BDNF and its pro-peptide are stored in presynaptic dense core vesicles in brain neurons

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Although brain-derived neurotrophic factor (BDNF) regulates numerous and complex biological processes including memory retention, its extremely low levels in the mature central nervous system have greatly complicated attempts to reliably localize it. Using rigorous specificity controls, we found that antibodies reacting either with BDNF or its pro-peptide both stained large dense core vesicles in excitatory presynaptic terminals of the adult mouse hippocampus. Both moieties were ~10-fold more abundant than pro-BDNF. The lack of postsynaptic localization was confirmed in Bassoon mutants, a seizure-prone mouse line exhibiting markedly elevated levels of BDNF. These findings challenge previous conclusions based on work with cultured neurons, which suggested activity-dependent dendritic synthesis and release of BDNF. They instead provide an ultrastructural basis for an anterograde mode of action of BDNF, contrasting with the long-established retrograde model derived from experiments with nerve growth factor in the peripheral nervous system.

Introduction

Polarized cells use well-conserved mechanisms to sort proteins into specific compartments (Mellman and Nelson, 2008), providing them with a directionality that is critically important for both their function and meaningful integration into tissues. Central nervous system (CNS) neurons are prototypically polarized cells with specialized axonal and dendritic compartments that play essential roles in intercellular signaling. Although neurons typically communicate by releasing low–molecular weight neurotransmitters accumulated in synaptic vesicles, they also store and release peptides or small proteins such as brain-derived neurotrophic factor (BDNF), a member of the neurotrophin family. BDNF is known to regulate a wide variety of brain functions in humans, ranging from food intake (Gray et al., 2006) to memory retention (Egan et al., 2003). Indeed, a single amino acid replacement in pro-BDNF has been convincingly shown to correlate with a diminished capacity to remember words and sentences (Egan et al., 2003; Cathomas et al., 2010). Furthermore, in animal models of disease, reduced levels of BDNF have been associated with several conditions, including depression (Calabrese et al., 2007), Rett syndrome (Chang et al., 2006), and Huntington’s disease (Zuccato et al., 2010).

Although many aspects of BDNF biology in the adult brain are thus beginning to be well appreciated, the subcellular localization of this secreted protein in neurons of the adult CNS is still very unclear, in large part because of the very low levels of endogenous BDNF. To address this question, several studies have resorted to neuronal cultures prepared from the embryonic...
rodent hippocampus (Goodman et al., 1996; Hartmann et al., 2001; Kojima et al., 2001; Egan et al., 2003; Adachi et al., 2005; Dean et al., 2009; Matsuda et al., 2009; Jakawich et al., 2010). In most cases, conclusions about the localization of BDNF were inferred from visualization experiments using transfected tagged BDNF constructs, with recent studies concluding that BDNF is transported in and released from both axons and dendrites (Adachi et al., 2005; Dean et al., 2009; Matsuda et al., 2009; Jakawich et al., 2010). As firmly established by in situ hybridization studies, the Bdnf gene is expressed in an activity-dependent fashion by numerous excitatory neurons (Zafra et al., 1990), with protein levels increasing by ~10-fold during the first 3 wk after birth, in parallel with the development of synaptic activity (Tao et al., 1998; Kolbeck et al., 1999). Nonetheless, BDNF remains, even in the adult brain, an extremely rare protein, making its unambiguous detection in vivo a challenging task that is further complicated by the early death of Bdnf-null mutant animals, thus precluding their use as age-matched controls (CONs; Ernfors et al., 1994; Jones et al., 1994). Given that an understanding of the mode of action of secreted proteins depends on a detailed knowledge of their subcellular localization, the present study investigates the distribution of BDNF in the adult hippocampus at both light microscopic and ultrastructural levels using three lines of transgenic animals: (1) a BDNF knockin line allowing the use of antibodies directed against a tagged version of the BDNF gene (Matsumoto et al., 2008), (2) a line conditionally lacking BDNF in adult neurons allowing for the specificity control of BDNF antibodies (Rauskolb et al., 2010), and (3) a mutant line displaying seizure episodes that are accompanied by markedly elevated levels of BDNF (Heyden et al., 2011). Antibodies against the BDNF prodomain were also used as an independent means of localizing BDNF as well as to generate information about the status of its cleaved prodomain (described hereon as the pro-peptide).

