Flotillin microdomains interact with the cortical cytoskeleton to control uropod formation and neutrophil recruitment

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Introduction

Microdomains defined by the proteins flotillin 1 and flotillin 2 (also termed reggie 2 and reggie 1, respectively) are an apparently ubiquitous feature of mammalian cells (Babuke and Tikkanen, 2007). They reside in the plasma membrane, as well as in endosomal and lysosomal organelles (Bickel et al., 1997; Lang et al., 1998; Volonte et al., 1999). The precise molecular function of flotillin microdomains is not known, but they are thought to be involved in processes including insulin signaling, T cell activation, phagocytosis, epidermal growth factor signaling, and regrowth of neurons (Babuke and Tikkanen, 2007; Langhorst et al., 2007; Hansen and Nichols, 2009). Non-exclusive models for how the microdomains act during these processes include mediation of endocytosis, scaffolding of signaling proteins, and interaction with the cortical cytoskeleton (Glebov et al., 2006; Kato et al., 2006; Babuke and Tikkanen, 2007; Langhorst et al., 2007).

Recently, it was shown that flotillin microdomains accumulate rapidly in the uropod of neutrophils after stimulation with the chemotactic bacterial peptide N-formyl-methionyl-leucyl-phenylalanine (fMLP; Rajendran et al., 2009; Rossy et al., 2009). The uropod is a plasma membrane protrusion found at the rear of motile leukocytes and other cell types (Sánchez-Madrid and Serrador, 2009). Generation of this structure requires segregation of different types of protein and lipid plasma membrane components, and local recruitment and activation of components of the cortical cytoskeleton (Gómez-Mouton et al., 2001; Seveau et al., 2001; Xu et al., 2003; Lee et al., 2004). The uropod of migrating leukocytes has been implicated in several functions, including intercellular signaling, regulation of cell adhesion, and migration through resistive environments. Whether flotillin microdomains are required for uropod morphogenesis and function has yet to be directly tested in loss-of-function experiments.

Here we use a combination of biochemistry, imaging, and generation of flotillin 1 knockout (−/−) mice to investigate the function of flotillin microdomains in neutrophils.
Results and discussion

Flotillin 1−/− mice lack flotillin microdomains
We produced C57BL6/J mice lacking exons 3–8 of the gene for flotillin 1 (Fig. 1, a and b). Western blots of tissues that express flotillin 1 in wild-type mice confirmed that flotillin 1 protein cannot be detected in flotillin 1−/− mice (Fig. 1 c). Heterozygous flotillin 1+−/− mice had reduced levels of flotillin 1 protein compared with controls. Flotillin 1−/− mice are viable, fertile, and have no readily apparent phenotypes.

Flotillin 1 and flotillin 2 function together to generate flotillin microdomains, and reduction in the expression of one flotillin protein causes a reduction in the expression of the other (Frick et al., 2007; Langhorst et al., 2008; Babuke et al., 2009). Western blotting of tissues from flotillin 1−/− mice confirmed that in the absence of flotillin 1 the expression of flotillin 2 is greatly reduced (Fig. 1 c). Mouse embryonic fibroblasts (MEFs) grown from flotillin 1−/− and isogenic control mice were fixed and stained with antibodies against flotillin 1 and flotillin 2. In the controls, flotillin 1 and flotillin 2 colocalized in plasma membrane puncta as predicted. In the flotillin 1−/− cells the limited, weak staining obtained with flotillin 2 antibodies was uniformly distributed across the plasma membrane (Fig. 1 d). Intact flotillin microdomains are insoluble in cold nonionic detergents (Bickel et al., 1997). Solubilization of extracts from flotillin 1−/− and wild-type control MEFs with cold Triton X-100 followed by ultracentrifugation in a sucrose density gradient revealed that the residual flotillin 2 found in these extracts is not present in the detergent-resistant fraction (Fig. 1 e). These observations show that deletion of flotillin 1 is likely to completely abrogate the function of flotillin microdomains.

