://www.jcb.org/cgi/content/full/jcb.200706015/DC1). Together, these results demonstrated that we had produced a series of mice with decreasing Bub1 protein dosage.

Mice with low amounts of Bub1 have a high percentage of aneuploid cells

To determine whether the reduced expression of Bub1 protein affects the accuracy of chromosome segregation, we collected splenocytes from Bub1+/+, Bub1−/H, Bub1+/−, and Bub1−/− mice at 5 mo of age and prepared metaphase spreads for karyotype analyses. Chromosome counts showed that <1% of wild-type splenocytes were aneuploid (Table I). In contrast, splenocytes from Bub1+/+, Bub1+/−, Bub1−/−, and Bub1−/− mice had a 6%, 16%, 35%, and 39% incidence of aneuploidy, respectively, revealing an inverse correlation between the level of Bub1 protein and the percentage of aneuploidy in this cell type. Moreover, the range of abnormal chromosome numbers broadened with the decreasing expression of Bub1 protein (Table I). We observed the premature separation of sister chromatids (PMSCS) in 14 and 15% of the mitotic figures from Bub1+/− and Bub1−/− splenocytes but only in 4% of the mitotic figures from Bub1−/− splenocytes (Table I). Thus, there seems to be no clear link between PMSCS and Bub1 dosage in splenocytes.

We further investigated the effect of Bub1 insufficiency on chromosome number stability by performing chromosome counts on metaphase spreads from Bub1+/+, Bub1+/−, Bub1+/−, Bub1−/+, and Bub1−/− MEFs at passage 5. We found that the percentage of aneuploid metaphases was much higher in Bub1−/− and Bub1−/− MEFs than in Bub1+/− and Bub1−/− MEFs, which, in turn, had a higher percentage than Bub1+/− MEFs (Table II). PMSCS was not increased in Bub1+/−, Bub1+/−, Bub1−/+, and Bub1−/− MEFs compared with Bub1+/+ MEFs (Table II). These data confirm that a high percentage of cells with low levels of Bub1 become aneuploid without the apparent requirement of PMSCS.

Kinetochore-associated proteins require distinct Bub1 levels for proper localization

Many of Bub1’s critical functions during mitosis occur at the kinetochore. Therefore, we tested whether the graded reduction of Bub1 expression corresponds to a graded reduction in Bub1 levels at kinetochores. Immunostaining of Bub1+/+, Bub1+/−, and Bub1−/− MEFs with affinity-purified Bub1-specific antibody showed that fluorescence signals at kinetochores progressively declined with decreasing cellular levels of Bub1 expression (Fig. 2 A). To examine how this graded reduction in kinetochore-bound Bub1 affected the localization of mitotic checkpoint proteins whose targeting to kinetochores is Bub1 dependent, we immunostained Bub1+/+, Bub1+/−, and Bub1−/− MEFs with antibodies against the mitotic checkpoint proteins Mad1, Mad2, BubR1, and CENP-E. In Bub1+/− prometaphase cells, Mad1 staining was concentrated on kinetochores visualized by anti-kinetochore antibody (Fig. 2 B). However, kinetochore-associated Mad1 signals were much less abundant in the corresponding Bub1−/− and Bub1−− cells. As Mad1 is required for the kinetochore localization of Mad2 (Chen et al., 1998, 1999), we anticipated that Mad2 staining patterns would also be reduced in Bub1−/− and Bub1−− prometaphase cells. We tested this prediction, but despite numerous attempts, we were unsuccessful in obtaining kinetochore-associated Mad2 signals in Bub1+/− and Bub1−/− MEFs with antibodies that are known to detect Mad2 at kinetochores of human prometaphase cells (see Materials and methods for details). Unlike Mad1, kinetochore-associated BubR1 and CENP-E signals were unaffected in Bub1−/− cells during prometaphase (Fig. 2, C and D). However, kinetochore signals for both of these proteins were dramatically reduced in prometaphase Bub1−/− cells. Western blot analysis showed that Mad1, BubR1, and CENP-E protein levels were similar in Bub1+/− and Bub1−/− cells (Fig. S2 A, available at http://www.jcb.org/cgi/content/full/jcb.200706015/DC1), excluding the possibility that the reduction in kinetochore localization of these proteins in Bub1−/− cells is caused by reduced protein stability.

