Dishevelled stabilization by the ciliopathy protein Rpgrip1l is essential for planar cell polarity

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Cilia are at the core of planar polarity cellular events in many systems. However, the molecular mechanisms by which they influence the polarization process are unclear. Here, we identify the function of the ciliopathy protein Rpgrip1l in planar polarity. In the mouse cochlea and in the zebrafish floor plate, Rpgrip1l was required for positioning the basal body along the planar polarity axis. Rpgrip1l was also essential for stabilizing dishevelled at the cilium base in the zebrafish floor plate and in mammalian renal cells.

In rescue experiments, we showed that in the zebrafish floor plate the function of Rpgrip1l in planar polarity was mediated by dishevelled stabilization. In cultured cells, Rpgrip1l participated in a complex with inversin and nephrocystin-4, two ciliopathy proteins known to target dishevelled to the proteasome, and, in this complex, Rpgrip1l prevented dishevelled degradation. We thus uncover a ciliopathy protein complex that finely tunes dishevelled levels, thereby modulating planar cell polarity processes.

Introduction

Many epithelia are characterized by a coordinated polarization of cells along the plane of the tissue, called planar cell polarity (PCP). This property has been extensively studied in Drosophila, where many actors involved in this process have been identified. Central to this process is the “core” PCP pathway, which involves the transmembrane proteins Frizzled (Fz), Strabismus/ Van Gogh (Stbm/Vang), and Flamingo (Fmi), as well as the cytosolic proteins Dishevelled (Dsh), Diego (Dgo), and Prickle (Pk). These proteins are enriched at adherens junctions and form two asymmetric complexes, localized at opposite positions along the planar polarity axis of the cell (Goodrich and Strutt, 2011). In vertebrates, one of the best model systems to study PCP is found in the mammalian cochlea, where all the mechanosensory cells in the organ of Corti possess a V- or W-shaped, actin-based stereociliary bundle with stereotyped orientation along the plane of the epithelium (Jones and Chen, 2008). Other examples of PCP signaling-dependent mechanisms in vertebrates include polarized cell intercalation and directed cell migration leading to convergence-extension (CE) movements (Karner et al., 2009; Roszko et al., 2009), and oriented cell division (Fischer and Pontoglio, 2009; Ségalen et al., 2010), which occur during gastrulation, neurulation, and renal tubule elongation. Proteins involved in vertebrate PCP, including the Stbm/Vang orthologue Van Gogh-like (Vangl2), Frizzled (Fz3/6), Prickle (Pki1/2), and Dishevelled (Dvl1-3), are distributed asymmetrically in the mouse cochlear and vestibular hair cells (Wang et al., 2005; Montcouquiol et al., 2006; Deans et al., 2007; Narimatsu et al., 2009).

Cilia are microtubule-based organelles projecting out from the cell surface of most eukaryotic cells and performing motile and/or sensory functions. In vertebrates, primary (nonmotile) cilia are present in virtually all cells and are involved in the Hedgehog (Hh), PDGFa, Notch, and Wnt pathways (Goetz and Deans, 2007; Montcouquiol et al., 2006). Additional factors, such as the ciliopathy proteins inversin (Invs; also known as nephrocystin-4), cilia associated protein (Carp) and Rpgrip1l, have been shown to stabilize Dishevelled and to control its function in planar polarity processes as described at http://creativecommons.org/licenses/by-nc-sa/3.0/).
Inversin negatively regulates Wnt–dishevelled for proteasomal degradation (Simons et al., 2005). In Zebrafish embryos depleted in Bbs proteins show CE defects enhanced by reduction in core PCP protein amounts (Ross et al., 2005; Gerdes et al., 2007; May-Simera et al., 2009). Moreover, mouse Ift88 and Bbs mutants show planar polarity defects in the cochlea (Ross et al., 2005; Jones et al., 2008). In several instances, the cilium itself acquires a polarized position within the cell and/or a polarized orientation of its basal body, and thereby constitutes a readout of planar polarity (Wallungford, 2010). This is the case for the multicilia of the Xenopus larval skin (Park et al., 2008; Mitchell et al., 2009) and of mammalian ependymal cells (Guirao et al., 2010), as well as for monocilia in the mouse node (Song et al., 2010), in the cochlea (Jones and Chen, 2008), and in the zebrafish floor plate (Borovina et al., 2010). This planar polarization of the cilium requires the core PCP genes Vangl2/2, Fz, and Dvl (Montcouquiol et al., 2003; Park et al., 2008; Borovina et al., 2010; Hashimoto et al., 2010; Song et al., 2010).