Flotillin microdomains are required for neutrophil recruitment in vivo
We measured recruitment of neutrophils in vivo, using a cell recruitment model in which a sterile air pouch is introduced subcutaneously. This establishes a vascularized cavity that is ideal for monitoring cellular migration in response to chemotactic agents (Kadl et al., 2009). After injection of fMLP, the air pouch was lavaged at different time points, and cellular infiltration was assessed. Total cell counts were performed, and cytospin slides were prepared and stained with Wright-Giemsa. Reduced total numbers of cells accumulated in the air pouch in response to fMLP in flotillin 1−/− mice, relative to wild type (Fig. 2 a). Counting of the infiltrating cells on Giemsa-stained cytospins demonstrated that the reduced recruitment of cells into the air pouch of flotillin 1−/− mice was due to an early defect in neutrophil, monocyte, and lymphocyte recruitment, with the effect being most prominent in the case of neutrophils (Fig. 2, a and c).

To further confirm that fMLP-induced recruitment of neutrophils and monocytes is impaired in flotillin 1−/− mice, cells infiltrating the air pouch were characterized by flow cytometry (Fig. 2 b). Surface expression of Ly6G and CD11b, as well as forward/side scatter profiles, were used to identify neutrophils (Ly6G+CD11b++) and monocytes (Ly6G+CD11b−/++; Cooper et al., 1993; Taylor et al., 2002). Monocytes were confirmed by F4/80 expression (not depicted). We detected a striking decrease in neutrophil recruitment in flotillin 1−/− mice, with only 20% of control levels observed after 2 h (Fig. 2, b and c). We also detected fewer monocytes (P = 0.01) in the flotillin 1−/− mice (Fig. 2 c).

The basal populations of different cell types in the air pouch model before addition of fMLP were essentially the same in control and flotillin 1−/− mice (Fig. 2, a–c; zero time points). Hematological analysis of blood obtained by cardiac puncture from control and flotillin 1−/− mice showed that levels of circulating leukocytes are again the same in both cases (Fig. S1). The pronounced defect in the chemotaxis of flotillin 1−/− neutrophils into fMLP-containing skin pouches is, therefore, likely to be due to a role for flotillin microdomains specifically in the process of recruitment itself.

Flotillin microdomains interact with myosin IIa and spectrins
To gain insight into the molecular mechanisms underpinning the function of flotillin microdomains we immunosolated flotillin microdomains, using anti-GFP antibodies and a HeLa cell line stably expressing flotillin 2–GFP. A second cell line expressing GFP provided a negative control. Elution of the immunosolutes was performed in a three-step protocol, to consecutively isolate detergent-resistant regions of the plasma membrane containing flotillin microdomains (E2), then specific protein complexes containing the microdomains themselves (E3; Fig. 3 a). After Coomassie staining, protein bands that were not present in the negative control samples were identified by mass spectrometry (Fig. 3 b). The most abundant proteins found in E3 were flotillins 1 and 2, indicating that the immunosolation protocol works efficiently. Other abundant proteins found specifically associated with flotillin microdomains were α- and β-spectrins and myosin heavy chain 9, the heavy chain of myosin IIa (Conti and Adelstein, 2008; Vicente-Manzanares et al., 2009; Fig. 3 b).

Western blotting was used to confirm the specific association of α-spectrin and myosin IIa with flotillin microdomains, using the same sequential elution procedure. As predicted, both proteins were found enriched in the E3 fraction after immunosolation of flotillin 2–GFP from HeLa cells (Fig. 3 c). Caveolin 1 was not found to be directly associated with flotillin microdomains. Only trace amounts of actin were detected in the flotillin microdomain (E3) fraction, and abundant plasma membrane proteins including cadherins, syntaxin 2, and the transferrin receptor were not detected (Fig. 3 c). We conclude that flotillin microdomains are specifically associated with myosin IIa and both α- and β-spectrins.

Flotillin microdomains may represent a point of contact between the plasma membrane and the underlying cell cortex. To support this idea, we coexpressed flotillin 1–GFP, flotillin 2–GFP, and myosin IIa–mCherry in HeLa cells. As predicted, both proteins were found enriched in the E3 fraction after immunosolation of flotillin 2–GFP. A second cell line expressing GFP provided a negative control. Elution of the immunosolutes was performed in a three-step protocol, to consecutively isolate detergent-resistant regions of the plasma membrane containing flotillin microdomains (E2), then specific protein complexes containing the microdomains themselves (E3; Fig. 3 a). After Coomassie staining, protein bands that were not present in the negative control samples were identified by mass spectrometry (Fig. 3 b). The most abundant proteins found in E3 were flotillins 1 and 2, indicating that the immunosolation protocol works efficiently. Other abundant proteins found specifically associated with flotillin microdomains were α- and β-spectrins and myosin heavy chain 9, the heavy chain of myosin IIa (Conti and Adelstein, 2008; Vicente-Manzanares et al., 2009; Fig. 3 b).

