Dynein light chain regulates axonal trafficking and synaptic levels of Bassoon

Anna Fejtova,1 Daria Davydova,1 Ferdinand Bischof,1 Vesna Lazarevic,1 Wilko D. Altrock,1 Stefano Romorini,1 Cornelia Schöne,1 Werner Zschatter,2 Michael R. Kreutz,3 Craig C. Garner,4 Noam E. Ziv,5,6,7 and Eckart D. Gundelfinger1

1Department of Neurochemistry and Molecular Biology, 2Special Laboratory for Electron and Laserscanning Microscopy, and 3Project Group Neuroplasticity, Leibniz Institute for Neurobiology, D-39118 Magdeburg, Germany
4Nancy Pritzker Laboratory, Department of Psychiatry and Behavioral Science, Stanford University School of Medicine, Stanford, CA 94305
5The Rappaport Family Institute for Research in Medical Sciences, 6Department of Physiology and Biophysics, and 7The Lorry Lokey Interdisciplinary Center for Life Sciences and Engineering, Technion–Israel Institute of Technology, Haifa 32000, Israel

Bassoon and the related protein Piccolo are core components of the presynaptic cytomatrix at the active zone of neurotransmitter release. They are transported on Golgi-derived membranous organelles, called Piccolo-Bassoon transport vesicles (PTVs), from the neuronal soma to distal axonal locations, where they participate in assembling new synapses. Despite their net anterograde transport, PTVs move in both directions within the axon. How PTVs are linked to retrograde motors and the functional significance of their bidirectional transport are unclear. In this study, we report the direct interaction of Bassoon with dynein light chains (DLCs) DLC1 and DLC2, which potentially link PTVs to dynein and myosin V motor complexes. We demonstrate that Bassoon functions as a cargo adapter for retrograde transport and that disruption of the Bassoon–DLC interactions leads to impaired trafficking of Bassoon in neurons and affects the distribution of Bassoon and Piccolo among synapses. These findings reveal a novel function for Bassoon in trafficking and synaptic delivery of active zone material.

Introduction

In neurons, the transport of membranous organelles along axons is based on molecular motors that propel organelles along microtubules, which, in axons, are oriented uniformly, with their plus ends pointing toward the growing tips (Burton and Paige, 1981). Anterograde transport is driven by members of the kinesin family of molecular motors, whereas retrograde transport relies mainly on the cytoplasmic dynein 1 motor complex (Vale, 2003). The role of anterograde transport in trafficking of axonal proteins is well documented, and molecular motors carrying axonal cargos were identified (Hirokawa and Takemura, 2005). Two classes of vesicular cargos were studied in the context of presynaptic assembly in neurons; synaptic vesicle (SV) precursors transported by kinesin-3 heavy chain KIF1A (Okada et al., 1995) probably linked via the cargo adapter liprin-α (Shin et al., 2003; Miller et al., 2005), and Piccolo-Bassoon transport vesicles (PTVs) linked via the syntaxin1–syntabulin adapter complex to KIF5B, which is the heavy chain of conventional kinesin-1 (Cai et al., 2007). Imaging of most anterogradely transported axonal cargos reveals that they move bidirectionally (Schroer et al., 1985; Shapira et al., 2003; Miller et al., 2005), suggesting that most cargoes are able to associate with both anterograde and retrograde motors. However, little attention has been paid to the understanding of the molecular mechanisms and physiological meaning of retrograde transport of material predestined for delivery to distal axonal locations.

Bassoon and Piccolo (also named Aczonin) are highly homologous core components of CAZ (cytomatrix at the active zone; tom Dieck et al., 1998; Wang et al., 1999; Fenster et al., 2000). They are large scaffold proteins believed to functionally and spatially organize presynaptic neurotransmitter release (Fejtova and Gundelfinger, 2006; Leal-Ortiz et al., 2008). After synthesis, they associate with Golgi-derived membranous...
Bassoon can interact with DLC1 and DLC2

To identify novel proteins interacting with Bassoon, the cDNA fragment Bsn2 covering aa 609–1,692 of rat Bassoon (Fig. 1) was used as bait in a yeast two-hybrid screen. Seven independent positive clones carried the cDNA of DLC1. In subsequent experiments, the DLC-binding interface of Bassoon was narrowed down to fragment Bsn12 (aa 1,360–1,441) and Bsn9 (aa 1,441–1,692). Baits covering the other parts of Bassoon did not show any binding (Fig. 1 A). The binding region (fragment Bsn7), which is situated between two Piccolo-Bassoon homology domains, does not contain any predictable structural features. Moreover, an alignment of the amino acid sequence between PBH4 and PBH5 with the corresponding Piccolo region showed only 10% identity and 17.4% similarity. In line with this, overlapping constructs Pclo1 and Pclo2 covering the corresponding region of rat Piccolo (aa 1,878–2,729) did not interact with DLC1 in yeast, implying that the DLC1 interaction interface of Bassoon is not conserved in Piccolo (Fig. 1 B). DLC2 also interacted with fragments Bsn12 and Bsn9 but not with Pclo1 and Pclo2 in yeast.

To confirm the yeast two-hybrid analyses, an interaction of Bassoon’s DLC-binding fragments with DLC1 and DLC2 was examined in corecruitment assays after heterologous expression in COS-7 cells. GFP-tagged DLC1 or DLC2 was targeted to the outer mitochondrial membrane by fusing it with the mito-targeting sequence of the rat mitochondrial protein TOM20. Staining of transfected cells with MitoTracker demonstrated mitochondrial targeting of Mitof-EGFP-DLC1, Mitof-EGFP-DLC2, and the control fusion protein Mitof-EGFP (Fig. S1). Fragment Bsn7 fused to monomeric RFP (mRFP) exhibited a rather uniform distribution of organelles that are transported along axons to sites of nascent synaptic contacts (Zhai et al., 2001; Bresler et al., 2004; Dresbach et al., 2006; Tao-Cheng, 2007). It was shown that the accumulation of Bassoon and Piccolo at nascent synaptic junctions temporally correlates with activity-induced SV recycling and often precedes clustering of postsynaptic elements (Friedman et al., 2000; Zhai et al., 2000; Shapira et al., 2003). Thus, it was postulated that they might play an important role in the formation of presynaptic release sites early in synaptogenesis (Fejtova and Gundelfinger, 2006).

Dynein light chain (DLC) LC8 represents one of three dimeric light chains of the cytoplasmic dynein motor complex (Vallee et al., 2004; Pfister et al., 2005). In mammals, two DLC isoforms, DLC1 and DLC2, were reported to link cargoes to the dynein motor (Schnorrer et al., 2000; Navarro et al., 2004; Lee et al., 2006) to associate with the actin-dependent motor myosin V (Espindola et al., 2000), where it might also function as a cargo adapter (Puthalakath et al., 2001), and to have additional motor-independent cellular functions (Jaffrey and Snyder, 1996; Vadlamudi et al., 2004).

In this study, we describe an interaction of DLC1 and DLC2 with Bassoon and demonstrate that DLC-binding fragments of Bassoon function as cargo adapters for retrogradely moving organelles. Bassoon associates with the dynein motor complex in neurons, and disruption of Bassoon–DLC binding results in deficits in axonal trafficking of PTVs in living neurons. Our findings reveal a novel function of Bassoon (i.e., directly connecting PTVs to molecular motors), thus ensuring their active transport toward nascent synapses. Moreover, they provide new insights on the importance of bidirectional transport for appropriate cargo trafficking during synapse formation.