The level at which cilia are involved in planar polarity, and their link with PCP signaling, are unclear. In the mouse cochlea, polarity defects are observed in Ift88 mouse mutants, which show total cilium loss, without disruption of the polarized distribution of the core proteins Vangl2 and Fz3 (Jones et al., 2008). This suggests that cilia act downstream or parallel to the PCP signaling pathway. Other data suggest that cilia mediate a switch between the Wnt–β-catenin and the Wnt-PCP pathways. The basolateral domain of the Nphp2/inversin (inv) gene, targets cytoplasmic dishevelled for proteasomal degradation (Simons et al., 2005). Inversin negatively regulates Wnt–β-catenin signaling in cultured mammalian cells and in Xenopus and zebrafish embryos; conversely, it interacts functionally with the PCP pathway during CE (Simons et al., 2005). Ciliary proteins also negatively control β-catenin stability (Gerdes et al., 2007; Corbit et al., 2008) and nuclear translocation (Lancaster et al., 2011). Several Wnt pathway proteins such as Vangl2, Diversin, β-catenin, and Jouberin, are enriched at the basal body and/or cilium (Ross et al., 2005; Itoh et al., 2009; Guirao et al., 2010; Lancaster et al., 2011), underlining the role of this structure as a regulatory platform for Wnt signaling. Despite these numerous studies, the function of cilia in the Wnt pathways remains poorly understood and controversial (Huang and Schier, 2009; Wallingford and Mitchell, 2011).

To better understand the role of different ciliary protein complexes in vertebrate PCP, we investigated the function of the ciliopathy gene Rpgrip1l (also called Mks5, Nphp8, and, for the mouse gene, Ftm) in this process. Rpgrip1l is a 175-kD protein containing several protein–protein interaction domains: three N-terminal coiled-coil domains, two C2 domains, and a C-terminal RPGR-interacting domain (RID; Vierkotten et al., 2007). The protein is mainly found at the ciliary transition zone where it forms a complex with nephrocystin-1 and nephrocystin-4 (Nphp4 gene; Roepman et al., 2005; Delous et al., 2007; Sang et al., 2011). In Caenorhabditis elegans sensory neurons, Rpgrip1l interacts functionally with NPHP and MKS proteins for the formation and function of the ciliary gate (Williams et al., 2011). The human RPRGIP1L gene is one of the causal genes in Meckel and Joubert type B syndromes, two autosomal-recessive multisystem ciliopathies characterized by polydactyly, kidney cysts, and central nervous system malformations such as cerebellar vermis hypoplasia and encephalocoele (Arts et al., 2007; Delous et al., 2007), and acts as a modifier gene in most ciliopathies (Khanna et al., 2009; Zaghloul and Katsanis, 2010). In mouse, Rpgrip1l is required for normal ciliogenesis in some but not all cell types (Arts et al., 2007; Delous et al., 2007; Vierkotten et al., 2007; Besse et al., 2011). Mice with a targeted inactivation of the Ftm/Rpgrip1l gene die around birth and recapitulate most malformations observed in MKS fetuses, including exencephaly, olfactory bulb agenesis, polydactyly, and kidney cysts (Delous et al., 2007; Vierkotten et al., 2007; Besse et al., 2011). Rpgrip1l is required for the correct regulation of Hh transduction (Vierkotten et al., 2007), and our recent data demonstrate that its function in telencephalic morphogenesis is mediated by the regulation of Gli3 proteolytic processing (Besse et al., 2011). Besides this essential role in Hh/Gli regulation, Rpgrip1l knock-down in zebrafish leads to CE defects, a process that has been linked to PCP signaling (Khanna et al., 2009).