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Figure 1. **Flotillin microdomains are absent in flotillin 1−/− mice.** (a) Gene targeting strategy. Positions of primers used for PCR genotyping are shown as black arrows; the primers are labeled a, b, c. (b) Genotyping of wild-type, C57Bl6/J flotillin 1+/+, flotillin 1 knockout C57Bl6/J flotillin 1−/− (−/−), and flotillin 1+/− heterozygous mice (+/−). (c) Flotillin 1−/− mice lack flotillin 1, and flotillin 2 expression in these mice is drastically reduced. Western blots using antibodies against flotillin 1 and flotillin 2. MEF, mouse embryonic fibroblast. (d) Residual flotillin 2 in flotillin 1−/− MEFs is not concentrated in flotillin microdomains. Staining of mixed culture of flotillin 1−/− and wild-type MEFs with flotillin 1 and flotillin 2 antibodies. Bar, 20 µm. (e) Residual flotillin 2 in flotillin 1−/− MEFs is not present in detergent-resistant membranes. Detergent-resistant membranes (fractions 9 and 10, boxed) were floated in a sucrose density gradient. Western blots of samples from control (+/+ and flotillin 1−/−−/−) mice were probed with antibodies to flotillin 2.
Staining of flotillin 2–GFP cells with anti-flotillin 1 antibodies showed that flotillin 1 expression is increased in these cells (Fig. S2). This suggests that exogenously expressed flotillin 2–GFP stabilizes endogenous flotillin 1. Flotillin 2–GFP cells frequently had altered morphology compared with untransfected controls, and the distribution (but not total expression) of myosin IIa was visibly altered (Fig. 4 b). The expression and distribution of α- or β-spectrin was not notably altered in these cells (not depicted). Staining of the flotillin 2–GFP cell lines with phosphospecific antibodies revealed a pronounced increase in myosin IIa RLC phosphorylation as compared with cocultured control HeLa cells (Fig. 4, c and d). Increased myosin IIa phosphorylation (Fig. 4 a, and Video 1). This provides direct evidence that flotillin microdomains are associated with the cell cortex in live cells.