Figure 1. Mapping of the DLC-binding region on Bassoon. (A) Overview of Bassoon fragments tested for binding to DLC1 in yeast two-hybrid assays. The extension of positive Bsn clones is displayed in black, and nonbinding clones are indicated in gray. (B) The homologous region of Piccolo does not bind DLC. Amino acid residues encoded by individual subclones are given in parentheses. Zn1/2, zinc fingers; CC1–3, coiled-coil regions; PDZ, PSD-95/Dlg/zonula occludens-1 homology domain; C2A/B, C2 domains; PBH1–10, regions of the Piccolo-Bassoon homology domain (depicted as gray boxes).
Western blot (WB) analysis confirmed the successful coprecipitation of EGFP-Bsn7 with overexpressed DLC1 and DLC2 (Fig. 2 F) and with endogenous DLC (Fig. 2 G), demonstrating the existence of Bassoon–DLC complexes in living cells. In control experiments, we were unable to coimmunoprecipitate DLC1 or DLC2 when coexpressed with EGFP. Collectively, binding assays in yeast and mammalian cells demonstrate that Bassoon can interact with both DLC1 and DLC2 via an interaction interface not conserved in Piccolo.
AAA, was unable to bind DLC1 or DLC2, indicating that the putative DLC-binding motif present in this fragment was inactivated by the mutation. Of note, the replacement of the central Gln residue by Ala in all identified motifs only partially disrupted DLC binding (unpublished data).

Next, we tested whether each of the three DLC-binding sites is sufficient to mediate binding to Bassoon by means of the mitochondrial corecruitment assay. As expected, fragment Bsn9 was recruited to mitochondria when coexpressed with Mito-EGFP-DLC1 (Fig. S2, C and D) or Mito-EGFP-DLC2 (not depicted). The mutation of both TQT motifs disrupted the recruitment of Bsn9II,III but not of construct Bsn7II,III, which still contained the SQT motif functioning as the DLC interaction interface. Additional mutation of this motif (in Bsn7I,II,III) completely disrupted DLC binding of this fragment. Construct Bsn12, which exhibited binding in yeast, was not recruited to mitochondria in COS-7 cells expressing Mito-EGFP-DLC1 or Mito-EGFP-DLC2, presumably as the result of steric hindrance. The results of the yeast two-hybrid and corecruitment assays in COS-7 cells are tabulated in Fig. 3B. To further confirm the importance of identified binding motifs, EGFP-Bsn7, -Bsn9, and -Bsn13 and their mutants EGFP-Bsn7I,II,III, -Bsn7II,III, -Bsn9I,II,III, and -Bsn13I,II,III were expressed in HEK293T cells and precipitated with anti-GFP antibody. Coprecipitation of

### Figure 3. Mapping of three independently functional DLC-binding sites in Bassoon.

(A) Bassoon constructs including the amino acid substitutions introduced at interaction sites I, II, and III and their relative lengths; bordering amino acid numbers refer to rat Bassoon. (B) Summary of binding assays in yeast (yeast two-hybrid [Y2H]) and in corecruitment assay by mito-targeting in COS-7 cells (Mito). (C) Molecular-binding activities (MBA) of purified Bsn fragments to DLC1 and DLC2 in surface plasmon resonance assays. Error bars indicate SEM. **, P < 0.01. (D) Recombinant proteins used for the binding assays are shown on a Coomassie-stained SDS gel. Molecular mass is indicated in kilodaltons.

Identification of three functional DLC-binding motifs on Bassoon

Two independent Bassoon regions located on fragments Bsn9 and Bsn12 bind DLCs. To identify the exact DLC-binding site, we screened the amino acid sequence of both regions for sequences similar to published DLC-binding consensus motifs (K/R)XTQT (Lo et al., 2001) and GIQVD (Liang et al., 1999; Fan et al., 2001). Two TQT motifs are present in the sequence of the Bsn9 fragment. Short constructs with the sequences ATAEFSTQTQ and PMVAQGTVTQ, covering the first and second TQT motifs, respectively, can indeed bind DLC in yeast. The respective mutant constructs with TQT motifs replaced by AAA did not bind DLC in this assay (Fig. 3, A and B). To verify that there are no additional DLC-interacting sequence motifs in Bsn9, we generated mutants in which the first (Bsn9II), the second (Bsn9III), or both (Bsn9II,III) TQT motifs were replaced by AAA (Fig. 3 A). Yeast two-hybrid analysis showed that the mutation of both binding motifs was necessary and sufficient to disrupt the interaction between Bsn9 and DLC1 or DLC2. Assessment of colony number and size suggested that DLC binding of Bsn9III is impaired to a greater degree than that of Bsn9II, which showed binding basically undistinguishable from Bsn9. Fragment Bsn12 contains an SQT motif instead of the canonical TQT motif. Construct Bsn12I, in which SQT was replaced by AAA, was unable to bind DLC1 or DLC2, indicating that the putative DLC-binding motif present in this fragment was inactivated by the mutation. Of note, the replacement of the central Gln residue by Ala in all identified motifs only partially disrupted DLC binding (unpublished data).

Next, we tested whether each of the three DLC-binding sites is sufficient to mediate binding to Bassoon by means of the mitochondrial corecruitment assay. As expected, fragment Bsn9 was recruited to mitochondria when coexpressed with Mito-EGFP-DLC1 (Fig. S2, C and D) or Mito-EGFP-DLC2 (not depicted). The mutation of both TQT motifs disrupted the recruitment of Bsn9II,III but not of construct Bsn7II,III, which still contained the SQT motif functioning as the DLC interaction interface. Additional mutation of this motif (in Bsn7I,II,III) completely disrupted DLC binding of this fragment. Construct Bsn12, which exhibited binding in yeast, was not recruited to mitochondria in COS-7 cells expressing Mito-EGFP-DLC1 or Mito-EGFP-DLC2, presumably as the result of steric hindrance. The results of the yeast two-hybrid and corecruitment assays in COS-7 cells are tabulated in Fig. 3B. To further confirm the importance of identified binding motifs, EGFP-Bsn7, -Bsn9, and -Bsn13 and their mutants EGFP-Bsn7I,II,III, -Bsn7II,III, -Bsn9I,II,III, and -Bsn13I,II,III were expressed in HEK293T cells and precipitated with anti-GFP antibody. Coprecipitation of
overexpressed and endogenous DLC was detected using an anti-DLC antibody (Fig. 2 G and Fig. S2, A and B), confirming that mutation of all identified motifs is required to disrupt the Bassoon–DLC interaction.

Inspection of the primary structure of Bassoon orthologues in different vertebrate species showed an absolute conservation of the second and third DLC-binding motifs despite the divergence of the adjacent sequences (Fig. S3). The first motif was conserved only in birds and mammals but not in zebrafish. The high degree of conservation throughout vertebrate species suggests that the DLC–Bassoon interaction might be of functional significance.

DLC1 and DLC2 are very similar proteins with 93% sequence identity. However, it has been proposed that they are sequestered to distinct protein complexes, as shown for dynein or myosin V motor complexes (Naisbitt et al., 2000; Puthalakath et al., 2001). To assess a possible preference of the binding sites on Bassoon for the two DLC isoforms, we expressed and purified a set of His-thioredoxin fusion proteins covering the first, second, and third or all three binding motifs (Fig. 3 D) and tested their relative binding affinities to purified GST-DLC1 or GST-DLC2 (Fig. 3 C) using surface plasmon resonance technology. The results did not reveal any binding preference for single motifs, but we observed significantly higher binding of fragments Bsn9 (motifs II and III) and Bsn13 (all three motifs) to GST-DLC2 compared with GST-DLC1 (Fig. 3 C and Fig. S3 B). The assay also confirms that the interaction of Bassoon and DLC is direct. Notably, we observed an increase of the relative binding affinities starting with fragments Bsn9II and Bsn9III (containing one DLC-binding site) to Bsn9 (containing two) and finally Bsn13 (containing three binding sites; Fig. 3 C). Thereby, site I seems to bind more tightly than sites II and III, and the relative binding affinities seem to be additive as DLC binding of Bsn13 represented roughly the sum of that of Bsn9 and Bsn12. In agreement with this set of data, we find decreased amounts of DLC coprecipitating with EGFP-Bsn7II,III compared with EGFP-Bsn7 (Fig. S2 B). These observations imply that the arrangement of three DLC-binding motifs in close proximity facilitates binding of Bassoon to DLC in vivo.