Results

Antibodies to BDNF and pro-BDNF reveal similar staining patterns

Hippocampal sections prepared from 8-wk-old animals were incubated either with the monoclonal BDNF antibody Mab#9 (anti-BDNF; Fig. 1 B) or with polyclonal antibodies recognizing the BDNF prodomain (anti–pro-BDNF; Fig. 1 H). Tissues from age-matched mice engineered to delete BDNF from neurons (cbdnf ko; Rauskolb et al., 2010) were used as a negative CON (Fig. 1, C, D, I, and J). In addition, sections from a knockin mouse line expressing Bdnf-Myc (Fig. 1 E; Matsumoto et al., 2008) were incubated with Myc antibodies (anti-Myc), with wild-type (WT) tissue used as a CON (Fig. 1, F and G). These three unrelated antibodies yielded strikingly similar staining patterns (Fig. 1, B, E, and H). Arrows denote the end bulb of the mossy fiber projection, which delineates CA3 and CA1. Note the relative lack of staining in CA1 in WT and Bdnf-Myc sections. Bars: (B, E, and H) 500 µm; (C, F, and I) 100 µm; (D, G, and J) 50 µm.
BDNF-IR is detected in subsets of neurons

Low-power examination of BDNF-IR in the dentate gyrus (DG) revealed a subset of immunopositive granule cells in the suprapyramidal and infrapyramidal blades (Fig. 2 A). BDNF-IR varied in intensity among the labeled cells, with staining concentrated in the somal apex (Fig. 2 A, inset). Anti–pro-BDNF staining was confined to exactly the same subset of granule cells containing BDNF-IR (Fig. 2, B and C), with somal pro-BDNF–IR also concentrated at the apex (Fig. 2 B, inset). In addition, the hilar region, which contains mossy fiber collateral axons of the granule cells, was intensely stained (Fig. 2, A–C). In the CA3 region, a subset of pyramidal neurons also showed both BDNF-IR (Fig. 2, D and F) and pro-BDNF–IR (Fig. 2, E and F). High-resolution examination of sections labeled with anti-Myc and anti–pro-BDNF, along with antibodies against the Golgi matrix protein GM130, revealed Myc-IR throughout the soma and the initial dendritic segments (Fig. 2 G), whereas pro-BDNF–IR showed a similar, albeit more punctate, distribution (Fig. 2 H). Comparison with GM130-IR showed pro-BDNF–positive puncta closely associated with the Golgi apparatus (Fig. 2, I and J). In line with this, anti–pro-BDNF immunogold labeling localized the protein to the Golgi complex of CA3 somata (Fig. 2 K). In CA1, BDNF and pro-BDNF containing was also detected in a small number of pyramidal neurons in temporal hippocampus sections (unpublished data).

BDNF-IR and pro-BDNF-IR are both detected in presynaptic terminals

The granule cells give rise to mossy fiber axons, whose targets include the complex spines on proximal dendrites of CA3 neurons. Mossy fibers project through and terminate in stratum lucidum (SL) and are characterized by prominent specialized endings known as mossy fiber boutons (MFBs). Accordingly, strong BDNF-IR and pro-BDNF–IR were observed within SL (Figs. 1 [B, E, and H] and 2 [D–F]). Using high-resolution confocal microscopy, both BDNF-IR and pro-BDNF–IR were found to be colocalized to the same subset of MFBs (Fig. 3, A–C).

Additional markers were then applied to identify the type of vesicles containing BDNF-IR and to compare the distribution of BDNF with other peptides known to be anterogradely transported by granule cells. As expected, BDNF-IR did not colocalize with the synaptic vesicle markers synaptophysin (SYP; Fig. 3, D and E) or VGLUT-1 (Fig. 3, F and G). We then tested possible colocalization with Met-enkephalin (Met-enk), an opioid peptide also derived from a larger precursor protein and stored in dense core vesicles (DCVs; Cheng et al., 1995). In a small proportion of granule cells and their axons, Met-enk–IR was detected throughout the soma and initial dendritic segment (Fig. 3 I). Although Met-enk–positive granule cells invariably coexpressed BDNF-IR (Fig. 3 H), the immunoreactive signals of these two precursor-derived molecules remained separate, suggesting that they do not reside together in the same secretory vesicles (Fig. 3, J and K–M). Similar conclusions were reached with cholecystokinin (CCK), a neuropeptide transported along the mossy fiber projection pathway of the ventral mouse hippocampus (Gall et al., 1986). Double labeling with anti-CCK and -BDNF revealed complete segregation of the two peptides within MFBs (Fig. 3, N–P).
Synaptic vesicles, numerous synaptic contacts with CA3 complex spines, nonsynaptic puncta adherentia at dendritic shafts, and a relatively large surface area. A subset of MFB profiles labeled with anti-BDNF (Fig. 4 A), anti-Myc (not depicted), or anti–pro-BDNF (Fig. 4 B) contained distinct aggregates of gold grains; at 10,000-fold magnification, these gold clusters were found to be associated with large vesicles encompassed by an electron-dense membrane. Although the vesicles were sometimes masked by gold grains, fortuitous grain distribution occasionally revealed an electron-dense core (Fig. 4, A and B, insets). Within MFB profiles of cbdnf ko sections, gold grains were never specifically associated with any type of organelle (Fig. 4, C and D). To assess the density of immunogold particles in MFB profiles, gold clusters and single gold grains were quantified in anti-BDNF and anti–pro-BDNF–labeled tissues.
and compared between sections from pooled WT/Bdnf-Myc mice and cbdnf ko mice. Anti-BDNF–labeled MFB profiles showed a mean density of 2.67 ± 0.35 clusters/µm² in WT/Bdnf-Myc animals compared with 0.38 ± 0.12 clusters/µm² (P < 0.005) in cbdnf ko profiles (Fig. 4 E). Moreover, single gold grain densities were significantly reduced in cbdnf ko MFB profiles, with 5.63 ± 0.94 grains/µm² compared with 21.99 ± 2.57 grains/µm² in WT/Bdnf-Myc (P < 0.005; Fig. 4 F). Similarly, pro-BDNF immunogold labeling showed a mean density of 2.01 ± 0.43 clusters/µm² and 27.44 ± 8.21 single grains/µm² in MFB profiles from WT/Bdnf-Myc animals, whereas in comparison, MFB profiles from cbdnf ko animals showed significant reductions in cluster (0.14 ± 0.10/µm²; P < 0.05; Fig. 4 E) and single grain (5.82 ± 2.06/µm²; P < 0.05; Fig. 4 F) densities, respectively.