Next, we tested how the graded reduction in kinetochore-bound Bub1 levels affected the subcellular localization of Sgo1 and Aurora B, both of which have been reported to require Bub1 for their proper localization to the centromeres (Tang et al., 2004b, 2006; Kitajima et al., 2005; Boyarchuk et al., 2007). Fewer Sgo1-positive centromeres were observed in Bub1−/− prometaphases than in Bub1+/− prometaphases (Fig. 2 E). In contrast, no such decrease was observed in Bub1+/− prometaphase cells (unpublished data). Immunostainings for Aurora B revealed that the localization of this protein was normal in both Bub1−/− and Bub1−/− prometaphase cells (Fig. S2 B). Thus, whereas most proteins that require Bub1 for proper localization to kinetochores/centromeres are mislocalized in Bub1−/− MEFs, only Mad1 is mislocalized in Bub1−/− cells.

Bub1 insufficient cells have a weakened mitotic checkpoint

To analyze the activity of the mitotic checkpoint in MEFs with a graded reduction in Bub1 expression, we performed a nocodazole challenge assay (Jeganathan et al., 2005; Baker et al., 2006). In this assay, MEFs were first transduced with a retrovirus encoding a YFP-tagged H2B fusion protein to allow the visualization of chromosomes by fluorescence microscopy (Fig. 3 A). MEFs were then challenged with nocodazole, and 20–30 cells undergoing nuclear envelope breakdown (NEBD) were marked and monitored at 15-min intervals to determine when their chromatin decondenses. The duration of arrest in mitosis, which is defined as the interval between NEBD (onset of mitosis) and chromatin decondensation (exit from mitosis without cytokinesis), was calculated and plotted. The time at which 50% of the cells have exited mitosis was used for comparison. Nocodazole-challenged Bub1+/+ MEFs typically remained arrested in

Bub1+/− and Bub1−/− prometaphase cells stained for kinetochores and Sgo1 showing that considerably fewer Sgo1-positive centromeres are present in Bub1−/− MEFs. Bar, 10 μM.
prometaphase for 7.2 h (Fig. 3B). Bub1<sup>+/+</sup> and Bub1<sup>+/−</sup> MEFs were impaired in their ability to maintain this arrest, with 50% of the cells exiting around 5.4 h. However, Bub1<sup>+/+</sup> and Bub1<sup>±/−</sup> MEFs exhibited a more profound defect, with 50% of the cells exiting mitosis at 3.8 h and 3.5 h, respectively. Thus, the mitotic checkpoint appears to be considerably weaker in Bub1<sup>+/+</sup> and Bub1<sup>±/−</sup> MEFs than in Bub1<sup>+/+</sup> and Bub1<sup>+/−</sup> MEFs.

**Bub1 insufficiency causes various chromosome segregation defects**

Next, MEFs with graded reductions in Bub1 expression were screened for chromosome segregation defects. In essence, we followed YFP-H2B-positive MEFs through an unchallenged mitosis by live cell imaging and determined the fraction of mitotic cells with chromosome segregation abnormalities. Two known defects underlying chromosome missegregation, congression failure and chromosome lagging (Fig. 4A and B), were observed at higher rates in Bub1<sup>+/+</sup>, Bub1<sup>+/−</sup>, Bub1<sup>+/−</sup>, and Bub1<sup>−/−</sup> MEFs than in Bub1<sup>+/+</sup> MEFs (Table III). The combined incidence of the aforementioned defects was remarkably similar in Bub1<sup>+/+</sup>, Bub1<sup>+/−</sup>, Bub1<sup>+/−</sup>, and Bub1<sup>−/−</sup> MEFs (Table III). Furthermore, anaphases with centrophilic chromosomes that segregate faster than the other chromosomes (Fig. 4C) were observed at an approximately two- to fivefold higher frequency in Bub1<sup>+/+</sup>, Bub1<sup>+/−</sup>, Bub1<sup>+/−</sup>, and Bub1<sup>−/−</sup> MEFs than in Bub1<sup>+/+</sup> MEFs (Table III). Whether this type of abnormality leads to chromosome missegregation is unclear, but even with the inclusion of this defect, the overall incidence of chromosome segregation abnormalities remains very similar in Bub1<sup>+/+</sup>, Bub1<sup>+/−</sup>, Bub1<sup>+/−</sup>, and Bub1<sup>−/−</sup> MEFs (Table III). Irrespective of Bub1 genotype, most cells with abnormal chromosome segregation events involved a single chromosome (or a duplicated chromosome). Occasionally, two or three chromosomes were implicated (Table IV and Table S1, available at http://www.jcb.org/cgi/content/full/jcb.200706015/DC1). Thus, the aforementioned analyses suggest that the accuracy of...
chromosome segregation is highly dependent on a full complement of Bub1 protein and that both small and large reductions in Bub1 cause chromosome missegregation at comparable rates.