**Bassoon-transporting vesicles associate with dynein and myosin V**

In young neurons, Bassoon is transported into the axon associated with membranous organelles, the PTVs (Zhai et al., 2001; Shapira et al., 2003; Dresbach et al., 2006; Tao-Cheng, 2007). One important cellular function of DLC is linking molecular motors to their cargo. Therefore, we wondered whether Bassoon already associates with DLC during its transport from the cell body toward nascent synapses and whether this interaction might be relevant for Bassoon’s delivery to the presynapse. To test this hypothesis, we cotransfected primary neurons with DLC1-EGFP and mRFP-Bsn at 3 d in vitro (DIV) and subjected them to live imaging 1 d later. Co-migration of Bassoon and DLC1–containing fluorescent puncta in both retrograde and anterograde direction (Fig. 4 B and Video 1) suggested cotransport of the two proteins.

In previous imaging experiments, we noted that mobile EGFP-tagged Bassoon puncta migrate in both directions along axons (Bresler et al., 2004), suggesting that anterograde and retrograde motors are involved. Anterograde transport of PTVs relies on kinesin-1 (Cai et al., 2007). In this study, we asked whether Bassoon transport organelles also recruit dynein- and/or myosin V–based motor complexes, both of which contain DLCs. To this end, we enriched PTVs by subcellular fractionation of embryonic brain homogenates (Zhai et al., 2001). Most Bassoon immunoreactivity was found between 0.8 and 1 M sucrose containing synaptic plasma membranes, but significant amounts were also present at the interface between 0.3 and 0.8 M sucrose containing floating PTVs (Fig. 5 A). In both membrane fractions, immunoreactivity was not observed if the samples were preincubated with Triton X-100, confirming that floating of Bassoon depends on its membrane association (Fig. 5 A). Intriguingly, DLC as well as IC74 (the intermediate chain of 74 kD) and...
dynein heavy chain (DHC), which are both exclusive components of dynein motor complexes, and myosin V were also detected in the 0.3–0.8 M sucrose interface (Fig. 5 A). Floating of all of these proteins was disrupted upon detergent treatment. This supports the view that Bassoon-carrying PTVs contain both dynein and myosin V motor complexes. To demonstrate conclusively the association of PTVs with multiple motors, we immunoisolated PTVs from the light membrane fraction using the Bassoon-specific monoclonal antibody mab7f (tom Dieck et al., 1998). As shown in Fig. 5 B, DLC, DHC, and myosin V were present in the fraction immunoisolated with mab7f but not with nonspecific IgGs. An antibody recognizing kinesin-1 heavy chains KIF5A/C (subclasses different from KIF5B, which was found on PTVs; Cai et al., 2007) did not detect significant levels of these proteins in the immunoisolated fraction (Fig. 5 B).

To examine the in situ colocalization of endogenous dynein motor complex components and PTVs, we performed immunostainings of neurons at 5 DIV with antibodies against Bassoon, DLC, IC74, and myosin V. A clear colocalization of Bassoon-positive puncta with DLC1/2, IC74, and myosin V was observed (Fig. 5 D). The degree of colocalization of Bassoon, DLC, IC74, and myosin V was quantified by determining the overlap coefficients (OCs) for each stain (Fig. S4) and comparing them with the overlap of Bassoon and cytochrome c, which labels mitochondria. The mean OC of coexisting for Bassoon and Piccolo (OC = 0.701 ± 0.017; n = 10), which is known to be cotransported on the same vesicle, was not significantly different from the OC of coexisting for Bassoon with DLC (OC = 0.661 ± 0.019; n = 21), IC74 (OC = 0.643 ± 0.015; n = 19), or myosin V (OC = 0.755 ± 0.029; n = 10). Similarly, the OC of coexisting for IC74 and DLC did not differ significantly (OC = 0.6614 ± 0.021; n = 9). In contrast, the OC of coexisting for Bassoon and cytochrome c was significantly lower (OC = 0.496 ± 0.021; n = 20; mean ± SEM; P < 0.0001). At the analyzed stage (5 DIV), neurons have formed only a few synapses, and the majority of Bassoon is associated with PTVs. Therefore, the colocalization of Bassoon-positive puncta with DLC, IC74, and myosin V in young neurons further supports the notion that these proteins are present on PTVs.

**Bassoon can function as a cargo adapter in COS-7 cells and associates with dynein motor complex in neurons**

When DLC1 or DLC2 were targeted to the outer mitochondrial membrane in COS-7 cells, the localization of mitochondrial was remarkably distinct from that of mitochondria in cells expressing the control construct Mito3-EGFP (Fig. 2, A1, B1, and D1). Normally, mitochondria are distributed throughout the cytoplasm of cells, sometimes even in their most distal regions (Figs. 2 A1 and 6 A1, arrows). In contrast, targeting of DLCs to mitochondria results in their accumulation near the cell center, presumably as the result of DLC-mediated retrograde transport along microtubules via dynein. To test whether Bassoon can function as a cargo adapter, we targeted the fragment Bsn9 to the outer mitochondrial membrane and observed the subcellular localization of mitochondria. After expression of Mito3-EGFP-Bsn9, mitochondria were clustered near the microtubule-organizing center (MTOC) of COS-7 cells, as visualized by costaining of microtubules (Fig. 6, B1 and B2). Targeting of the mutant fragment Bsn9DBM, which cannot bind DLC, to mitochondria did not alter their subcellular localization, confirming that an interaction with DLC was required for retrograde transport in COS-7 cells (Fig. 6 C1). Treatment of COS-7 cells expressing Mito3-EGFP-Bsn9 with the microtubule depolymerizing drug nocodazole for 2 h before fixation and staining led to a disruption of clusters, whereas treatment with the actin polymerization inhibitor cytochalasin D had no effect on cluster formation (Fig. 6, D1 and E1). These data suggest that fragment Bsn9 can operate as a cargo adapter via its binding to DLC and induces a microtubule-dependent retrograde transport of organelles in living cells.

To test whether Bassoon associates with the dynein motor complex in neurons, we performed immunoprecipitation experiments from the brain extract of newborn animals. Using Bassoon antibodies, we could coimmunoprecipitate DLC but not other components of the dynein motor complex, (i.e., IC74 or DHC; Fig. 4 C), suggesting that either the interaction is disrupted by the harsh conditions necessary to solubilize Bassoon or the major fraction of cellular Bassoon is not directly associated with motors but still binds DLC. To address this issue further, we immunoprecipitated the dynein motor complex using IC74 antibodies. In this study, significant amounts of Bassoon could be detected in the precipitates together with DHC and DLC (Fig. 5 C), confirming the association of a fraction of Bassoon with the dynein motor complex. Synaptophysin, representing an independent vesicular cargo in the brain at this developmental stage, was not coprecipitated with the dynein motor complex, and unspecific IgGs did not precipitate any of the analyzed proteins.