Although BDNF immunogold labeling was mostly concentrated in presynaptic terminals, labeled vesicles were also occasionally observed within unmyelinated axon segments in SL (Fig. 5 A), which in fortuitous sections could be seen to give rise to giant MFBs (Fig. 5 B).

Ultrathin sections of SR (CA1) labeled with anti-BDNF and anti–pro-BDNF immunogold were also examined, and, as expected, large cluster-labeled secretory vesicles were observed within small axon terminals (Fig. 5 C). Depending on the proximal distal level of SR, these infrequent labeled boutons likely correspond to Schaffer collateral terminals or entorhinal terminals. No such labeled terminals were observed in cbdnf ko sections (Fig. 5 D).

**BDNF-IR and pro-BDNF-IR are not detected in dendrites**

Next, we examined the possible localization of BDNF in dendrites by analyzing sections double labeled with anti-Myc and anti–microtubule-associated protein–2 (MAP-2) (MAP-2). Both in BDNF-positive granule cells and CA3 neurons, Myc-IR only extended as far as the initial dendritic segments, and there was no evidence of colocalization in SL (Fig. 6, A–C). Sections were alternatively labeled with antibodies to Arc/Arg3.1, an immediate early gene product up-regulated in somata and dendrites during elevated synaptic activity (Lyford et al., 1995). Confocal scanning revealed that the majority of granule cells labeled with BDNF-IR and pro-BDNF-IR also expressed Arc/Arg3.1-IR (Fig. 6, D–G). Although Arc/Arg3.1-IR was seen throughout the soma and dendritic arbor, strong coexpression of BDNF-IR and pro-BDNF–IR was confined to the cell soma (Fig. 6, D–G).

In ultrathin sections prelabeled either with anti-BDNF or anti–pro-BDNF immunogold, gold grains were sparsely distributed within dendritic profiles (Fig. 6, H and K). When profiles were thoroughly scrutinized for specifically labeled vesicles or endosomes, none was found to be stained above the background levels observed in dendrites from cbdnf ko sections (Fig. 6, I and L). Gold grain quantification of anti-BDNF–labeled sections (Fig. 6 J) revealed a mean density of 3.56 ± 0.45 grains/µm² in WT/Bdnf-Myc mice versus 4.72 ± 0.91 grains/µm² in cbdnf ko mice (P = 0.81). Similarly, in anti–pro-BDNF–stained sections, mean densities were comparable between WT/Bdnf-Myc mice (4.69 ± 1.09 grains/µm²) and cbdnf ko mice (4.08 ± 2.02 grains/µm²; P = 0.31; Fig. 6 M).