Here, we used different model systems to investigate the function of Rpgrip1l in planar polarity and its interaction with the PCP pathway. We showed that Rpgrip1l is required for planar polarity in sensory hair cells of the mouse cochlea, in CE during zebrafish gastrulation and neurulation, and in the polarized positioning of motile cilia of the zebrafish floor plate. In the zebrafish floor plate and in cultured mammalian kidney cells, we showed that the enrichment of dishevelled at the base of the cilium was dependent on Rpgrip1l. Rpgrip1l positively controls dishevelled levels, and Dvl overexpression in zebrafish embryos rescues CE and floor plate cilia localization in rpgrip1l morphants. We provide a model for Rpgrip1l function in a ciliopathy protein complex modulating dishevelled stability.

**Results**

Rpgrip1l is essential for localization of the cilium along the PCP axis in cochlear hair cells

Rpgrip1l is present at the kinocilium transition zone in E18.5 cochlear hair cells (Fig. 1 A). To investigate the involvement of Rpgrip1l in PCP, we first analyzed the polarity of cochlear hair
cells in the Ftm mouse mutant line, which harbors a null mutation in the Ftm/Rpgrip1l gene (Vierkotten et al., 2007). Because Ftm mice die at birth, cochlear defects were analyzed in E18.5 fetuses, when PCP is already established in the cochlea. Ftm<sup>−/−</sup> cochleae were slightly shorter than that of control (Ftm<sup>+/+</sup> or Ftm<sup>+/−</sup>) fetuses (Fig. 1 C), suggesting a mild CE defect. The four rows of hair cells were slightly disorganized, with additional hair cells between the three rows of outer hair cells (OHCs; Fig. 1, H and I). In scanning electron microscopy, the stereociliary hair bundles appeared less cohesive and did not present the normal V shape (Fig. 1, B and D–G). Misorientation of the hair bundles was observed in few cells (Fig. 1 E, arrows). This was confirmed and quantified by staining F-actin with phalloidin (Fig. 1, H and I). At the base of Ftm<sup>−/−</sup> cochleae, 4.8% of sensory cells had a misoriented bundle (over 30° of angle deviation; Fig. 1 I, arrows) and 2.5% had a round bundle (n = 522
cells from 4 Ftm<sup>−/−</sup> fetuses), whereas control cells had 1% mis-oriented and no round bundles (n = 595 cells from 5 control fetuses). In control cochleae, the kinocilium is positioned at the vertex of the hair bundle (Fig. 1, B, K, and L1) and is connected to the stereocilia by extracellular links (Jones and Chen, 2008). In Ftm<sup>−/−</sup> cochleae, the kinocilium was incorrectly positioned and oriented, and was disconnected from the stereociliary bundle (Fig. 1, G, L, and L1). In addition, ciliogenesis was impaired in a small proportion of cells: the kinocilium was either shorter in 16% or absent in 12% of the cells (n = 193; Fig. 1, L and L1).

We then performed γ-tubulin staining in order to accurately position the two centrioles within the cells. In control hair cells, the two centrioles were close to each other and aligned along the proximo-distal axis (n = 122 cells from 3 control fetuses) (Fig. 1, B, J, M, and M1). In two thirds of hair cells in Ftm<sup>−/−</sup> cochleae, the centrioles were not in the distal quadrant and/or not properly aligned along the proximo-distal axis (n = 83 cells from 3 Ftm<sup>−/−</sup> fetuses; Fig. 1, J, N, and N1). In rare cases (2 of 100 cells), the two centrioles were far from each other. Our results on kinocilium and centriole position defects in hair cells of Ftm<sup>−/−</sup> cochleae indicate a role for Rpgrip1l in PCP.