**Mutation of DLC-binding sites in Bassoon affects mobility of PTVs in axons**

The physical association of Bassoon with dynein motor complex suggests that the Bassoon–DLC interaction may mediate the trafficking of Bassoon-carrying vesicles within neurons. To test this, a GFP-tagged DLC-binding mutant (DBM) of Bassoon (GFP-BsnDBM) unable to interact with DLC was produced by introducing the aforementioned amino acid exchanges into all three DLC-binding motifs of Bassoon. We observed GFP-Bsn and GFP-BsnDBM particles in neurons transfected at 3 DIV in time-lapse experiments performed 48 h later. At this stage, moderate GFP signals were attained, and bidirectionally moving fluorescent particles could be observed in axons of cells transfected with either construct (Fig. 7 A). This implies that both GFP-Bsn and GFP-BsnDBM were sorted to appropriate transport vesicles. However, during a time period of 32 s (500 frames), a significantly larger fraction of immobile particles was observed in cells expressing GFP-BsnDBM as compared with wild-type Bassoon (72.5 ± 4.1% vs. 53.4 ± 4.1%; mean ± SEM; n = 6 cells and 534 and 450 puncta analyzed, respectively; P = 0.008; Fig. 7 B and Videos 2 and 3).

To determine velocities and running distances of vesicles carrying GFP-Bsn or GFP-BsnDBM, traces of mobile particles were visualized on kymographs of axonal segments with clear polarity. Traces showing processive movement (without stops and changes in velocities or movement directions) were analyzed.
Figure 5. DLC and other motor components associate with Bassoon transport packets.

(A) Membranes of rat embryonic brain (E18) were subjected to a floatation assay. Equal volumes of each fraction were analyzed on WBs. Membrane association was tested by incubation with the detergent Triton X-100 (+TX-100), which prevents floating of membrane-associated proteins. (B) PTVs were isolated from the light membrane fraction (0.3–0.8 M sucrose) by immunoprecipitation with Bassoon-specific antibodies, solubilized, and probed for their protein content on WBs (mBsn column). PTV marker proteins Bassoon (Bsn) and Piccolo (Pclo) were detected as well as DLC, DHC, and myosin V (myoV). KIF5A/C was not precipitated at detectable levels. Precipitation with unspecific IgGs (mlgG column) confirmed the specificity of binding. I, input; UB, unbound material; B, bound material.

(C) Coprecipitation of protein complexes with anti-Bassoon (αBsn column) and anti-IC74 (αIC74 column) antibody from brain lysate of P1 rats. The presence of DLC in Bassoon-containing complexes as well as the association of Bassoon with assembled dynein motor complex (containing IC74, DHC, and DLC) can be seen. Lanes with input and control precipitations with unrelated IgG are indicated. Synaptophysin (Sph) was not detected in any of the immunoprecipitated complexes. HC, heavy chain; and LC, light chain of coupled antibody.

(D) Costaining of Bassoon (magenta) with IC74, DLC1/2, and myosin V (green) in distal axons (left image sequences) and growth cones (right image sequences) of neurons at day 6 after plating. Outlines of axons and growth cones were created according to cell autofluorescences in raw images. Bar, 5 µm.
mobility and movement characteristics imply that Bassoon’s interaction with DLC regulates its axonal trafficking in neurons.

Interaction with DLC controls CAZ protein levels at the synapse

The impaired trafficking of GFP-BsnDBM in the live imaging experiments suggests that the inability to bind DLC might affect the supply of GFP-BsnDBM to synapses. To test this, we analyzed the localization of GFP-BsnDBM in relation to the endogenous pre- and postsynaptic markers synapsin and PSD-95 in 15-DIV neurons (Fig. 8 A). 98.8 ± 0.4% (mean ± SEM) of GFP-Bsn fluorescent puncta colocalized with synapsin, and 87.2 ± 2.5% colocalized with PSD-95. This was not significantly different from the localization of GFP-Bsn, where 94.5 ± 2.6% and 82.4 ± 2.8% fluorescent puncta colocalized with synapsin and PSD-95, respectively. Thus, the DLC-binding motifs of Bassoon are not absolutely required for its synaptic targeting.
DLC-binding sites II and III. This fragment was able to efficiently pull down DLC from brain lysates (Fig. S5 A). Moreover, trafficking of GFP-Bsn when coexpressed with the Bsn9 fragment (Fig. S5, B and C) very much resembled that of GFP-BsnDBM. To assess the impact of DLC-dependent Bassoon transport on its synaptic delivery, we analyzed amounts of synaptic proteins in 9-DIV neurons when many synapses are newly formed. To this end, we expressed Bsn9 together with EGFP-synapsin translated from the same bicistronic transcript to identify synapses formed by transfected neurons. The relative synaptic amounts of Bassoon were determined as the intensity of immunostaining normalized to mean Bassoon immunoreactivity of synapses formed by axons of nontransfected neurons in the same image. The relative synaptic amounts of Bassoon in cells expressing fragment Bsn9II,III (lacking the DLC-binding sites) with EGFP-synapsin were used as a reference. Thus, effects related to EGFP-synapsin overexpression or to the overexpression of functional elements encoded by Bsn9 that are not related to DLC binding were excluded. The quantitative analysis revealed a significant increase in mean amounts of Bassoon (1.75 ± 0.02 [n = 3,141] vs. 1.50 ± 0.02 [n = 1,827]; mean ± SEM; P < 0.0001; Fig. 8 B) and Piccolo (1.57 ± 0.03 [n = 1,420] vs. 1.21 ± 0.02 [n = 1,487]; mean ± SEM; P < 0.0001; Fig. 8 C) at synapses of neurons transfected with dominant-negative Bsn9 compared with Bsn9II,III. Notably, the relative intensities of Bassoon and Piccolo were much more variable between individual synapses compared with the control situation (Fig. S5, E, F, K, and L). Interestingly, synaptic levels of Munc13, a CAZ protein that can be transported to nascent synapses independently of PTVs (unpublished data), remained unaffected (1.47 ± 0.03 [n = 1,417] vs. 1.43 ± 0.03 [n = 1,038]; mean ± SEM; P = 0.23; Fig. 8 D). The amount of postsynaptic PSD-95 was also unchanged (1.40 ± 0.03 [n = 1,372] vs. 1.33 ± 0.03 [n = 1,441]; mean ± SEM; P = 0.07; Fig. 8 E). Surprisingly, synaptic levels of synaptophysin (1.568 ± 0.02707 [n = 2,109] vs. 1.712 ± 0.02689 [n = 2,399]; mean ± SEM; P = 0.0002; Fig. 8 F) and vGLUT1 (2.083 ± 0.05944 [n = 720] vs. 2.429 ± 0.07300 [n = 772]; mean ± SEM; P = 0.0003; Fig. 8 G) were slightly but significantly reduced upon overexpression of Bsn9 (Fig. S5, I, J, O, and L).

**Discussion**

We report a novel interaction between the presynaptic active zone protein Bassoon and DLC1 and DLC2. We identified three independently functioning DLC-binding sites in Bassoon and...
showed that these could function in linking cargo to the dynein motor complex in vivo. Interfering with the DLC–Bassoon interactions affects the trafficking of PTVs and the distribution of Bassoon and the related protein Piccolo among synapses, resulting in an increased variability in the Bassoon and Piccolo content of individual presynaptic boutons.

Three functional DLC-binding sites on Bassoon

We identified three independently functional DLC-binding sites on Bassoon, all resembling but not exactly matching the DLC-binding consensus sequence (K/R)XTQT (Lo et al., 2001). This is in line with the previously reported high diversity of binding sites identified among known DLC-interacting partners (Lajoix et al., 2004). Notably, none of three DLC-binding sites of Bassoon is conserved in its parologue Piccolo. Mutated Bassoon fragments, in which the essential (T/S)QT motifs were replaced by AAA, did not interact with DLCs neither in biochemical assays nor in yeast or mammalian cells. Therefore, it is likely that Bassoon binds DLCs via the target-interacting groove of DLCs, which is the interaction interface also shared by other known DLC interaction partners (Liang et al., 1999; Fan et al., 2001; Lo et al., 2001). Quantitative in vitro binding assays revealed that all three sites are active, that site I has a higher relative affinity for DLCs than sites II and III, and that binding strengths seem to be additive. Moreover, fragment Bsn7II,III, bearing only one DLC-binding motif, coprecipitated DLC from HEK293T cells less efficiently than fragment Bsn7, which bears all three. Thus, the cluster of three DLC-binding motifs constitutes a multivalent interaction interface in Bassoon that is likely to facilitate the DLC–Bassoon interaction.
In agreement with the high homology of DLC1 and DLC2, Bassoon was observed to bind both isoforms. However, quantitative binding assays with Bassoon fragments containing two or three DLC-binding interfaces showed significantly higher affinity for DLC2 than for DLC1. Because of the lack of DLC1- and DLC2-specific antibodies, we were unable to examine binding partner preferences of Bassoon in vivo.