**Localization of BDNF-IR and pro-BDNF-IR in Bsn mutants**

Next, we examined the hippocampus of mice lacking the presynaptic protein Bassoon, as these mutants develop episodic generalized seizures (Altrock et al., 2003) and have enlarged cortices and hippocampi (Angenstein et al., 2007). Concurrent with the development of seizures, BDNF protein levels become significantly higher than those measured in adult CON littermates (see Fig. 10 C; Heyden et al., 2011). Whereas Bassoon (Bsn) mutants showed the typical distribution pattern of BDNF-IR and pro-BDNF–IR, a dramatic increase in staining intensity largely confined to the neuropil was observed (compare CON in Fig. 7 [A–D] with Bsn in Fig. 7 [E–H]). In contrast, granule cell
bodies from Bsn mutants did not show increases in anti-BDNF (Fig. 7 I) or anti–pro-BDNF (Fig. 7 J) staining intensities, although a much higher proportion of cells was labeled in comparison with CON tissues. Granule cell dendrites in the molecular layer remained unlabeled in Bsn mutants, whereas in the CA3 region, the stark increase in BDNF-IR (Fig. 7 G) and pro-BDNF–IR (Fig. 7 H) was confined to SL. Closer examination revealed intense presynaptic labeling in MFB profiles (Fig. 7, K and L), which was confirmed by a lack of colocalization with the postsynaptic markers synpo (Fig. 7 K) and MAP-2 (Fig. 7 L), respectively.

Enhanced BDNF staining in Bsn mutants was not only confined to the granule cell–CA3 projection pathway. In the CA1 region, where BDNF-IR can usually only be detected at high magnification (compare Fig. 1 with Figs. 3 Q and 5 D), a conspicuous band of punctate BDNF-IR and pro-BDNF–IR was observed at the border of SR and stratum lacunosum–molecular (SL–M; Fig. S1, A–C), corresponding to a region known to harbor fibers from the entorhinal cortex (Amaral and Lavanex, 2006). Importantly, these increased signals colocalized (Fig. S1 D), showing no overlap with either synpo-IR (Fig. S1 E) or glial fibrillary acidic protein (GFAP)–IR (Fig. S1 F), which label dendritic spines and reactive astrocytes, respectively. This suggests that entorhinal neurons also represent a possible presynaptic source of BDNF for CA1 neurons.

Immunogold-labeled sections from both WT and Bsn mutants were then examined at 2,000-fold magnification. In comparison with CON animals (Fig. 8, A and B), a higher number of labeled MFB profiles containing more BDNF-positive DCVs was observed in sections from Bsn mutants (Fig. 8, C and D), with clusters accumulated at the synaptic membrane. This was confirmed by quantification, with anti-BDNF–labeled MFBs containing a significantly higher density of gold clusters (1.7 ± 0.3 clusters/µm²; Fig. 8 E) and grains (14.4 ± 2.5 grains/µm²; Fig. 8 F) compared with CONs (0.8 ± 0.01 clusters/µm² with P < 0.005 and 6.9 ± 0.5 grains/µm² with P < 0.05, respectively); these relatively low increases in mean cluster densities are a result of the larger areas of MFB profiles in Bsn mutants.

Gold grain distribution and density in dendrites were also compared between Bsn mutant (Fig. 9, C and D) and CON tissues (Fig. 9, A and B). Dendritic profiles were again scrutinized for evidence of labeled vesicles, but none was found in tissues from either group. Density measurements revealed background values in Bsn mutant mice similar to those observed in CON mice, both in tissues labeled with anti-BDNF (5.4 ± 0.4 grains/µm² for Bsn mutant vs. 5.7 ± 0.5 grains/µm² for CON; P = 0.66; Fig. 9 E, left) and anti–pro-BDNF (5.1 ± 0.1 grains/µm² for Bsn mutant vs. 6.3 ± 0.3 grains/µm² for CON; P = 0.12; Fig. 9 E, right). Increased labeling density was not detected in the extra-cellular space or in nonneuronal cell types such as astrocytes.

Verification of background immunogold labeling
To determine whether background gold labeling is evenly distributed over different subcellular compartments, we extended the quantitative gold grain analysis to dendritic spine profiles and myelinated axon profiles in SL, as they are devoid of clustered-labeled organelles. Quantification of single gold grains overlying these profiles revealed that the density of background gold labeling depends on the type of subcellular compartment (Table 1). The mean values for gold grain densities in spine and myelinated axon profiles did not differ between WT/Myc versus cbcdnf ko or WT versus Bsn mutant tissues. Therefore, spines and myelinated axons from WT mice do not exhibit anti-BDNF and anti–pro-BDNF immunogold labeling above the background levels observed in cbcdnf ko tissues, nor do they contain specifically labeled organelles.

Biochemical detection and quantification of the BDNF pro-peptide
The identity of the molecules recognized by the BDNF antibodies in the immunocytochemistry experiments was then determined.
and incubation conditions used, no measurable proteolysis of recombinant pro-BDNF (500 pg) added to the lysates at the beginning of the extraction procedure could be detected (Fig. S2). The recovery of added recombinant pro-BDNF was 102.2 ± 5.8% (n = 4).