Zebrafish rpgrip1l morphants display planar polarity defects

To further study the involvement of Rpgrip1l in PCP, we turned to the zebrafish model. We identified a single orthologue of the human RPGRIP1L gene in the zebrafish genome (Ensembl prediction GenBank/EMBL/DDBJ accession no. XM_002666951).
and determined its full-length coding sequence (GenBank accession no. JN051142). The zebrafish rpgrip1l gene encodes a 1259-amino acid protein with 51% identity and 67% similarity with the human RPGRIP1L protein. Zebrafish rpgrip1l is expressed both maternally and zygotically, widely at early somite stages, then mainly in the central nervous system after 24 hours post-fertilization (hpf; Fig. S1). Injection of RNA coding for a tagged form of Rpgrip1l revealed that the protein was enriched in the transition zone of the cilium in all monociliated cell types examined, including the neural tube, floor plate, and notochord cells (Fig. 2 A). We next performed rpgrip1l loss-of-function experiments using two antisense morpholinos, targeting the translation start site (Mo-1) and a region 55–31 nucleotides upstream of the translation start site (Mo-55). Morphological examination of the morphants showed shortened body axis, kinked notochord/neural tube, and malformed somites at early somite stages, abnormal body curvature after 24 hpf, and hydrocephaly at 2 dpf (Fig. 2, B–K). The embryos were separated into three classes depending on the severity of the axis elongation phenotype, as measured by the body gap angle (bga; angle between the tip of the head and the tip of the tail) at the 10–12-s stage (Fig. 2, B–D and P). Morphants that survived up to 5 dpf were shorter and had a curved body (Fig. 2, H and I), as found in PCP mutants (Jessen et al., 2002). Shorter axis and curved body strongly suggested the existence of CE defects, which was confirmed using molecular landmarks (Fig. S2, A–F). The number and size of cilia, the localization of several ciliary proteins within the cilium shaft, and the structure of basal bodies appeared unchanged in most tissues of rpgrip1l morphants as compared with controls (Fig. 3, A–C; and Fig. S2, G–Q). Because both morpholinos gave similar and dose-dependent phenotypes, more robust with Mo-1 than with Mo-55 (unpublished data), Mo-1 was used in most experiments (hereafter called Mo-Rpgrip1l). Co-injection of human RPGRIP1L mRNA with the morpholinos rescued the CE phenotype, with a significant decrease in the number of affected embryos and the severity of the defects (Fig. 2 P), as well as hydrocephaly (Fig. 2 Q), demonstrating that these defects were specific for the loss of Rpgrip1l function. Rpgrip1l morphants also displayed defective left–right asymmetry, as illustrated by randomized expression of the early laterality marker spw (Fig. 2, L–O and R). Thus, rpgrip1l morphants display a ciliary phenotype characterized by abnormal CE and laterality, without general ciliary defects, allowing us to study Rpgrip1l function in planar polarity independently of its role in ciliogenesis.

In addition to CE, another manifestation of PCP in the early zebrafish embryo is the posterior position of the basal body in floor plate cells, which defects are associated with aberrant motile cilia tilting (Borovina et al., 2010). Because rpgrip1l transcripts are enriched in floor plate cells, we investigated its function in this process. In control embryos from the 14-s stage onward, 80% of the cells presented posterior basal bodies, as
assessed by γ-tubulin staining in embryos injected with membrane-associated GFP to detect cell membranes (Fig. 3, D, G, and J). In rpgrip1l morphants, the posterior bias in basal body position was significantly reduced (Fig. 3, E, F, H, I, and J). These data demonstrate that Rpgrip1l is required for positioning the cilium along the antero-posterior (planar polarity) axis in the zebrafish floor plate.

Rpgrip1l acts in PCP by stabilizing dishevelled

We analyzed the functional interaction of rpgrip1l with several PCP genes and with inversin by performing combined knockdown in zebrafish embryos. Co-injection of low doses of Mo-Rpgrip1l with morpholinos specific for vangl2 (Mo-Vangl2), prickle1 (Mo-Pk1), for all three zebrafish dvl genes (Mo-3Dvl; Ségalen et al., 2010), or for inv (Mo-inv; Sayer et al., 2006) led to an increase in the severity of the axis elongation phenotype as compared with individual morphants (Fig. 4 A and Fig. S3, A–C). These results suggest either that Rpgrip1l and these proteins functionally interact during zebrafish CE, or that they act in parallel to promote common cellular processes.

Because dishevelled is a major actor of the Wnt pathways whose stability is modulated by the Rpgrip1l interactors nephrocalcin-4 and inversin (Simons et al., 2005; Burcklé et al., 2011), we investigated a direct role of Rpgrip1l on its stability. For that purpose, we co-injected an RNA coding for a Myc-tagged form of Xenopus Dvl2 with Mo-Rpgrip1l into zebrafish embryos and analyzed by Western blots the amounts of Dvl2-Myc protein assessed by γ-tubulin staining in embryos injected with membrane-associated GFP to detect cell membranes (Fig. 3, D, G, and J). In rpgrip1l morphants, the posterior bias in basal body position was significantly reduced (Fig. 3, E, F, H, I, and J). These data demonstrate that Rpgrip1l is required for positioning the cilium along the antero-posterior (planar polarity) axis in the zebrafish floor plate.