Reduced cell death after chromosome missegregation as Bub1 levels decline

Initially, we were surprised that chromosome missegregation rates were similar in Bub1+/H, Bub1−/+, Bub1+/H, and Bub1−/− MEFs because the percentage of aneuploid cells was much higher in Bub1+/H and Bub1−/− cultures than in Bub1+/+ and Bub1+/+ cultures (Table II). One explanation could be that cell survival after chromosome missegregation increases with decreasing Bub1 levels. To explore this possibility, we infected Bub1+/+, Bub1+/−, Bub1+/H, and Bub1−/− MEFs with the YFP-H2B virus and monitored the fate of cells undergoing chromosome missegregation for up to 12 h by live cell imaging. Typically, 95% of Bub1+/+ MEFs died within several hours after chromosome missegregation (Table V and Videos 5 and 6, available at http://www.jcb.org/cgi/content/full/jcb.200706015/DC1). In addition, decreased Bub1 expression also did not increase survival to prolonged exposure to nocodazole, a spindle poison that induces tetraploidization by driving prometaphase cells into G1 without chromosome segregation (Fig. S4 D). These experiments suggest a rather specific role for Bub1 in mediating cell death after the missegregation of one or a few chromosomes.

Consistent with this interpretation, we found that micronuclei, which we observed by live cell imaging to result from misaligned, centripetal, or lagging chromosomes, accumulated steadily with decreasing Bub1 levels (Fig. S3).

To explore whether Bub1 plays a more general role in cell death signaling, we measured cell survival to various kinds of DNA-damaging agents. MEFs with graded reduction in Bub1 protein levels were exposed to increasing concentrations of doxorubicin, mitomycin C, or paraquat for 48 h. Cell survival was then determined by using the MTS assay. Cell survival in these agents was similar for Bub1+/+, Bub1+/−, Bub1+/H, Bub1+/H, and Bub1−/− MEFs (Fig. S4, A–C; available at http://www.jcb .org/cgi/content/full/jcb.200706015/DC1). In addition, decreased Bub1 expression also did not increase survival to prolonged exposure to nocodazole, a spindle poison that induces tetraploidization by driving prometaphase cells into G1 without chromosome segregation (Fig. S4 D). These experiments suggest a rather specific role for Bub1 in mediating cell death after the missegregation of one or a few chromosomes.

Spontaneous tumorigenesis is increased in Bub1 hypomorphic mice

To determine the long-term consequences of Bub1 down-regulation, we created and monitored cohorts of Bub1+/+(n = 160), Bub1+/−(n = 142), Bub1+/H(n = 137), and Bub1−/−(n = 238) mice on a mixed 129 × C57BL/6 background. Earlier, we reported that BubR1 hypomorphic mice have a short lifespan, are infertile, and develop various early aging–associated phenotypes (Baker et al., 2004). We note that no such phenotypes were observed in any of our Bub1 mutant mice (unpublished data). However, we found that Bub1−/− and Bub1+/H mice were significantly more prone to spontaneous tumors than Bub1+/+.

<table>
<thead>
<tr>
<th>MEF genotype (n)</th>
<th>Mitotic cells inspected</th>
<th>Metaphases with misaligned chromosomes</th>
<th>Anaphases with lagging chromosomes</th>
<th>Anaphases with centripetal chromosomes</th>
<th>Cells with segregation defects</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bub1+/−(5)</td>
<td>106</td>
<td>0.9</td>
<td>3.8</td>
<td>0.9</td>
<td>4.7 (5.6)</td>
</tr>
<tr>
<td>Bub1+/−/−(4)</td>
<td>91</td>
<td>3.3</td>
<td>6.6</td>
<td>3.3</td>
<td>9.9 (13.2)</td>
</tr>
<tr>
<td>Bub1+/−/−/(6)</td>
<td>142</td>
<td>7.7</td>
<td>6.3</td>
<td>2.1</td>
<td>12 (14.1)</td>
</tr>
<tr>
<td>Bub1+/−/−(4)</td>
<td>122</td>
<td>6.5</td>
<td>4.9</td>
<td>4.8</td>
<td>11.6 (16.4)</td>
</tr>
<tr>
<td>Bub1+/−/−(5)</td>
<td>168</td>
<td>7.1</td>
<td>6.6</td>
<td>2.4</td>
<td>13.1 (15.5)</td>
</tr>
</tbody>
</table>

All cells scored as metaphases with misaligned chromosomes displayed congression failure at anaphase onset. 