We next performed experiments designed to assess whether flotillin microdomains can impact on the activity of myosin IIa. Phosphorylation of myosin IIa regulatory light chain (RLC) at Ser19 increases motor activity and plays a key role in the regulation of its cellular activity (Umemoto et al., 1989; Conti and Adelstein, 2008). Overexpressing flotillins, particularly flotillin 2, induces changes in cell morphology, and hence presumably in the distribution or activity of cortical proteins (Neumann-Giesen et al., 2004, 2007; Hoehne et al., 2005). We generated individual HeLa cell lines stably overexpressing flotillin 2–GFP. Staining of flotillin 2–GFP cells with anti-flotillin 1 antibodies showed that flotillin 1 expression is increased in these cells (Fig. S2). This suggests that exogenously expressed flotillin 2–GFP stabilizes endogenous flotillin 1. Flotillin 2–GFP cells frequently had altered morphology compared with untransfected controls, and the distribution (but not total expression) of myosin IIa was visibly altered (Fig. 4 b). The expression and distribution of α- or β-spectrin was not notably altered in these cells (not depicted). Staining of the flotillin 2–GFP cell lines with phosphospecific antibodies revealed a pronounced increase in myosin IIa RLC phosphorylation as compared with cocultured control HeLa cells (Fig. 4, c and d). Increased myosin IIa
Figure 3. **Flotillin microdomains associate with myosin IIa and spectrins.** (a) Protocol for isolation of flotillin microdomains. (b) Identification of proteins associated with flotillin microdomains in HeLa cells by mass spectrometry. The specific bands from Coomassie-stained gels indicated with arrows were identified by mass spectrometry. Numbers in brackets indicate the number of unique peptides identified. MW, molecular weight (kD). (c) Confirmation that α-spectrin and myosin IIa associate specifically with flotillin microdomains in HeLa cells. Immunoprecipitation was performed as in panels a and b above; samples were analyzed by Western blotting with antibodies against the proteins indicated.
Figure 4. Flotillin microdomains interact with the cell cortex in vivo, and regulate myosin IIa activity. (a) Co-migration of flotillin microdomains and myosin IIa. Time-lapse TIR imaging of a HeLa cell coexpressing flotillin–GFP and myosin IIa–mCherry. Times indicate interval after addition of 50 µM cytochalasin D. The yellow area highlights the region of the plasma membrane from which both flotillin microdomains and cortical cytoskeleton retract. Bar, 10 µm. (b) Stable overexpression of flotillin 2–GFP alters the distribution of myosin IIa. A HeLa cell line overexpressing flotillin 2–GFP was co-cultured with HeLa cells, and stained with antibodies against the heavy chain of myosin IIa. Bar, 25 µm. (c) Stable overexpression of flotillin 2–GFP increases phosphorylation of myosin IIa RLC on S19. (d) Histogram of myosin IIa RLC phosphorylation (S19) in control and flotillin 2–GFP clones. (e) Western blots of F2-GFP clones, control and treated samples. (f) Western blots of myosin IIa S19-P and actin.
RLC phosphorylation in the flotillin 2–GFP cell lines was confirmed by Western blotting (Fig. 4 e). Use of the myosin light chain kinase inhibitor ML7 (Chou et al., 2004), and the Rho kinase inhibitor Y27632 (Saito et al., 2002), confirmed the specificity of the antibodies and identified Rho kinase as being responsible for the bulk of the detected phosphorylation (Fig. 4 e). We used siRNAs to look at the effect of reduced flotillin 1 or flotillin 2 expression on phosphorylation of myosin IIa RLC in HeLa cells. Two separate siRNAs against each flotillin all caused a clear reduction in myosin IIa RLC phosphorylation (Fig. 4 f). Therefore, an increase in the abundance of flotillins is sufficient to increase the activity of myosin IIa, and loss of flotillins causes a reduction in activity.

Uropod formation and cell migration is compromised in the absence of flotillin microdomains

We sought to apply the data derived from experiments in HeLa cells described above to begin to characterize the functional deficit in neutrophils from flotillin 1/−/− mice in more detail. Both spectrins and myosin IIa, like flotillin microdomains, are found concentrated in uropods (Gregorio et al., 1992; Eddy et al., 2000; Jacobelli et al., 2009; Sánchez-Madrid and Serrador, 2009) and myosin IIa activity is required for uropod formation (Doyle et al., 2009; Jacobelli et al., 2009; Vicente-Manzanares et al., 2009). We asked, therefore, whether the absence of flotillin microdomains impacts on neutrophil migration ex vivo, uropod formation, or myosin IIa activity.

Chemotaxis assays following the migration of neutrophils from control and flotillin 1/−/− mice on serum-coated glass in a concentration gradient of fMLP, using Dunn chambers and video microscopy, did not detect any difference between the two populations of cells (Fig. S3, a and b). Additionally, phosphorylation of ERK (extracellular signal–regulated kinase), which acts downstream of the fMLP receptor (Coxon et al., 2003), occurs identically in control and flotillin 1/−/− cells (Fig. S3, c and d). We also tested the ability of control and flotillin 1/−/− neutrophils to migrate in Dunn chambers in response to a second chemoattractant, MIP-2 (macrophage inflammatory protein-2; Kobayashi, 2006). Again, no difference in behavior was detected (unpublished data). Flotillin microdomains are, therefore, neither required for neutrophils to sense different chemoattractants nor for locomotion of neutrophils on planar glass surfaces.