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recovered at different stages (Fig. 4 B). The amounts of Dvl2-Myc were not significantly different between control and rpgrip1l morphants at 80% epiboly (8.5 hpf). However, at 12 s (15 hpf), rpgrip1l morphants displayed a 40% reduction in Dvl2-Myc levels compared with controls, suggesting that Rpgr1p1l stabilized dishevelled. Dvl protein levels are known to be important for zebrafish CE (Angers et al., 2006) and the knock-down of all three zebrafish dvl genes led to defects in asymmetric basal body localization in floor plate cells (Fig. S3 D). To test whether Rpgr1p1l function in PCP is mediated by dishevelled stabilization, we performed a rescue experiment by co-injecting Mo-Rpgr1p1l together with an RNA coding for a GFP-tagged form of Drosophila dishevelled (Dsh-GFP). This led to a significant rescue of the CE phenotype (Fig. 4 C), as well as of the posterior localization of basal body in floor plate cells (Fig. 3 K). Together, these data show that in these two processes, Rpgr1p1l acts at least in part by stabilizing dishevelled.

**Rpgr1p1l localizes dishevelled to the basal body of zebrafish floor plate cells**

To test whether dishevelled stability was affected uniformly in the embryo, or differentially in specific cell types or subcellular compartments, we performed whole-mount GFP immunofluorescence on embryos injected with Dsh-GFP mRNA. NLSLacZ mRNA, which codes for a nuclear form of β-galactosidase (βgal), was co-injected as a control of the presence of injected RNA in the cells. In control embryos at 12 s, Dsh-GFP concentrated in many cell types including the floor plate and notochord cells, in apical granules located at or close to the cilia (in 61% of the injected embryos, n = 39; Fig. 4 D). In double staining for βgal and GFP, all of the βgal-positive cells (n = 86 cells, 9 embryos) in the floor plate displayed Dsh-GFP granules at the cilia base, in the immediate vicinity of the basal body (Fig. 4, F and H). Co-injection of Dsh-GFP with Rpgr1p1lMyc mRNA did not reveal colocalization between GFP and Myc, indicating that the dishevelled and Rpgr1p1l proteins were enriched in adjacent structures at the cilia base (Fig. 4 I). In 95% of rpgrip1l morphants (n = 41), Dsh-GFP granules were either absent or severely reduced in number and size (Fig. 4 E) and found at the base of the cilia in only 9% of the βgal-positive cells (n = 78, 7 embryos; Fig. 4 G). In contrast, in inv morphants the cytoplasmic pool of Dsh-GFP was increased and the basal body pool was not grossly affected. In combined inv, rpgr1p1l morphants the ciliary pool of Dsh-GFP was absent and the cytoplasmic pool was still present although reduced compared with inv morphants (Fig. S3, E–J). A Vangl2-GFP fusion protein showed a different behavior, with a homogenous localization at cell membranes in both controls and rpgr1p1l morphants (Fig. S3, K–R). Together, these experiments strongly suggest that Rpgr1p1l is required to maintain dishevelled levels in zebrafish floor plate cells and that it acts primarily but not exclusively on a pericentriolar pool of dishevelled.