One important cellular function of DLCs is to link their binding partners to dynein- or myosin V-dependent transport processes. In neurons, DLCs were shown to bind postsynaptic scaffold molecules like guanylate kinase-associated phosphatase (Naisbitt et al., 2000) or gephyrin (Fuhrmann et al., 2002), and active retrograde transport was proposed as a mechanism contributing to activity-dependent remodeling of the postsynaptic receptor apparatus (Maas et al., 2006). However, the functional role of DLCs as cargo adapters for dynein motors has become a subject of recent debate (Vallee et al., 2004; Barbar, 2008). The binding of DLC cargo proteins occurs in the same binding groove as the binding to dynein motor complex via IC74 (Williams et al., 2007). In this configuration, the DLC interaction partners would probably compete with the dynein motor complex for binding to DLC rather than being linked to the dynein motor complex via DLC. In this study, targeting of DLC-binding Bassoon fragments to mitochondria led to their accumulation near the cell center (probably the MTOC). This accumulation was dependent on both intact DLC-binding motifs and assembled microtubules and is best explained by assuming retrograde transport of Bassoon-tagged mitochondria. Similarly, a point mutation of the DLC-binding motif in the protein Egalitarian (Navarro et al., 2004) or deletion mutants of Swallow (Schnorrer et al., 2000) or Dazl (Lee et al., 2006) deficient in DLC binding showed defects in their dynein-mediated transport. In this study, we observed that Bassoon associates with the dynein motor complex in vivo and that the mutation of its DLC-binding motif has profound effects on microtubule-dependent transport when expressed in primary neurons. These findings support the view that Bassoon, via its interaction with DLCs, might function as a cargo adapter for the retrograde motor dynein.

**Bassoon links dynein and myosin motors to PTVs**

Recent studies established that newly synthesized Bassoon is transported toward nascent synapses associated with Golgi-derived vesicular structures, PTVs (Zhai et al., 2001; Shapira et al., 2003; Dresbach et al., 2006). Imaging studies using GFP-tagged Bassoon revealed that PTVs move along axons in both directions until they stop at sites of synaptogenesis (Shapira et al., 2003; Bresler et al., 2004). Fast long-range transport in axons is achieved by anterograde and retrograde movement of molecular motors along microtubules (Vale, 2003). Recently, the anterograde motor kinesin-1 was reported to link to PTVs via a syntaxin1–syntabulin–KIF5B adapter complex, mediating their transport toward synapses (Su et al., 2004; Cai et al., 2007). As the molecular link to retrograde transport remained unknown, we followed the hypothesis that interaction of Bassoon with DLC might be relevant in this context. Our data suggest that DLCs associate with mobile Bassoon packets, which also contain DHCs possibly linked through DLCs. Accordingly, the retrograde transport of PTVs along microtubules could be driven by the dynein motor complex recruited via Bassoon, although Bassoon is most likely not the only cargo adapter for dynein on PTVs. For example, voltage-dependent Ca channels, which were identified as potential components of PTVs (Shapira et al., 2003) and can interact with the DLC tctex-1 (Lai et al., 2005), might represent such an alternative cargo adapter.

Why are organelles such as PTVs, which are designated for axonal targeting, equipped with both anterograde and retrograde motors when unidirectional anterograde transport might be simpler and more economical? Regulated bidirectional movement of organelles might assure their relative uniform distribution in the outgrowing axon to be available for efficient synapse formation all along the axon.

**What might be the physiological role of the Bassoon-DLC interaction in neurons?**

To assess whether the Bassoon–DLC interaction links PTVs to microtubule-based axonal transport, the mobility of vesicles carrying either GFP-tagged wild-type Bassoon or DLC binding-deficient Bassoon were compared. Bidirectional movement observed in both cases implies that the perturbed DLC binding does not interfere with the appropriate sorting of Bassoon to transport packages, which are expected to also carry endogenous Bassoon. Nonetheless, a significant decrease in the overall mobility of GFP-BsnDBM-bearing vesicles as compared with GFP-Bsn–carrying vesicles was observed. A plausible explanation might be that bidirectional motors are tightly cross-regulated, i.e., when a motor for one direction is disturbed, the opposite motor does not simply take over. This is well demonstrated by mutants of kinesin heavy chain (Goldstein, 2001), kinesin light chain (Gindhart et al., 1998), or DHC (Martin et al., 1999), which result in the accumulation of their respective cargoes in axons but not strictly in their accumulation in the cell body or in distal part of axons. Moreover, it was hypothesized that the presence of multiple classes of motor proteins on one vesicle might cooperate to avoid collision when two vesicles are transported in opposite direction on the same microtubule track (Welte, 2004). Thus, the increased number of immobile particles may reflect the predisposition of Bassoon particles with perturbed links to dynein motor complexes to be involved in “traffic jams” on microtubule tracks.

The analysis of vesicle movements in retrograde and anterograde directions revealed significant decreases in retrograde and anterograde velocities as well as in running distances in both directions for DLC binding–deficient Bassoon compared with the wild-type GFP-Bassoon. Effects on the anterograde transport by interference with components of retrograde molecular motors like DHC or dynactin have also been reported in previous studies (Gross et al., 2000, 2002; He et al., 2005). This has been attributed to the necessity of efficient coordination of opposite-polarity motors by one another.

Despite the numerous immobile vesicles seen at 5 DIV in neurons expressing GFP-BsnDBM, normal localization of the construct with respect to pre- and postsynaptic markers was observed at 14 DIV, when synaptogenesis declines and PTVs are
reduced in cultured neurons (Zhai et al., 2000). However, interference with the Bassoon–DLC interaction leads to significant increase in mean Bassoon and Piccolo levels at individual synapses of transected neurons compared with controls. DLC-mediated transport of Bassoon and Piccolo away from synapses might be part of a protein exchange process fine-tuning synaptic amounts of these proteins (Tsuriel et al., 2006, 2009). Furthermore, our analysis revealed a striking increase in variability of both Bassoon and Piccolo contents at individual synapses. This confirms that the transport of both proteins is tightly linked and suggests that interference with Bassoon’s interaction with DLC causes a defect in the trafficking of their common carrier.

It was proposed that synaptogenesis occurs in two phases. Initially, SV precursors traveling along microtubule tracks pause or stop at locations predefined by yet unknown mechanisms. This leads to the formation of “orphan” transmitter release sites that can accumulate active zone proteins (Krueger et al., 2003; Sabo et al., 2006). In the second step, the initially formed site can split, and release-capable packages containing SVs and active zone material start to travel bidirectionally along axons (Krueger et al., 2003). Thus, the primary capturing of presynaptic scaffold molecules can occur at nascent synaptic sites in a random fashion. Their subsequent bidirectional active distribution among adjacent synaptic boutons might represent a mechanism assuring formation of a rather homogeneous synapse population. In line with this hypothesis, PTVs, after interference with Bassoon–DLC interactions, are stranded at the places where their transport paused because they cannot be transported further as a result of their impaired capacity to bind dynein. The high variance in synaptic levels of Bassoon and Piccolo could reflect the inability to more evenly redistribute the stochastically deposited clusters of PTVs during initial phases of synaptogenesis.