It is likely that there are different functional requirements for cells to migrate through resistive, three-dimensional environments than for migration on planar surfaces (Even-Ram and Yamada, 2005; Doyle et al., 2009). We assayed migration of neutrophils through Matrigel (a matrix of laminin, collagen, and other basement membrane components; Kleinman et al., 1982). Generation of projections based on confocal Z-stacks allowed measurement of the proportion of cells migrating into a layer of Matrigel in response to chemoattractant (Fig. 5 a). We found MIP-2 gave more reproducible results in this assays than fMLP. A marked reduction in the efficiency of migration into Matrigel was observed in flotillin 1/−/− neutrophils when compared with controls (Fig. 5, a and b). This suggests that the defect in neutrophil recruitment in flotillin 1/−/− mice reported in Fig. 2 is, at least in part, due to a cell-intrinsic effect of the lack of flotillin microdomains specifically in neutrophils.

To look for possible defects in neutrophil polarization and uropod formation, we fixed fMLP-stimulated neutrophils from control and flotillin 1/−/− mice, and stained them with antibodies against myosin IIa, α-spectrin, and against the uropod membrane marker CD44 (del Pozo et al., 1995; Fig. 5, c and d; spectrin staining shown in Fig. S3 e). The number of uropods was quantified using CD44 as a marker. Flotillin 1/−/− neutrophils had a well-defined uropod half as frequently as was observed in the controls (Fig. 5 e).

To analyze whether myosin IIa activity was altered in flotillin 1/−/− neutrophils we used phosphospecific antibodies to measure phosphorylation of myosin IIa RLC after stimulation with fMLP. Staining with the phosphospecific antibodies was efficiently concentrated in the uropod (Fig. 5 f). Quantification of fluorescence intensity revealed an ∼50% drop in signal in flotillin 1/−/− neutrophils stimulated with fMLP for 15 min (Fig. 5 g). Reduced myosin IIa RLC phosphorylation in flotillin 1/−/− neutrophils during stimulation with fMLP was confirmed by Western blotting (Fig. 5, h and i).

**Conclusion**

The data presented here demonstrate a requirement for flotillin microdomains during neutrophil recruitment in vivo. Our data also reveal that one of the functions of flotillin microdomains is to coordinate formation of the uropod in neutrophils; that, at least in part, this function probably involves regulation of the activity of myosin IIa; and that flotillins are required for efficient migration of neutrophils through Matrigel.

Our previous studies, and those of other laboratories, suggest that flotillins define a specific mechanism for endocytosis (Payne et al., 2007; Blanchet et al., 2008; Schneider et al., 2008; Babuke et al., 2009; Riento et al., 2009; Zhang et al., 2009). It is intriguing that immunoisolation of flotillin microdomains and analysis of associated proteins did not reveal any abundant membrane proteins. The function of flotillin-mediated endocytosis thus remains unclear. We speculate that one function may be to regulate the number and distribution of flotillin microdomains within the plasma membrane, as this could provide a way of controlling plasma membrane–cortex interactions in space and time.
Figure 5. Flotillin microdomains are required for uropod formation and activation of myosin IIa. (a) Migration of control and flotillin 1<sup>−/−</sup> neutrophils into Matrigel in a gradient of MIP2. The dashed white line indicates the top of the Matrigel layer. Triangles represent the direction of the MIP2 concentration gradient; vertical bar, 50 µm. Images are maximum intensity projections derived from 20–25 confocal Z-slices. (b) Quantification of neutrophil migration into Matrigel. The proportion of cells entering the Matrigel was derived by quantifying the fluorescence above and below the dashed white line in panel a. Combined data from four separate experiments are shown. (c) Indirect immunofluorescence with antibodies against flotillin 1 and α-spectrin to show colocalization in the uropod of fMLP-stimulated neutrophils. Bar, 20 µm. (d) Comparison of uropod formation in control (+/+) and flotillin 1<sup>−/−</sup> neutrophils. Cells were fixed 15 min after stimulation with fMLP and labeled with antibodies against CD44. Arrows indicate structures defined as uropods. Bar, 20 µm.
The biochemical association between flotillin microdomains and IIa RLC. (i) Densitometric quantification of Western blot of lysates from control (+/+) and lin 1
dependent of uropod formation (Doyle et al., 2009).

Matrices implies that myosin II has roles in cell migration in-fiber fibroblasts (which do not form uropods) to migrate through the other. Indeed, the finding that myosin II activity is needed for fibroblasts (which do not form uropods) to migrate through matrices implies that myosin II has roles in cell migration independent of uropod formation (Doyle et al., 2009).