*Percentage of cells with misaligned and/or lagging chromosomes. The percentage of cells with chromosome segregation abnormalities is presented in parentheses.

Table III. Analysis of chromosome segregation abnormalities in Bub1-insufficient MEFs

<table>
<thead>
<tr>
<th>Mitotic figures inspected (n)</th>
<th>Number of cells with segregation defectsa</th>
<th>Number of cells with one abnormally segregated chromosome</th>
<th>Number of cells with two or three abnormally segregated chromosomesb</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bub1+/−/− (106)</td>
<td>6</td>
<td>6 (100%)</td>
<td>0 (0%)</td>
</tr>
<tr>
<td>Bub1+/−/− (91)</td>
<td>12</td>
<td>11 (92%)</td>
<td>1 (8%)</td>
</tr>
<tr>
<td>Bub1+/−/− (142)</td>
<td>20</td>
<td>15 (75%)</td>
<td>5 (25%)</td>
</tr>
<tr>
<td>Bub1+/−/− (122)</td>
<td>20</td>
<td>17 (85%)</td>
<td>3 (15%)</td>
</tr>
<tr>
<td>Bub1+/−/− (168)</td>
<td>26</td>
<td>21 (81%)</td>
<td>5 (19%)</td>
</tr>
</tbody>
</table>

aThe chromosomes involved were either single chromosomes or duplicated chromosomes.
bNone of the cells had more than three abnormally segregated chromosomes (see Table S1).
mice (Fig. 6, A and B). Bub1–/– mice had a significantly shorter median tumor-free survival (530 d) than Bub1H/H mice (676 d), which, in turn, had a significantly shorter median tumor-free survival than Bub1+/+ mice (772 d; Fig. 6 B). Moreover, Bub1–/– and Bub1H/H mice developed a different spectrum of tumors than did Bub1+/+ mice (Fig. 6, C–F). Bub1–/– mice developed significantly more sarcomas, lymphomas, and lung tumors. Bub1H/H mice were also prone to develop sarcomas but not lymphomas and lung tumors. Bub1H/H mice were also prone to develop sarcomas but not lymphomas and lung tumors. Bub1H/H mice were highly susceptible to hepatocellular carcinomas, a tumor type that was not significantly increased in Bub1–/– mice. In contrast to Bub1–/– and Bub1H/H mice, Bub1+/– mice showed a trend toward decreased tumor formation, particularly in liver and lung tissue (Fig. 6, B and C). Collectively, these data establish a causal relationship between the down-regulation of Bub1 expression and cancer development and suggest that there is a threshold level of Bub1 below which the incidence of neoplastic transformation progressively increases. Our data further imply that Bub1 reductions above the threshold may slightly inhibit tumor formation in particular tissues.

**Increased incidence of DMBA-induced tumors in Bub1 mutant mice**

Based on the aforementioned data, we conclude that Bub1+/– mice have enough Bub1 protein to protect themselves against spontaneous tumorigenesis. To determine whether this level is sufficient to guard against carcinogen-induced tumors, we administered a single dose of 0.5% DMBA (9,10-dimethylbenz-A-athracene) in acetone to the dorsal skin of 3–5-d-old pups generated from Bub1+/– × Bub1+/+ intercrosses. 5 mo after