The levels of SV proteins synaptophysin and vGLUT1 were decreased in synapses of neurons overexpressing the DLC-capturing fragment Bsn9. The actual reason for this observation is currently unclear. As Piccolo was recently identified to negatively regulate SV exocytosis (Leal-Ortiz et al., 2008), the decrease of SV markers might be caused by increased levels of Piccolo. In the dominant-negative approach achieved by overexpression of Bsn9, synaptic levels of the CAZ protein Munc13 were not changed. This argues that interference with the Bassoon–DLC interaction did not have a general pleiotropic effect on axonal transport and is in line with described mechanisms of Munc13 synaptic targeting. Munc13 is recruited to synapses via the Rab3-interacting molecule (Andrews-Zwilling et al., 2006), which, in turn, is localized to synapses depending on the presence of CAZ-associated structural protein/ERC2 (Ohtsuka et al., 2002). Synaptic targeting of CAZ-associated structural protein is partly independent from Bassoon, which is consistent with the finding that it is not transported exclusively on PTVs (Ohtsuka et al., 2002). Preliminary experiments confirm that specific blockade of Bassoon’s exit from the trans-Golgi network does not disrupt the transport or appearance of Munc13 at synapses (unpublished data).

The amount of the postsynaptic marker PSD-95 also remained unchanged. As the size of pre- and postsynaptic elements is tightly coregulated (Schikorski and Stevens, 1999), additional Bassoon and Piccolo molecules detected in a subpopulation of synapses after expression of dominant-negative Bsn9 might not be inserted into presynaptic plasma membrane but accumulate within the presynaptic bouton. Perhaps PTVs are affected, after they dissociate from microtubules in synaptic areas, in their switch to myosin V driving their transport toward the presynaptic plasma membrane along actin filaments. Our immunostaining and immunoprecipitation experiments of PTVs confirm an association of myosin V with PTVs. Whether this switch from microtubule- to actin-based motors also involves DLC–Bassoon interactions will be the subject of future investigations.

Altogether, our experiments have shown that Bassoon interacts with DLCs in neurons, that this interaction is essential for proper movement of Bassoon transport packets along the axon, and that disruption of the interaction affects the accurate distribution of the CAZ scaffolding proteins Piccolo and Bassoon to the sites of neurotransmitter release.

Materials and methods

Cloning of constructs

The pMito3-EGFP vector was generated by introducing aa 1–33 of rat TOM20, coding for its mitochondrial targeting signal (Kanaji et al., 2000), into a eukaryotic expression vector, which was derived from pCMV-Tag3A (Agilent Technologies) by excision of the myc tag. The EGFP coding sequence was subsequently inserted in frame at the 3’ end of the TOM20 sequence. The GFP-Bsn construct used in this study was described previously as GFP-Bsn9s-3938 (Dresbach et al., 2003), and the mRFP-Bsn construct, in which GFP was replaced by mRFP, is its analogue. Cloning of Bsn1–4 fragments was described previously (Dresbach et al., 2003). All other Bassoon and Piccolo constructs were generated using PCR with extended primer, thereby adding BamHI and Xhol restriction sites at the 5’ and 3’ ends of the fragments, respectively. The introduced restriction sites were used for in-frame cloning of fragments into the vectors pGADT7, pGBK7, pEGFP-C2 (Clontech Laboratories, Inc.), pMito3-EGFP, and pmRFP-C2, which was constructed by introducing the mRFP sequence (gift from R.Y. Tsien, University of California, San Diego, La Jolla, CA) instead of the EGFP sequence into the pEGFP-C2 plasmid. The coding sequence of DCL1, flanked at its 5’ end with EcoRI and at its 3’ end with Xhol sites, was generated by PCR using extended primers. Accordingly, the coding sequence of DLC2 was flanked with BamHI (5’ end) and Xhol (3’ end). The introduced restriction sites were used for in-frame cloning of fragments into the plasmids pGADT7, pGBK7, pCMV-Tag2C (Agilent Technologies), pMito3-EGFP, pEGFP-N1 (Clontech Laboratories, Inc.), and pGEX-5X (GE Healthcare). The His-Strep expression vector pETS-32a+ was generated by ligation of a Strep-tagII (IBA) encoding synthetic DNA fragment with 5’ Xhol and 3’ Sall into the Xhol site of the plasmid pMAD32a+ (EMD). Bassoon fragments were cloned into the multiple cloning site of the resulting plasmid using appropriate restriction sites. Short fragments coding for the second and third DCL-binding interfaces of Bassoon were constructed by in-frame insertion of synthetic oligo-deoxynucleotides coding for the respective sequence into pGADT7 or pGBK7 vectors. All mutations described were introduced by inverse PCR using primers with mutated sequence and corresponding Bassoon fragments subcloned in pBluescript SK+ (Agilent Technologies) as a template. cDNA of EGFP-synapsin was a gift from T.A. Ryan (Cornell University, New York, NY).

Yeast two-hybrid experiments

For cDNA library screening, the Matchmaker Two-Hybrid System 2 (Clontech Laboratories, Inc.) was used with a rat brain Matchmaker CDNA library (Clontech Laboratories, Inc.) as prey and Bassoon fragment Bsn2 as bait. Transformation and selection was performed according to the manufacturer’s protocols. For narrowing down the DCL interaction region in Bassoon and for interaction analysis of Piccolo and mutated Bassoon construct, the Matchmaker system 3 (Clontech Laboratories, Inc.) was used. In detail, the Bassoon and Piccolo constructs based on pGBK7 were cotransformed with DCL1 or DCL2 in pGADT7 vectors into AH109 yeast cells using standard transformation protocols. Cotransformed cells were selected by growth on Leu- and Trp-lacking medium. The interaction of coexpressed proteins
activating expression of reporter genes was monitored as growth on selection media lacking adenine, His, Leu, and Trp and supplemented with 0.1% d-adenosine, 100 µM flavine adenine dinucleotide, 100 µM thiamine, 0.1% casamino acids, 0.1% yeast extract, 50 µM histidine, 50 µM methionine, 50 µM phenylalanine, 50 µM tryptophan, 0.05% glucose, 0.2% glycerol. Cells were incubated for 4 h at 37°C in 5% CO₂ atmosphere before exchange of growth media. Cells were lysed in 50 mM Tris-HCl, pH 7.4, and 0.5% Triton X-100, 100 mM NaCl, 10% glycerol, 1.5 mM MgCl₂, and complete protease inhibitors (Roche) for 1 h on ice before 48 h after transfection. Cleared cell lysate was used for immunoprecipitation using MicroMACS anti-GFP MicroBeads and MicroColumns (Millenyi Biotec) according to the manufacturer’s instructions, but using lysis buffer in all washing steps.