Discussion

Our study reveals that in the adult hippocampus, BDNF and its cleaved pro-peptide are stored in large DCVs located in the presynaptic terminals of excitatory neurons. Both in WT and Bsn mutants, BDNF and its pro-peptide are stored at roughly equimolar ratios and are ~10-fold more abundant than pro-BDNF in hippocampal lysates. Together with the lack of any detectable BDNF staining in dendrites or spines, our results provide direct support for an anterograde mode of action for BDNF in the intact CNS. They also offer a morphological substrate for recent findings indicating that the release of endogenous BDNF accounts for some of the rapid calcium transients observed at synaptic sites on dendrites of CA3 neurons (Lang et al., 2007).
Immunohistochemical localization of BDNF in the CNS

The specificity of BDNF immunostaining in the adult brain is very difficult to ascertain, as Bdnf knockout animals die before reaching maturity, a fact that complicates the interpretation of previous immunohistochemical studies (Wetmore et al., 1991; Dugich-Djordjevic et al., 1995; Schmidt-Kastner et al., 1996; Altar et al., 1997; Conner et al., 1997; Yan et al., 1997; Luo et al., 2001; Danzer and McNamara, 2004; Tongiorgi et al., 2004; Zhou et al., 2004; Agassandian et al., 2006; Avvenagha et al., 2006, Salio et al., 2007). Our study deals with the mature hippocampus, as much of the current interest in BDNF relates to its role in the adult brain, and the staining specificity of our BDNF monoclonal antibody was verified with tissues from age-matched mice lacking BDNF in the CNS (Kolbeck et al., 1999; Rauskolb et al., 2010). The choice of BDNF antibody was also important, as none of the commercially available BDNF antibodies we tested yielded specific staining (unpublished data). In addition, a mouse line was used in which Bdnf was replaced with a Myc-tagged version of the gene (Matsumoto et al., 2008), allowing additional control experiments to be performed with anti–human Myc antibodies on WT tissue sections. We note that a recent light microscopy study that also used a knockin strategy to tag endogenous BDNF came to a conclusion very similar to our own, revealing identical labeling of the mossy fiber pathway with antibodies directed against either the HA epitope used to tag BDNF or the BDNF prodomain (Yang et al., 2009). However, our biochemical results (see later part of Discussion) now indicate that pro-BDNF antibodies primarily detect the BDNF pro-peptide, which is present at much higher levels than uncleaved pro-BDNF. The pro-BDNF antibodies further validate our localization data, as they recognize an epitope unrelated to those detected by the antibodies to BDNF or Myc but similarly label presynaptic large DCVs. These organelles are well known to be involved in the regulated secretion of many other neuronal signaling peptides including Met-enk (Cheng et al., 1995), which was also used here as a presynaptic marker. DCVs are typically found in presynaptic terminals but rarely in postsynaptic structures such as dendrites or spines, with the notable exception of the DCVs containing the neuromodulators vasopressin and oxytocin, found in the dendrites of specialized neurosecretory cells (Kennedy and Ehlers, 2011).

Using cultured cells to study the cell biology of BDNF

Numerous previous studies have used cultured hippocampal neurons to study the storage and release of BDNF (Haubensak et al., 1998; Hartmann et al., 2001; Wu et al., 2004; Adachi et al., 2005; Brigadiiski et al., 2005; An et al., 2008; Dean et al., 2009; Matsuda et al., 2009). They all come to the conclusion that BDNF is stored in and released from both axonal and dendritic compartments, raising the question of what could underlie the apparent discrepancy with our findings. Although the majority of these studies utilize neurons transfected with a cDNA encoding a fluorescently tagged form of BDNF, some of the conclusions are also based on the detection of endogenous BDNF in vitro. However, cultured embryonic neurons grow in a very different environment compared with neurons in vivo, with fewer and less intimate cell–cell interactions between neighboring neurons and astrocytes, an incomplete ECM, and a lack of laminar input. In vivo, the development of neuronal circuits is spatially tightly regulated (Frotscher et al., 2000), whereas in vitro, synapses seem to form randomly. As a result of these cell-extrinsic differences, it is conceivable that cultured neurons fail to sort cargo-loaded vesicles as strictly as they do in vivo (Mellman and Nelson, 2008). In addition, we note that cultured neurons express significantly higher levels of BDNF than are detectable even in adult hippocampal extracts (unpublished data).
BDNF signaling have inferred the translation of BDNF mRNA in dendrites (Tanaka et al., 2008), it has not been possible thus far to assess directly whether dendritic BDNF mRNAs are translated locally. Thus, it is of particular significance that our experiments revealed a complete lack of specific BDNF protein signals in dendrites not only in WT but also in Bsn mutant animals. Although our experiments cannot rule out that functionally relevant quantities of BDNF may be released from dendrites, they indicate that, at present, there is no structural basis for this speculation.