### Table V. Cell death after chromosome missegregation decreases as Bub1 expression declines

<table>
<thead>
<tr>
<th>MEF genotype (n)</th>
<th>Cells with missegregation monitored</th>
<th>Apoptosis incidence</th>
<th>Apoptosis incidence (1 cell)</th>
<th>Apoptosis incidence (2 cells)</th>
<th>Cells with normal mitosis monitored</th>
<th>Apoptosis incidence</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bub1+/+ (3)</td>
<td>16</td>
<td>94</td>
<td>0</td>
<td>100</td>
<td>38</td>
<td>3</td>
</tr>
<tr>
<td>Bub1+/– (3)</td>
<td>28</td>
<td>68</td>
<td>26</td>
<td>74</td>
<td>17</td>
<td>6</td>
</tr>
<tr>
<td>Bub1–/– (3)</td>
<td>23</td>
<td>65</td>
<td>9</td>
<td>91</td>
<td>7</td>
<td>0</td>
</tr>
<tr>
<td>Bub1H/H (3)</td>
<td>26</td>
<td>42</td>
<td>12</td>
<td>88</td>
<td>16</td>
<td>0</td>
</tr>
<tr>
<td>Bub1–/– (3)</td>
<td>31</td>
<td>32</td>
<td>14</td>
<td>86</td>
<td>14</td>
<td>0</td>
</tr>
</tbody>
</table>

*Cells analyzed had either one or a few missegregated chromosomes.
treatment, we killed the mice and screened for tumors. Irrespective of the genotype, tumors were exclusively detectable in the lungs. Bub1<sup>−/−</sup> mice exhibited a two- to threefold higher incidence of lung tumors than in Bub1<sup>+/+</sup> mice (Fig. 7 A). Moreover, the tumor burden of Bub1<sup>−/−</sup> mice was increased approximately threefold (Fig. 7 B). From this experiment, we conclude that Bub1 heterozygous knockout mice are prone to carcinogen-induced tumorigenesis.

### Discussion

In this study, we produced a series of mutant mice in which the expression of Bub1 is reduced in a graded fashion from normal to zero by the use of wild-type, hypomorphic, and knockout alleles to determine the physiological role of Bub1. As anticipated, we find that the complete loss of Bub1 leads to embryonic lethality. Strongly reduced Bub1 expression (up to approximately...
When Bub1 is down-regulated. We speculate that Mad2, lar, the recruitment of Mad1 to kinetochores is dramatically protein for their normal recruitment to kinetochores. In particu-
lar, the recruitment of BubR1 and CENP-E to kinetochores is also sensitive to Bub1 down-regulation, but not as sensitive as Mad1, as their localization is normal in Bub1 heterozygous MEFs. Recent studies have presented evidence that Bub1 functions to recruit Sgo1 to centromeres to prevent the precocious separation of sister kinetochores (Tang et al., 2004b; Kitajima et al., 2005). Consistent with these studies, we find that centromeric Sgo1 levels are reduced in Bub1 mutant MEFs, but only when Bub1 expression is strongly down-regulated. However, this did not result in premature sister kinetochore separation, implying that an even further drop in centromeric Sgo1 is required to trigger the cleavage of cohesin molecules that link sister centromeres. A recent study has implicated Bub1 in the targeting of chromosomal passenger complexes to centromeres in early mitosis (Boyarchuk et al., 2007). Our finding that very low amounts of Bub1 are sufficient for directing these complexes to centromeres suggests that near complete Bub1 depletion is required to dislocate the passenger complex from mitotic centromeres.

One of our more surprising findings is the observation that a relatively small reduction in Bub1 expression has a major impact on the accuracy of chromosome congression. What could be the explanation for this observation? Although the precise role of Bub1 in chromosome congression is currently not known, it is believed that this role involves kinetochore assembly (Meraldi and Sorger, 2005). Of the mitotic checkpoint proteins whose recruitment is Bub1 dependent, only CENP-E has so far been implicated in chromosome congression. Thus, one explanation for the congression failure in Bub1 mutant MEFs might be a CENP-E recruitment defect. Consistent with this, we find that the targeting of CENP-E to kinetochores is perturbed in Bub1 hypomorphic MEFs. On the other hand, Bub1 haplo-insufficient MEFs, which display similar rates of congression failure as Bub1 hypomorphic MEFs, exhibit normal CENP-E recruitment to kinetochores, implying that the mechanism of congression failure is CENP-E independent. This conclusion is further supported by data of Meraldi and Sorger (2005) demonstrating that the depletion of Bub1 in HeLa cells by RNA interference causes chromosome congression defects in the absence of CENP-E mislocalization. Therefore, it remains unclear how Bub1 promotes proper chromosome congression. Nonetheless, we suspect that it involves a known or novel kinetochore-associated protein that functions in microtubule capture and whose recruitment to kinetochores is highly dependent on a full complement of Bub1.