The reduction in myosin IIa phosphorylation in both flotillin I−/− neutrophils and in flotillin siRNA-treated HeLa cells, the biochemical association between flotillin microdomains and myosin IIa, and the effect of overexpression of flotillins on myosin IIa phosphorylation all indicate that flotillin microdomains are important in regulation of myosin IIa activity. Spectrin function in uropod formation is not well characterized, but, as these proteins may restrict lateral diffusion in the plasma membrane, recruitment of spectrins could play an important role in establishment of the uropod as a distinct membrane domain (Gregorio et al., 1992; Seaveu et al., 2001; Tang and Edidin, 2003; Sánchez-Madrid and Serrador, 2009). We propose that flotillin microdomains regulate uropod formation by spatially organizing the plasma membrane and the cortical cytoskeleton during neutrophil polarization, thereby allowing activation of myosin IIa and other uropod components at one specific location in the cell cortex.

Materials and methods

Mice

All animal experiments were performed in compliance with Irish Department of Health and Children regulations and approved by the Trinity College Dublin’s BioResources ethical review board, or with the approval of the UK Home Office and MRC-UMB ethical review board. Flotillin I−/− C57Bl6/J mice were generated by Artemis GmbH. Gene targeting was performed in C57Bl6/J embryonic stem cells. The selectable cassette used for gene targeting (Fig. 1 a) was removed by crossing with C57Bl6/J mice to establish the uropod as a distinct membrane contrast (black and white levels) was adjusted in Adobe Photoshop without

Image acquisition and processing

All fluorescence images were obtained using a confocal microscope (LSM 510; Carl Zeiss, Inc.), 63x 1.4 NA objective, and standard filter sets, with the exception of Fig. 4 a and Video 1. These were obtained using a total internal reflection microscope (Olympus), 100x 1.49 NA objective. Image contrast (black and white levels) was adjusted in Adobe Photoshop without gamma adjustment. Fluorescence intensities were quantified using ImageJ (NIH, Bethesda, MD).

Isolation of neutrophils

Primary mouse neutrophils were isolated from bone marrow of age- and gender-matched mice. In brief, bone marrow cells were harvested into HBSS, 15 mM Hepes, pH 7.2, and 0.05% FAF-BSA. The cell suspension (10 ml) was loaded onto a discontinuous 60/70% Percoll gradient (in HBSS, 10 ml each) and spun at 1,300 g for 30 min at 4°C. Neutrophils were collected from the 60/70% Percoll interface and washed with 50 ml HBSS, 15 mM Hepes, pH 7.2, and 0.05% FAF-BSA. Red blood cells were lysed by hypotonic shock and the final neutrophil cell pellet washed twice and resuspended in HBSS plus Ca2+ and Mg2+, 15 mM Hepes, pH 7.2, and 0.05% FAF-BSA.

fMLP time course in neutrophils

3 x 106 neutrophils in HBSS plus Ca2+ and Mg2+, 15 mM Hepes, pH 7.2, and 0.05% FAF-BSA were equilibrated to 37°C in a heat block and stimulated with 1 μM fMLP for the indicated time points. Cells were spun at 6,000 g for 1 min at 4°C and immediately lysed in RIPA buffer for 30 min on ice. Lysates were cleared of debris by centrifugation and precipitated using MeOH/Chloroform. Equal protein amounts were subjected to Western blotting.

Immunofluorescence

HeLa cells stably expressing flotillin 2–GFP or GFP were scraped into PBS, pelleted, and homogenized in 2 ml ice-cold HB (320 mM sucrose, 1 mM EDTA, 2 mM MgCl2, 20 mM tricine, pH 7.8, plus protease inhibitor cocktail; Roche) using a Dounce homogenizer. A post-nuclear supernatant (PNS) was generated by centrifugation of the homogenate at 1,000 g for 10 min. The PNS was buffered by the addition of one-tenth volume of 10x PBS.

Phospho-myosin light chain 2 (Ser19; 3671) and phospho-myosin light chain 2 (Thr18/Ser19; 3674) antibodies were from Cell Signaling Technology. Anti-transferrin receptor antibody (13-6800) was from Invitrogen. Mouse anti-phospho-Erk1/2 (M8159) and polyclonal anti-actin (A2066) antibodies were from Sigma-Aldrich.