**Rpgr1p1l stabilizes dishevelled at the cilium base of mammalian kidney cells**

Overexpressed dishevelled protein tends to form aggregates whose physiological significance is unclear (Schwarz-Romond et al., 2005). To investigate Rpgr1l function in the localization of endogenous dishevelled, we used polarizing monolayered MDCK cells, in which we have previously shown the endogenous expression of Dvl2, Dvl3, and RPGRIP1L at the base of the cilium (Burcklé et al., 2011). To test whether RPGRIP1L was required for dishevelled localization, we generated stable RPGRIP1L knock-down MDCK cell lines with two different shRNA sequences (RPGRIP1L-KD-sh1/2). Silencing of RPGRIP1L led to ~40% residual expression of mRNA and protein expression compared with control cell lines, as assessed by quantitative RT-PCR and Western blotting (Fig. S4 A). In both RPGRIP1L-KD cell lines, RPGRIP1L depletion resulted in a reduction of both the number and the length of cilia, as well as in a disruption of the pericentriolar material (Fig. S4, B–D). We then examined whether endogenous dishevelled localization was altered upon RPGRIP1L depletion. In most polarizing control MDCK cells, Dvl2 and Dvl3 were detected in and around the basal body, and sometimes in the cytoplasm (Fig. 5 A, diagrams). In contrast, Dvl2 or Dvl3 pericentriolar staining was lost or severely reduced in a large majority (75–80%) of RPGRIP1L-depleted cells (Fig. 5 A). This correlated with a global reduction in total Dvl2 and Dvl3 protein amounts, without modification of mRNA levels (Fig. 5, B and C), suggesting that Rpgr1p1l stabilizes dishevelled proteins in these cells. The decrease in Dvl2 and Dvl3 levels at the basal body was rescued upon reexpression of RPGRIP1L-Myc in RPGRIP1L-KD cells (Fig. S5, A–C). Using other experimental conditions allowing the detection of Dvl2 and Dvl3 at the cell membrane, we showed that the membrane localization of dishevelled was still observed after RPGRIP1L depletion (Fig. S5 D). Because inversin and nephrocystin-4 have been shown to bind and destabilize dishevelled (Simons et al., 2005; Burcklé et al., 2011), we tested whether RPGRIP1L depletion prevented their localization at the base of the cilium. RPGRIP1L depletion led to a reduction of the pericentriolar accumulation of inversin and nephrocystin-4 along with a disruption of γ-tubulin staining in ~50% of the cells. However, in the 50% of RPGRIP1L-KD cells where γ-tubulin staining remained, inversin and nephrocystin-4 as well as two other ciliopathy proteins, nephrocystin-3 and CEP290, accumulated around the basal body, similar to control MDCK cells (Fig. 5, D and E; and Fig. S4, D–G). These results suggest that RPGRIP1L depletion in these cells leads to a drastic and widespread reduction of Dvl accumulation at the cilium base, which partially correlates with a destabilization of the pericentriolar material.

**Rpgr1p1l acts in a complex with inversin and nephrocystin-4 to modulate dishevelled stability**

We next investigated the mechanisms by which RPGRIP1L stabilizes dishevelled. Inhibition of the proteasome by treatment of RPGRIP1L-KD cells with clasto-lactacystin led to a rescue of Dvl3 amounts to levels comparable to that found in control PSICOR cells (Fig. 6 A), strongly suggesting that RPGRIP1L stabilizes dishevelled by preventing its proteasomal degradation. To determine whether RPGRIP1L is sufficient for dishevelled stabilization, we overexpressed RPGRIP1L-Myc in HEK293T
We found that inversin coimmunoprecipitated both RPGRIP1L and Dvl2, and that coexpression of RPGRIP1L and Dvl2 with inversin increased the levels of immunoprecipitated Dvl2, in a manner consistent with the stabilization of Dvl2 by RPGRIP1L in total lysates (Fig. 6 C). Moreover, when coexpressed, nephrocystin-4 coimmunoprecipitated both Dvl2 and RPGRIP1L (Fig. 6 D). We next tested in this assay dishevelled stabilization by mutant forms of these proteins in HEK293T cells and compared its effect to that of inversin-Flag (Inv-Flag) overexpression. Whereas inversin overexpression strongly decreased Dvl2-Myc protein levels, RPGRIP1L-Myc overexpression led to a significant increase in Dvl2-Myc levels (Fig. 6 B).

To determine whether RPGRIP1L, inversin, nephrocystin-4, and dishevelled could act together in a same complex or whether Rpgrip1l would compete with inversin and nephrocystin-4 for dishevelled binding, we performed coimmunoprecipitation experiments in HEK293T cells with tagged forms of these proteins. We found that inversin coimmunoprecipitated both RPGRIP1L and Dvl2, and that coexpression of RPGRIP1L and Dvl2 with inversin increased the levels of immunoprecipitated Dvl2, in a manner consistent with the stabilization of Dvl2 by RPGRIP1L in total lysates (Fig. 6 C). Moreover, when coexpressed, nephrocystin-4 coimmunoprecipitated both Dvl2 and RPGRIP1L (Fig. 6 D).
of RPGRIP1L identified in cases of Joubert type B syndrome and known to interact less efficiently with nephrocystin-4 (Delous et al., 2007). We found that these mutant forms had a reduced capacity to stabilize Dvl2 (Fig. 6 E), consistent with their reduced capacity to rescue CE defects in zebrafish (Khanna et al., 2009).