With regard to the biochemistry of BDNF, initial studies suggested that the most abundant, or indeed the only detectable, translation product in adult brain extracts was uncleaved pro-BDNF (Chen et al., 2005; An et al., 2008). However, a signal of similar size and intensity to pro-BDNF can readily be detected in mice lacking BDNF in the CNS (Matsumoto et al., 2008). Furthermore, pulse-chase experiments showed that pro-BDNF is a short-lived intermediate that is rapidly processed intracellularly (Matsumoto et al., 2008). BDNF, like all other neurotrophins, is initially translated as a glycosylated precursor protein, presumably allowing the proper folding and disulfide bridging of the mature protein (Leibrock et al., 1989; Rattenholl et al., 2001). Accordingly, we detected specific pro-BDNF–IR in the

BDNF biosynthesis and processing

Although in situ hybridization experiments have long established that BDNF mRNA can be unambiguously detected in the cell bodies of excitatory neurons (Hofer et al., 1990; Isackson et al., 1991), some studies have also suggested its presence in dendrites (Tongiorgi et al., 1997, 2004; An et al., 2008) and its active transport upon stimulation (Tongiorgi et al., 1997, 2004; Chiaruttini et al., 2009; Baj et al., 2011; Louhivuori et al., 2011; Wu et al., 2011). However, these experiments revealed that BDNF mRNA is predominantly detected in the most proximal part of the dendrite, whereas other actively transported mRNAs can be detected in dendrites hundreds of micrometers away from the soma, as is the case for the Arc/Arg3.1 (Lyford et al., 1995), CaMKII-α (Burgin et al., 1990; Miller et al., 2002), or dendrin (Link et al., 1995) mRNAs. This leaves the possibility that BDNF mRNA localizes to proximal dendrites by diffusion rather than active transport. Selective trafficking has also been difficult to assess, as activity-dependent transcription augments the intensity of the proximal mRNA signal, allowing more distal diffusion of the in situ hybridization enzymatic reaction product (Tongiorgi et al., 1997, 2004). Although elegant experiments aimed at blocking protein synthesis and interfering with
Golgi apparatus of dentate granule cells and CA3 neurons but not in other organelles such as lysosomes or endosomes. Although the intracellular site of pro-BDNF cleavage and the participating proteases are yet to be identified, the existence of the BDNF pro-peptide in brain lysates has not been previously reported (see Fig. 10). This pro-peptide has hitherto escaped detection presumably because it is readily washed away from blotting membranes. This loss could be prevented by fixing the membranes with glutaraldehyde after electrophoretic transfer (Karey and Sirbasku, 1989). Although we observed that prolonged stimulation of cultured neurons results in the cosecretion of BDNF and its pro-peptide (unpublished data), the biological significance of this corelease is unclear.

**BDNF detection in the hippocampus of Bsn mutants**

Mutant mice expressing a truncated version of the large presynaptic protein Bassoon develop spontaneous epileptic seizures for reasons that are only partially understood (Altrock et al., 2003). In line with the well-established link between excitatory neuronal activity and Bdnf transcription, BDNF protein levels were found to increase in parallel with, but not precede, the development of seizure episodes (Heyden et al., 2011). It seems unlikely that Bassoon is directly involved in the increase in BDNF levels, as a mutation in the potassium channel Kv1.1 that also leads to seizure activity is accompanied by a marked increase in BDNF mRNA and megencephaly (Diez et al., 2003; Lavebratt et al., 2006), as is the case for the Bsn mutation (Angenstein et al., 2007). Blocking activity with carbamazepine in the Kv1.1 mutant prevents the development of megencephaly and restores BDNF mRNA to normal levels (Lavebratt et al., 2006). Despite the marked increase in BDNF and its pro-peptide in the Bsn mutant, both immunoreactive signals remained confined to presynaptic terminals, with no detectable staining in either granule or pyramidal cell dendrites or glial cells (Bergami et al., 2008). In comparison with WT animals, the only remarkable change observed in the Bsn mutant was an increase in the density of BDNF-positive DCVs in MFBs. These observations are in line with earlier studies in the rat, in which chemically induced seizure activity led to an increase in presynaptic BDNF staining (Vezzani et al., 1999; Scharfman et al., 2002; Danzer and McNamara, 2004). Despite the substantially higher BDNF levels, we did not detect any BDNF staining in the extracellular space, suggesting that after its release, BDNF becomes rapidly diluted by diffusion as well as by cellular reuptake, thus preventing accumulation to a degree that would allow immunohistochemical detection.

**BDNF and hippocampal mossy fibers**

The hippocampal mossy fiber projection stands out as the most heavily stained structure in the adult CNS using either BDNF or pro-BDNF antibodies (Fig. 1). This pathway is thought to exhibit presynaptic long-term potentiation (LTP; Nicoll and Schmitz, 2005) and to be important for pattern completion as well as for the storage and recall of information.
brains were obtained from the following lines of mice, all of which were used at peripheral nervous system and the CNS. In addition, 6-burst–induced release of endogenous BDNF from mossy fibers is sufficient to induce intracellular changes in postsynaptic CA3 neurons (Lang et al., 2007). Furthermore, several previous studies in CA1 have pointed to the need for BDNF in order for LTP to reach maximal values (Korte et al., 1995; Patterson et al., 1996), and our results uncover a presynaptic location of BDNF in this hippocampal area (Figs. 5 and S1).