Our analysis of MEFs with graded reduction in Bub1 expression indicates that relatively small shortages in Bub1, such as those seen in Bub1+/− and Bub1−/− MEFs, weaken the mitotic checkpoint considerably. It is plausible that the impaired recruitment of Mad1 (and presumably Mad2) to kinetochores undermines the mitotic checkpoint in these cells, as kinetochore-associated Mad1–Mad2 complexes generate soluble Mad2–Cdc20 complexes that bind to and inactivate APC/C. Larger reductions in Bub1, as present in Bub110H and Bub1−/− MEFs, had an even
more profound impact on mitotic checkpoint activity. We propose that this is caused, at least in part, by the added loss of CENP-E and BubR1 from kinetochores, as kinetochore-bound CENP-E and BubR1 molecules have been implicated in the assembly of various inhibitory protein complexes that target APC/C (Mao et al., 2003, 2005). The Bub1 kinase also can inhibit APC/C directly through the phosphorylation of Cdc20 (Chung and Chen, 2003; Chen, 2004; Tang et al., 2004a). We have not addressed whether the phosphorylation of Cdc20 is affected in our mutant series of MEFs as a result of the current lack of antibodies that recognize phosphorylated mouse Cdc20.

Although it has been well established that gross abnormalities of chromosome segregation (frequently referred to as mitotic catastrophe) often cause cell death (Castedo et al., 2004), the fate of cells undergoing the random missegregation of only one or a few chromosomes has been unknown. Here, we show by the use of live cell imaging that wild-type primary MEFs die at very high rates after minor abnormalities in chromosome segregation. The implication of this finding is that aneuploidy rates in cultured wild-type MEFs are substantially higher than metaphase spread karyotypes reveal. Our discovery that cell death rates after chromosome missegregation dramatically decline with decreasing levels of Bub1 creates a molecular entry point for studying the underlying cell death mechanism. Whether Bub1 plays a unique role in this mechanism or whether there is a broader connection between mitotic checkpoint damage and decreased cell death after chromosome missegregation is an important question for future analysis. Bub1’s dual function as a guardian of high fidelity chromosome segregation and as a mediator of cell death after aberrant segregation is reminiscent of proteins such as ATM (ataxia telangiectasia mutated) and p53 that function in both DNA repair and apoptosis in response to DNA damage (Sancar et al., 2004). A recent study showed that Bub1-depleted cancer cell lines display increased mitotic cell death when they are exposed to agents that perturb kinetochoremicrotubule attachment, such as nocodazole (Niikura et al., 2007). We observed no such effect in nocodazole-treated MEFs with graded reduction in Bub1 expression (Fig. S4 D), suggesting that the impact of the Bub1 level of expression on mitotic cell death induced by spindle poisons is cell type and/or transformation status dependent.

Bub1 expression is reduced in several human cancers, including colorectal, gastric, and esophageal tumors (Shigeishi et al., 2001; Shichiri et al., 2002; Doak et al., 2004); however, it was unknown whether the reduced expression of this mitotic checkpoint protein is causally implicated in tumorigenesis. Analysis of our series of Bub1 mutant mice firmly establishes that the reduced expression of Bub1 leads to the development of spontaneous tumors in mice, but only when Bub1 levels fall below a threshold level. Those with the most drastic reductions of Bub1 expression have the shortest tumor latency and the highest incidence of tumors. The level of Bub1 required to prevent spontaneous tumorigenesis appears to vary per tissue, as illustrated by the fact that only mice with the most profound reduction in Bub1 are predisposed to lymphomas and lung tumors. In the liver, the optimal level of Bub1 down-regulation is not the lowest level, as Bub1<sup>H10</sup> mice but not Bub1<sup>-40</sup> mice are prone to hepatocellular carcinomas. Adding even more complexity is the discovery that Bub1 haploinsufficiency exerts a slight tumor-suppressive effect in both liver and lung tissue. This finding is consistent with the recent discovery that CENP-E haploinsufficiency inhibits tumorigenesis in certain mouse tissues (Weaver et al., 2007). However, unlike CENP-E insufficiency, Bub1 haploinsufficiency does not inhibit DMBA-induced tumorigenesis. In fact, Bub1 haploinsufficient mice are highly susceptible to lung tumors when challenged with this carcinogen. This observation implies that the loss of one Bub1 gene copy acts to accelerate the development of tumors initiated by particular cancer gene mutations.