**Light brain membrane preparation, immunosolation of Bassoon-containing vesicles, and immunoprecipitation of Bassoon and the dynnein motor complex**

Light brain membrane preparation and vesicle immunosolation were performed as described previously (Zhai et al., 2001), with minor modifications. Embryonic day [E] 18 brains were homogenized in 5 mM MES, pH 7.0, 1 mM EDTA, and 0.32 M sucrose with complete protein inhibitors. The homogenate was centrifuged for 20 min at 15,000 g at 4°C. Mitochondria and crude membranes in the supernatant were lysed hypotonically by adding 9 vol of homogenization buffer without sucrose. The crude membrane extract was then centrifuged at 100,000 g for 1 h. The pellet was resuspended in homogenization buffer with 2 M sucrose and loaded as a layer of a discontinuous sucrose gradient under layers of 1, 0.8, and 0.3 M sucrose. The sucrose gradient was spun for 3 h at 350,000 g. Fractions were taken from the top of the gradient to the bottom. Immunoprecipitations were performed with superparamagnetic beads (Dynabeads Protein G; Invitrogen) according to the manufacturer’s protocol. In brief, mab7f or unspecific IgGs (50 µg/100 µl of beads) were bound to beads for 2 h at RT and, after being washed three times with PBS, were cross-linked using dimethyl pimelimidate. Beads were collected using a magnetic device (Dynal MPG; Invitrogen). Dynabeads coupled with mab7f, anti-IC74 antibody, or normal mouse IgGs were covalently coupled to 50 µl of GammaBind Plus Sepharose beads (GE Healthcare) using dimethyl pimelimidate. Postnatal day (P) 1 rat brains were homogenized in 25 mM Tris-Cl, pH 8.1, 50 mM NaCl, 2 mM EDTA, and 0.5% Triton X-100 plus complete protease and PhosSTOP phosphatase inhibitors cocktail (Roche). After centrifugation of 27,000 g for 15 min, the lysate was precleared by incubation with beads for 30 min, diluted 1:1 with NET gel (150 mM NaCl, 5 mM EDTA, 50 mM Tris, pH 7.4, 0.25% gelatin, 0.05% Triton X-100, and complete protease inhibitors cocktail), and, finally, incubated with antibody-coupled beads for 3 h. The beads were washed three times with buffer A (PBS, pH 7.4, 0.5% Triton X-100, 0.05% sodium deoxycholate, 0.01% sodium dodecyl sulfate, and 0.02% sodium azide) followed by three washes with buffer B (25 mM Tris, pH 8.1, 500 mM NaCl, 0.5% Triton X-100, 10 mM EDTA, and 0.02% sodium azide). The precipitates were eluted into SDS loading buffer. Full-downs were performed using purified Bassoon fragments bound to HisPur cobalt resin using the analogous homogenization and washing procedure as described for the aforementioned immunoprecipitation experiments.

**Neuronal cultures**

 Dissociated primary hippocampal cultures were prepared as described previously (Frischnecht et al., 2008). Transfection of cultured neurons was performed as described previously (Dresbach et al., 2003). Neurons were fixed in 4% paraformaldehyde in PBS, pH 7.4, for 5 min at RT. Before immunostaining, blocking was performed for 1 h in PBS containing 10% FCS, 0.1% Gli, and 0.3% Triton X-100. Primary and secondary antibodies were diluted in PBS containing 3% FCS. Images were taken with a confocal microscope (SP2; Leica) equipped with LCS software (Leica) or with an upright microscope (Axioplan2; Carl Zeiss, Inc.) equipped with a camera (Spot RTKE; Diagnostics, Instruments, Inc.) and MetaVue software (MDS Analytical Technologies). Nocodazole and cytochalasin D (both from EMD) were used in concentrations of 10 µM, and 1 mM, respectively. HEK293T cells grown in 75-cm² flasks were transfected using the Ca phosphate method. In brief, 0.5 ml of 0.5 M CaCl₂ were mixed with 25 µg DNA (for double transfection; 12.5 µg of each DNA). Then, 0.5 ml of 140 mM NaCl, 50 mM Hepes, and 1.5 mM Na₂PO₄, pH 7.05, were added and, after 1 min, and applied to cells in culture. The cells were incubated for 4 h at 37°C in 5% CO₂ atmosphere before exchange of growth media. Cells were lysed in 50 mM Tris-HCl, pH 7.4, 0.5% Triton X-100, 100 mM NaCl, 10% glycerol, 1.5 mM MgCl₂, and complete protease inhibitors (Roche) for 1 h on ice before 48 h after transfection. Cleared cell lysate was used for immunoprecipitation using MicroMACS anti-GFP MicroBeads and MicroColumns (Millenyi Biotec) according to the manufacturer’s instructions, but using lysis buffer in all washing steps.
shows how the DLC-binding motifs are conserved among Bassoon ortho-

Fig. S1 shows a map of pMito3-EGFP and the colocalization of Mito3-

performed with Prism 4 software (GraphPad Software, Inc.).

all statistical analyses were

maximal projections of confocal stacks were created, and synapse detection,

analyzed time frame (32 s corresponding to 500 frames) relative to the total de-

the percentage of particles that did not change their position within the ana-

colocalization between both channels. Immobile particles were determined as

using the JACoP plugin (Bolte and Cordelieres, 2006) of ImageJ. The OC ex-

In general, images were adjusted and analyzed using ImageJ (National Insti-

119 mM NaCl, 2.5 mM KCl, 2 mM CaCl2, 2 mM MgCl2, 30 mM glucose,

characteristics. Experiments were performed under constant perfusion with

fraction and with 10 frames/s for analysis of bidirectional vesicle movement

imaging of mRFP-Bassoon and DLC1-EGFP co-migration. Stream acquisition

MetaMorph Imaging software (MDS Analytical Technologies) using a 100×

Instruments) and a camera (QuantEM 512SC; Photometrics) controlled by

and a standard GFP or double GFP/mRFP excitation/emission filter set (Chroma

stream flow mode was used with 33 or 66 frames/s for analysis of the immobile vesicle

projection, and with 10 frames/s for analysis of bidirectional vesicle movement

34x25]354

JCB • VOLUME 185 • NUMBER 2 • 2009

Andrews­

Accepted: 24 March 2009

Submitted: 28 July 2008

We are grateful to H. Wickborn, J. Juhle, and B. Kracht for expert technical assistance, R. Frischknecht and M. Heine for helpful discussions and suggestions, T.A. Ryan for the EGFP-synapsin construct, and M. Sheng and R. Jahn for rabbit anti-DLC1/2 and synaptophysin antibodies, respectively.

This work has been supported by grants from the European Commission (SynScaff), the Land Saxony-Anhalt (LSA-N2), the European Union Structural Funds 2007–2013, and the Deutsche Forschungsgemeinschaft (SFB779/GK167) and a Max Planck Award from the Alexander von Humboldt Foundation/Max Planck Society to E.D. Guldengartner. This work was also supported by grants from the German Federal Government (BMFB/01GA0505) to M.R. Kreutz, E.D. Guldengartner, and N.E. Ziv and the National Institutes of Health (NS53947 and NS535862) to C.C. Garner. During part of the work, A. Fejtova was supported by a fellowship from the Swiss National Science Foundation.

Submitted: 28 July 2008
Accepted: 24 March 2009

References


Bolte, S., and F.P. Cordelieres. 2006. A guided tour into subcellular colocaliza-


Cai, Q., P.Y. Pan, and Z.H. Sheng. 2007. Syntabilin-kinses-1 family member 5B-mediated axonal transport contributes to activity-dependent pre-