Materials and methods

Mouse lines

Brains were obtained from the following lines of mice, all of which were used at ~8 wk of age: WT (C57Bl/6), Bdnf-Myc (Matsumoto et al., 2008), cbdnf ko (Rauskolb et al., 2010), and Bsn mutant (BsnEX4/5; Altrock et al., 2003). All experiments were performed in accordance with the institutional guidelines of the University of Freiburg and the University of Basel.

Preparation of tissue for immunostaining

Mice (n = 7 for WT, n = 7 for Bdnf-Myc, n = 4 for cbdnf ko, and n = 7 for Bsn mutant) were transcardially perfused with 0.9% NaCl followed by a solution of 3% M.O.M. (Mouse-on-Mouse blocking serum; Vector Laboratories) in TBS for 1 h. Primary antibodies were diluted in a solution of 3% BSA, 2% normal donkey serum, and 0.2% Triton X-100 in TBS to yield the following final concentrations/dilutions: 10 µg/ml anti-BDNF, 0.4 µg/ml anti-Myc, 1.33 µg/ml anti–pro-BDNF, 3.3 µg/ml anti–VGLUT-1, 4 µg/ml anti–anti-SYP, 0.9 µg/ml anti–GM130, 2 µg/ml anti–Arc/Arg3.1, 4 µg/ml anti–MAP-2, 1,200 anti-synpo, 3 µg/ml anti–M-Met-enk, 1,200 anti–CCK, and 1:1,000 anti-GFAP. Sections were incubated for at least one night at 4°C in either one or more of these primary antibodies. For fluorescent signal detection, the following Alexa Fluor-conjugated secondary antibodies were used: donkey anti–rabbit-488 or -555, donkey anti–goat-555 or -488, donkey anti–mouse-555 or -647, and donkey antiguinea pig–Cy5 (all purchased from Millipore). Labeled sections were mounted onto glass slides and coverslipped with fluorescent mounting medium (Dako).

Confocal microscopy

Slide-mounted sections of immunolabeled hippocampi were viewed on an inverted microscope (Axiovert 200; Carl Zeiss) equipped with Plan Apochromat 20× (0.75 NA) or 63× (all differential interference contrast; 1.4 NA) objectives and attached to a spectral confocal laser system (LSM510; Carl Zeiss) powered by LSM 4.2 Meta software (Carl Zeiss). The tissue was scanned at room temperature with 488-, 543-, and 633-nm laser lines to detect the Alexa fluorophores 488, 555, and 647, respectively, and high-resolution images of optical sections (z slices) were captured using sequential line [mean of four] scanning. Colocalization of two or three fluorophores was quantitatively assessed in the x, y, and z planes of each optical section. Maximal projection images of confocal z series (stacks) were generated where indicated in the figure legends. Minimal adjustments to image contrast and intensity were made in Photoshop CS (version 8.0.1; Adobe) using the levels or contrast/brightness functions. Images were arranged and annotated using CorelDRAW 12 software (Corel Corporation).

Immunogold EM

Immunogold EM prelabeling. 50-µm vibratome sections of perfusion-fixed hippocampi from WT, Bdnf-Myc, cbdnf ko, and Bsn mutant mice were thoroughly washed in 50 mM TBS, pH 7.4, and immersed for 2 h in a cryoprotecting solution (25% sucrose and 10% glycerol in 0.05 M PB) before being snap frozen in liquid nitrogen–cooled isopentane. After immediate thawing and washing in 0.1 M PB, sections were incubated for 1 h in blocking solution (0.3% M.O.M. in TBS for anti-BDNF staining and 20% normal goat serum in TBS for anti-pro-BDNF and anti-Myc staining).
followed by one to three nights at 4°C in a 3% BSA/TBS solution contain-
ing either 20 µg/ml anti-BDNF, 1.33 µg/ml anti-pro-BDNF (AN-03), or 0.4 µg/ml anti-Myc antibody. After thorough washing in TBS, the sections were incubated overnight at 4°C in 3% BSA/TBS solution containing either 1.4 nm of gold-conjugated goat anti-mouse, goat anti-rabbit, or rabbit anti-goat IgG. Sections were rinsed in TBS and then fixed for 10 min in a 1% glutaraldehyde solution. Tissue-bound gold particles were washed using a silver filtration kit (HQ Silver; Nanoprobes). The sections then underwent osmification for 40 min in a solution of 0.5% OsO₄ and 6.86% sucrose in 0.1 M PB.