Because Bub1 hypomorphic mice have a high percentage of aneuploid cells and are predisposed to spontaneous tumors, whereas Bub1 haploinsufficient mice have a relatively low percentage of aneuploid cells and are not tumor prone, it is tempting to speculate that it is the increase in aneuploidy that drives tumorigenesis in Bub1 hypomorphic mice. However, the fact that both Rae1/Bub3 and Rae1/Nup98 double-haploinsufficient mice develop aneuploidy at rates very similar to that of Bub1 hypomorphic mice but are not prone to spontaneous tumors argues against this idea (Babu et al., 2003; Baker et al., 2006; Jeganathan et al., 2005, 2006). One possible explanation for this discrepancy could be that as a result of the decreased cell death in response to chromosome missegregation, Bub1 hypomorphic mice may develop a wider variety of abnormal karyotypes than Rae1/Bub3 and Rae1/Nup98 double-haploinsufficient mice, thereby perhaps increasing the incidence of karyotypes that have the ability to drive tumorigenesis. However, the role of aneuploidy in tumorigenesis is clearly highly complex, and it will be necessary to carefully examine each individual regulator of chromosome segregation for its involvement in tumorigenesis through the use of animal models. We expect these efforts to allow the identification of a subset of mitotic regulators that are particularly important for tumor prevention. Among them may be mitotic regulators that serve as molecular hubs within the mitotic checkpoint or other networks that regulate proper chromosome segregation or mitotic regulators with connectivity to other pathways that guard against neoplastic transformation.

In this study, we have used a series of mutant mice to demonstrate that only after reducing Bub1 levels beyond a threshold level do mice start to develop spontaneous tumors. Had we used only Bub1 haploinsufficient mice rather than a series of mice with graded reduction in Bub1 expression, our conclusions would have been dramatically different in that we would conclude that Bub1 does not act as a tumor suppressor itself. Heterozygous knockout models for several other mitotic checkpoint genes are also not predisposed to spontaneous tumorigenesis. For a more definitive understanding of the roles these genes have in tumor prevention, it will be useful to use hypomorphic alleles to further reduce their level of expression in mice.

Materials and methods

Generation of Bub1 mutant mice and analyses of tumorigenesis

An 8.5-kb Bub1<sup>129Sv/J</sup> genomic DNA fragment was used to generate both targeting vectors used. Gene-targeting procedures were performed as previously described (von Deursen et al., 1996). We identified targeted ES
cell clones by Southern blot analysis using a 3’ probe on BamHI-cut genomic DNA (Fig. 1 A). Mutant mice were derived from targeted ES cell clones through standard procedures. These mice were maintained on a mixed 129/Sv/E × C57BL/6 genetic background. Mice in tumor susceptibility experiments were observed daily for the development of overt tumors or signs of ill health. Moribund mice were killed, and all major organs were screened for overt tumors using a dissection microscope (SZX12; Olympus). Tumors that were collected were processed by standard procedures for histopathology. Prism software (GraphPad Software, Inc.) was used for the generation of tumor-free survival curves and for statistical analyses. DMSA treatment was performed as previously described (Serrano et al., 1996; Babu et al., 2003). All major organs were screened for overt tumors using a dissection microscope (SZX12; Olympus). Harvested tumors were routinely processed for histopathological confirmation. We note that all mice were housed in a pathogen-free barrier environment.

Western blot analysis and indirect immunofluorescence

Western blot analyses and indirect immunofluorescence were performed as previously described (Kasper et al., 1999). A laser-scanning microscope (LSM 510 v3.2SP2; Carl Zeiss MicroImaging, Inc.) as well as a microscope (Axiovert 100M; Carl Zeiss MicroImaging, Inc.) with a c-Apochromat 100× oil immersion objective (Carl Zeiss MicroImaging, Inc.) was used to analyze immunostained cells and to capture representative images. Primary antibodies were visualized with appropriate secondary antibodies conjugated to AlexaFluor594, -488, or -647 (Invitrogen). Western blot analysis and indirect immunofluorescence were housed in a pathogen-free barrier environment.

Karyotyping of MEFs and splenocytes

Chromosome counts on metaphase spreads were performed as previously described (Babu et al., 2003). We note that cells were scored as diploid (n = 40 chromosomes), tetraploid (n = 80 chromosomes), or aneuploid (Weaver et al., 2007).