Gross, S.P., M.A. Welte, S.M. Block, and E.F. Wieschaus. 2000. Dynem- 


Downloaded from jcb.rupress.org on July 10, 2017


**Figure S1.** Mitochondrial localization of fusion constructs using TOM20 mitochondrial targeting domain. (A) Map of expression vector for mito-targeted EGFP, DLC1, DLC2, and Bassoon fragments were cloned in frame using the multiple cloning site (MCS). bla, β-lactamase; CMV, cytomegalovirus; HSV-TK, herpes simplex virus thymidine kinase; neo/kan, neomycin/kanamycin; ori, origin of replication. (B–E) Cells were transfected with mito-targeting constructs, and, 18 h after transfection, mitochondria were stained using MitoTracker before fixation of cells. Colocalization of GFP fluorescence (first and third column) with MitoTracker (second and third column) confirmed correct targeting of expressed Mito3-EGFP and its fusion proteins. Note the change in distribution of mitochondria after mito-targeting of EGFP (B1–B3) in contrast to mito-targeting of DLCs (C1–C3 and D1–D3) or Bsn7 fragment (E1–E3). Bar, 20 µm.
Figure S2. **Bassoon interacts with DLC1 and DLC2 in mammalian cells.** (A and B) Coimmunoprecipitation of Bassoon–DLC complexes from HEK293T cells. Lysates from transfected cells with EGFP-tagged Bsn9 and Bsn13 and the point-mutated constructs Bsn9II,III and Bsn13I,II,III (A) or with Bsn7, Bsn7II,III, or EGFP (B) and Flag- (A) or myc-tagged (B) DLC1 were incubated with anti-GFP antibodies coupled to magnetic beads. The samples of input material (Input), unbound material (Unbound), and eluted material (Elution) were separated by SDS-PAGE and detected with antibodies against GFP (top), DLC8 antibody (A, bottom), or anti-DLC antibody (B, bottom) on WBs. All EGFP-tagged proteins and EGFP were expressed (I and Input lanes) and successfully immunoprecipitated (arrows in α-GFP WBs in A and B). Overexpressed DLC1 and endogenous DLCs were coimmunoprecipitated with EGFP-tagged Bsn9 and Bsn13 but not with the constructs carrying point mutations in their DLC-binding sites (A). EGFP-tagged Bsn7 coprecipitated overexpressed DLC1 and endogenous DLCs more efficiently than EGFP-Bsn7II,III, which contains only one active DLC-binding site (site I), whereas sites II and III were mutated. DLC was not coprecipitated with EGFP. Black/white lines indicate that intervening lanes have been spliced out. Molecular mass is indicated in kilodaltons. (C and D) Mito-targeting assays in COS-7 cells of Bsn9 fragment. Cells were fixed 18 h after transfection. Mito-targeted EGFP or EGFP-DLC1 are localized at mitochondria (C3 and D3; green in overlay images), as shown by colocalization with MitoTracker (C5 and D5; magenta in overlay images C4 and D4). Bassoon fragment Bsn9 (C1 and D1; magenta in overlay images C2 and D2) shows a uniform cytoplasmic distribution when coexpressed with Mito-EGFP control construct (C1–C3) but is corecruited to mitochondria when mito-targeted DLC1 is coexpressed (D1–D3). Bar, 10 µm.
**Figure S3. DLC-binding motifs of Bassoon.** (A) Conservation during vertebrate evolution. Alignment of amino acid sequences of human (hs), rat (rn), mouse (mm), platypus (oa), chicken (gg), and zebrafish (dr) Bassoon orthologues. The DLC-binding motifs are highlighted in gray. Note that the second and third motifs are completely conserved in all species displayed, whereas the first motif is not conserved in fish. Asterisks indicate identical residues, colons indicate conserved substitutions, and periods indicate semiconserved substitutions. (B) Results of surface plasmon resonance binding assays (Fig. 3 C). The molecular-binding activity (MBA) was calculated from three to five association–dissociation cycles performed for each interaction pair. The results shown were confirmed by a second measurement set in which proteins coming from independent purifications were used.

<table>
<thead>
<tr>
<th></th>
<th>DLC1 MBAx10⁻³ mean±SD</th>
<th>DLC2 MBAx10⁻³ mean±SD</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bsn 9</td>
<td>34.3±3.1</td>
<td>52.7±2.5</td>
</tr>
<tr>
<td>Bsn 9&lt;sup&gt;II&lt;/sup&gt;</td>
<td>19.3±1.2</td>
<td>17.7±3.5</td>
</tr>
<tr>
<td>Bsn 9&lt;sup&gt;III&lt;/sup&gt;</td>
<td>18.3±3.6</td>
<td>21.6±3.1</td>
</tr>
<tr>
<td>Bsn 9&lt;sup&gt;IV&lt;/sup&gt;</td>
<td>6.3±0.6</td>
<td>6.7±1.2</td>
</tr>
<tr>
<td>Bsn 12</td>
<td>50±7</td>
<td>44±5.3</td>
</tr>
<tr>
<td>Bsn 12&lt;sup&gt;I&lt;/sup&gt;</td>
<td>10.3±1.2</td>
<td>9.3±2.1</td>
</tr>
<tr>
<td>Bsn 13</td>
<td>91.3±6.3</td>
<td>110.3±4</td>
</tr>
<tr>
<td>Bsn 13&lt;sup&gt;III&lt;/sup&gt;</td>
<td>1.7±0.6</td>
<td>2±4</td>
</tr>
</tbody>
</table>
Figure S4. Quantification of colocalization of Bassoon with myosin V, IC74, DLC8, Piccolo, and cytochrome c and of DLC8 with IC74 in distal axonal segments of 5 DIV neurons. (A) Example images used for colocalization analysis are shown. (B) Bar graph of mean OC calculated from at least 10 independent images. Error bars indicate SEM; ***, P < 0.001 (one-way analysis of variance). Bsn, Bassoon; CytoC, cytochrome c; MyoV, myosin V; Pclo, Piccolo. Bar, 10 µm.
Figure S5. Interference with Bassoon–DLC interaction in neurons. (A) Pull-down experiment showing that endogenous DLC is efficiently captured by Bsn9 but not by Bsn9II,III (bearing point-mutated DLC-binding sites) from brain lysate. Molecular mass is indicated in kilodaltons. (B and C) The overexpression of Bsn9 affects GFP-Bsn trafficking in neurons. Running distances of movements in both directions were analyzed. (B) Bar graphs show mean values of running distances for GFP-Bsn with co-overexpressed Bsn9 (open bars) or Bsn9II,III (shaded bars). Error bars indicate SEM; **P < 0.01; ***P < 0.001. (C) Relative distribution of analyzed running distances (significant difference: P < 0.001 in two-way analysis of variance of cumulative frequency distribution of values). (D) Scheme illustrating the expression cassette used for interference with DLC–Bassoon interaction in a dominant-negative approach. Dominant-negative construct Bsn9 carrying two of Bassoon’s DLC-binding sites or control construct Bsn9II,III lacking these sites (see Fig. 3) was overexpressed in primary neurons transfected at 3 DIV and evaluated at 9 DIV. Synapses were labeled by coexpression of EGFP-synapsin from the same vector encoding a bicistronic transcript. CMV, promoter or cytomegalovirus; IRES, internal ribosomal entry site. (E–P) Plots show the cumulative distribution of relative fluorescent intensities for respective stainings. Note the different distribution of Bassoon and Piccolo staining intensities in cells transfected with Bsn9 construct as compared with the control Bsn9II,III. Scatter plots show distribution of values of intensities of individual synapses. Of note, the mean staining intensities and the variance of intensities at individual synapses (K and L) is increased for Bassoon and Piccolo but not for Munc13, PSD-95, or SV markers synaptophysin and vGLUT1.
Video 1. **DLC1-EGFP co-migrates with mRFP-Bsn in neurons.** The video shows a section of an axonal tree of a neuron transfected with DLC1-EGFP and mRFP-Bsn at 3 DIV and analyzed 2 d later. Recording of DLC1-EGFP fluorescence is shown in the left part and mRFP-Bsn fluorescence in the right part of the combined image. Note the co-migration of mRFP-Bsn particles with DLC1-EGFP (compare with Fig. 4 A). The video is shown at 30 frames/s.

Video 2. **Mobility of GFP-Bsn in neurons.** The video shows a section of an axonal tree of a neuron transfected with the GFP-Bsn construct at 3 DIV. The video was recorded 48 h after transfection. Note the smaller, dimmer fluorescent particles that move faster and more directionally than the bigger, brighter particles, which show more restricted forward and backward movements. The video is shown at 90 frames/s.

Video 3. **GFP-BsnDBM shows decreased mobility compared with GFP-Bsn in neurons.** The video shows a section of an axonal tree of a neuron transfected with the GFP-BsnDBM construct lacking all three DLC-binding sites at 3 DIV. The video was recorded 48 h after transfection. Note the striking difference in overall mobility of both smaller and bigger fluorescent particles as compared with Video 2. The video is shown at 90 frames/s.