Embedding. After osmification, sections were washed in 0.1 M PB followed by 50% ethanol (EtOH). The tissue was then incubated in 1% uranyl acetate in 70% EtOH for 35 min followed by 10-min dehydration steps in increasing grades of EtOH. After washing in propylene oxide, the tissue was embedded in Durcupan (Fluka). Ultrathin sections (60 nm) of selected hippocampal areas (DG, CA3, and CA1) were cut and mounted on formvar-coated nickel grids. Sections were viewed and examined in an electron microscope (LEO 906E; Carl Zeiss).

Quantification of immunogold staining
Gold-labeled sections from WT, Bdnf-Myc, cbdnf ko, and Bsn mutant ani-
mals were first examined in the electron microscope at magnifications ranging from 6,000 to 12,000 x, and areas containing labeled structures in SL of CA3 were then photographed using a dual-speed charge-coupled device camera (TRS SharpEye; Troeneldle) and analySIS acquisition soft-
ware (Soft Imaging System; Olympus). MFBs were unambiguously identi-
fied based on their well-defined morphology (see criteria in Results). An
organelle in an MFB was deemed labeled if associated with at least four gold grains (described in this study as a cluster). MFBs containing labeled organelles were then selected for quantitative analysis, in which the number of single gold grains per dendritic profile was counted and expressed as a density measurement (i.e., the number of grains per µm² of MFB). Although the rare gold clusters observed in cbdnf ko sections showed no association with any particular organelle, they were still in-
cluded in the counts. In addition to cluster quantification, the respective
number of grains per µm² was determined against known amounts of purified BDNF pro-peptide produced in Escherichia coli (Koshimizu et al., 2009). The COS-7-derived cleavage-resistant pro-BDNF was used as a molecular mass marker, whereas WT pro-BDNF was used in recovery experiments with tissue lysates (Fig. 52).

Statistical analysis
All values are expressed as mean ± SEM. For BDNF and pro-BDNF immuno-
gold quantification, 10–20 MFBs and 30–40 dendrites per WT, Bdnf-Myc, cbdnf ko, and Bsn mutant mice were analyzed, and mean values for each animal were calculated. Group means (pooled WT/BDNF-Myc vs. cbdnf ko and WT vs. Bsn mutant) were compared using a Student’s t-test. A probability of P < 0.05 was considered to be statistically significant.

Online supplemental material
Fig. S1 shows an increase in BDNF-IR and pro–BDNF-IR in area CA1 of the Bassoon mutant hippocampus. Fig. S2 shows the complete recovery of WT recombinant pro-BDNF from hippocampal lysates. Online supple-
mental material is available at http://www.jcb.org/cgi/content/full/jcb.20121038/DC1.

We thank Dr. J. Kowalski and M. Sibbe for helpful statistical advice.
This work was supported by the Swiss National Foundation (31003A_124902/1), the German Research Foundation (SFB 780, project A4), and a Postdoctoral Research Fellowship [to S. Dien] from the Alexander von Humboldt Foundation. M. Frotscher is a Senior Research Professor of the Hertie Foundation.

Submitted: 9 January 2012
Accepted: 15 February 2012

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Figure S1. **BDNF labeling is increased in the CA1 region of Bsn mutants.** (A–C) BDNF-IR and pro–BDNF-IR in Bsn mutants demonstrates a laminar-specific increase in staining at the border of SR and SL-M (arrows). The dashed lines mark the hippocampal fissure [HF]. GCL, granule cell layer; H, hilus; ML, molecular layer; PCL, pyramidal cell layer; SO, stratum oriens. (D) High-resolution confocal image (represented by box d in C) reveals colocalization of BDNF-IR and pro–BDNF-IR at the border of SR and SL-M from a Bsn mutant. (E) Colabeling with antibodies against BDNF and synpo reveals segregation of anti-BDNF (presynaptic) and anti-synpo (postsynaptic) signals at the border of SL and SL-M. (F) Colabeling with antibodies against BDNF and GFAP shows a lack of BDNF-IR in astrocytes. Bars: (A–C) 200 µm; [D] 30 µm; [E and F] 10 µm.

Figure S2. **Recovery of recombinant WT pro-BDNF and mBDNF from hippocampal lysates.** Hippocampal lysates were immunoprecipitated with anti-BDNF for 3 d in the presence or absence of protease inhibitors. IP was performed with or without known amounts of recombinant WT BDNF containing pro-BDNF (pro) and mature BDNF (m). After IP, WB was performed with BDNF-N20 antibodies. Recombinant (rec.) mature BDNF was used as a reference. HIP, hippocampus; IB, immunoblot.