Live cell imaging experiments

To allow the visualization of chromosomes by fluorescent microscopy on living cells, we used a retrovirus expressing YFP-tagged H2B (Jeganathan et al., 2006). Passage 2 MEFs were seeded in 6-well plates and cultured in DME/10% FBS at 3% oxygen. 12 h after seeding and again every 12 h for at least three times, the medium was replaced with medium harvested from EcoPACK pMSCV-puro-H2B-YFP viral producer cell lines. Cells were then seeded onto 35-mm glass-bottomed culture dishes (MatTek Corp.) and cultured in DME/10% FBS. Approximately 24 h later, experiments were performed using a microscope system (Axio Observer; Carl Zeiss MicroImaging, Inc.) with CO2, Module S, TempModule S, Heating Unit XL S, a plan Apo 63× NA 1.4 oil differential interference contrast III objective (Carl Zeiss MicroImaging, Inc.), camera (AxioCam MRm; Carl Zeiss MicroImaging, Inc.), and AxioVision 4.6 software (Carl Zeiss MicroImaging, Inc.). The imaging medium was DME/10% FBS. The temperature of the imaging medium was kept at 37°C. The exposure times in nocodazole challenge experiments were 100 ms at 2 × 2 binning. Time of arrest in mitosis was defined as the interval between NEBD (onset of mitosis) and chromatin decondensation (exit from mitosis without cytokinesis). Interframe intervals were 1.5 min for nocodazole challenge. Analysis of mitotic defects was performed as previously described (Baker et al., 2006). For analysis of the incidence of cell death after chromosome missegregation, MEFs undergoing abnormal chromosome segregation were marked and followed with an interframe interval of 30 min for up to 12 h. Cell death was preceded by severe nuclear blebbing and cytoplasmic fragmentation. For each of the aforementioned experiments, we examined at least three independent clones per genotype unless otherwise noted. Prism software (GraphPad Software, Inc.) was used for statistical analyses. To evaluate the incidence of micronuclei formation, at least 600 YFP-H2B–expressing interphase MEFs were screened for the presence of micronuclei by live cell microscopy.

Cell survival assays

Analyses of cell survival in response to doxorubicin, mitomycin C, and paraquat were performed as described previously (Baker et al., 2004) with the exception that passage 3 MEFs were used instead of passage 2 MEFs. For analysis of cell death in response to nocodazole treatment, 103 passage 3 MEFs were seeded in duplicate for three independent cell lines of each genotype. After ~12 h, normal medium was replaced with medium containing 100 ng/ml nocodazole, and cells were cultured for an additional 72 h. All cells were collected after this time, and trypsin blue exclusion was used to count living cells.

Online supplemental material

Fig. S1 shows that truncated proteins encoded by the Bub3 knockout and hypomorphic alleles are undetectable by immunoblotting. Fig. S2 shows that Mad1, Bub3, and CENP-E protein levels were similar in Bub1+/− and Bub1−/− cells and that Aurora B is not mislocalized in Bub1−/− cells. Fig. S3 shows that the incidence of micronuclei increases with declining levels of Bub1. Fig. S4 shows that the Bub1 level of expression has no impact on MEF cell survival to DNA-damaging agents and prolonged nocodazole exposure. Videos 1 and 2 show videos of the Bub1−/− MEF presented in Fig. S. A. Videos 3 and 4 show videos of the Bub1+/− MEF presented in Fig. S. B. Videos 5 and 6 show a Bub1+/− MEF undergoing chromosome missegregation in mitosis. Both daughter cells die after exit from mitosis. Videos 7 and 8 show a Bub1−/− MEF undergoing chromosome missegregation. One of the two daughter cells undergoes cell death. Videos 9 and 10 show a Bub1+/− MEF undergoing normal chromosome segregation. Both daughter cells survive. Table S1 presents data about the number of chromosomes that are abnormally segregated in cells with multiple segregation defects. Online supplemental material is available at http://www.jcb.org/cgi/content/full/jcb.200706015/DC1.

We thank Paul Galardy, Rick Bram, Robin Riche, and Debbie Pearson for critical reading of the manuscript and helpful discussions. We are grateful to Hongtao Yu, Tim Yen, and Don Cleveland for providing Sgo1, Mad1, and CENP-E antibodies, respectively. This work was supported by a National Institutes of Health grant to J.M. Van Deursen.

Submitted: 4 June 2007
Accepted: 19 September 2007

References


BUB1 INSUFFICIENCY CAUSES ANEUPLOIDY AND CANCER • JEGANATHAN ET AL. 267