Cell culture and immunofluorescence

HeLa cells and mouse embryonic fibroblasts were cultured in DME, 10% fetal bovine serum (FBS), and penicillin/streptomycin at 37°C, 10% CO2. For immunofluorescence, cells were fixed with 4% paraformaldehyde for 3–4 min followed by 6 min in −20°C methanol (MeOH). Cells were stained with primary and secondary antibodies in PBS, pH 7.4, 5% FBS, and 0.2% saponin.

Primary mouse neutrophils were stimulated with fMLP suspended in Heps-buffered saline solution (HBSS), 15 mM Hepes, pH 7.2, and 0.05% fatty acid-free bovine serum albumin (FAF-BSA), and allowed to settle for the time indicated (typically 15 min) onto FBS-coated coverslips. They were then fixed and processed as described for HeLa cells above.

siRNA knockdown in HeLa cells

HeLa cells were transfected with 50 nM flotillin siRNAs using Oligo-lectamine [Invitrogen] according to the manufacturer’s recommendations. The following siRNAs were used: F1.2: 5′-GCAGAGAGGCACAAAU3′; F1.2: 5′-GUUGCCACUGUACACUGA3′; F2.2: 5′-UGAUUGGGUGG-GCACCCCA3′; F2.2: 5′-AGGAGAGGGUGACCGGGAU3′. All siRNAs were ordered from Thermo Fisher Scientific. Cells were lysed with RIPA buffer 72 h after transfection and lysates precipitated with MeOH/Chloroform. Equal protein amounts were subjected to Western blotting.
Chemotaxis assays

For chemotaxis in Dunn chambers, 50,000 bone marrow–derived neutrophils from age- and gender-matched flotillin 1−/− and flotillin 1+/+ mice were allowed to adhere to glass coverslips for 10 min at room temperature in chemotaxis buffer (HBSS, 15 mM NaCl, 1% NP-40, 1% sodium deoxycholate, and 0.1% SDS) and incubation on ice for 30 min. The lysates were cleared by centrifugation and the samples precipitated using MeOH/CHCl3.

Western blotting of tissue samples

Tissues from flotillin 1+/+ and flotillin 1−/− littersmates were homogenized in HB using a Dounce homogenizer. Equal protein amounts were solubilized by addition of RIPA buffer (25 mM Tris, pH 7.6, 150 mM NaCl, 1% NP-40, 1% sodium deoxycholate, and 0.1% SDS) and incubation on ice for 30 min. The lysate was mixed thoroughly and left on ice for 30 min with occasional mixing. Nuclei were pelleted at 1,000 × g for 5 min. The lysate was adjusted to 40% sucrose by addition of 2 ml 80% sucrose in TNE and placed on the bottom of an ultracentrifuge tube. 6 ml 35% sucrose and 2 ml 5% sucrose (in TNE) was layered on top and the gradient spun at 50,000 rpm (SW40 rotor; Beckman Coulter) for 16 h. Twelve 1-ml fractions were collected from the bottom by tube puncture and precipitated using MeOH/CHCl3.

Chemokine assays

Cells were stained with eBioscience mAbs; APC anti-CD11b (Mac-1 α; ICRF44), PE anti-Ly6G (RB6-8C5), and PerCP anti-F4/80 (BM8). Flow buffers used contained 2 mM EDTA to exclude doublets. Using appropriate isotype controls, gates were drawn and data were plotted on logarithmic scale density plots or dot plots.

Slide preparation and differential cell counting

Slides were prepared from the lavage fluid (50,000 cells/slide) from air pouches using a cytospin (ThermoShandon). Blood smears were prepared after cardiac puncture. All slides were stained with Wright-Giemsa (ThermoShandon) to depict leukocyte subsets. A total of 200 leukocytes were counted per slide.

Statistical analysis

All data are representative of mean ± SEM of between 3 and 7 replicates. Statistical analysis was performed using a Student’s t test with InStat software. P values of less than 0.05 were considered significant.

Online supplemental material

Fig. S1 shows hematological analysis of leukocyte populations in blood obtained by cardiac puncture of wild-type and flotillin 1−/− mice. Fig. S2 shows that stable overexpression of flotillin 2–GFP increases expression of flotillin 1.


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