**Discussion**

In this paper we investigated the function of the ciliopathy protein Rpgrip11 in planar polarity. We show that, in the mammalian cochlear sensory epithelium and in the zebrafish floor plate, Rpgrip11 is required for correct positioning of the basal body along the PCP axis. We demonstrate that Rpgrip11 is essential for stabilizing dishevelled in different experimental systems and that in the zebrafish floor plate this stabilization of dishevelled by Rpgrip11 mediates basal body positioning. Finally, our experiments in cell culture suggest that Rpgrip11 acts in a complex with nephrocystin-4 and inversin to finely regulate dishevelled stability.
Our data uncover a function for Rpgrip1l in several PCP processes characterized by a stereotyped position of the centrioles along the PCP axis (Jones and Chen, 2008; Borovina et al., 2010; Sepich et al., 2011). Rpgrip1l could act at several levels in these processes. It could be a general modulator of planar polarity, controlling the polarized localization of core PCP pathway proteins. Alternatively, it could be involved in interpreting a preexisting polarity information that would result in the asymmetric positioning of the basal body. Finally, it could act in parallel to and independently of the PCP pathway. Our results are not in favor of the first possibility. First, in cochleae, Rpgrip1l mutation leads to a mild disruption of the orientation of the stereociliary bundles, confirming that distinct planar polarity events in these cells can be uncoupled in ciliary mutants (Jones et al., 2008; Sipe and Lu, 2011). Moreover, the rescue of CE defects and of ciliogenesis in the zebrafish floor plate by dishevelled shows that Rpgrip1l function is mediated at least in part by dishevelled stabilization. This places Rpgrip1l either downstream of, or in parallel to, cortical asymmetric cues, and upstream of intracellular basal body positioning. In the Drosophila wing (Goodrich and Strutt, 2011) as in the Xenopus skin (Mitchell et al., 2009), dishevelled acts cell-autonomously to establish polarity within cells, while Strabismus/Vangl and Frizzled transmembrane proteins also act in the coordination of planar polarity between adjacent cells. Thus, we propose that Rpgrip1l, by maintaining dishevelled levels, is specifically involved in one planar polarity process, i.e., the asymmetric localization of the basal body.

The function of dishevelled in basal body positioning in the zebrafish floor plate is consistent with what is described in the mouse node (Hashimoto et al., 2010). In the cochlea, dishevelled loss of function perturbs stereocilia orientation (Wang et al., 2006), whereas Rpgrip1l loss of function does not. In this system, dishevelled has been described in a cortical crescent located at the distal membrane of the hair cells, where the centrioles are anchored (Wang et al., 2006; Etheridge et al., 2008; Sipe and Lu, 2011). It would be very interesting to know whether Rpgrip1l is required for dishevelled stability in this system, and we are currently investigating this question. Also, dishevelled effectors in asymmetric basal body localization in the zebrafish floor plate remain to be identified. A candidate is Rac1, which is a downstream effector of dishevelled in other cellular contexts (Gao and Chen, 2010; Ishida-Takagishi et al., 2012), and whose inhibition prevents both posterior positioning of the basal body in node cells (Hashimoto et al., 2010) and positioning of the kinocilium on the distal side of cochlear hair cells (Sipe and Lu, 2011).

Our data point to a functional interaction between dishevelled and Rpgrip1l, which leads to dishevelled stabilization. What are the mechanisms of this functional interaction? Our experiments in HEK293T cells suggest that a physical interaction between Rpgrip1l, nephrocystin-4, inversin, and dishevelled can occur. The Rpgrip1l–dishevelled interaction is likely to be indirect, through a macromolecular complex comprising nephrocystin-4 and inversin (Simons et al., 2005; Delous et al., 2007; Burcklé et al., 2011). Rpgrip1l-dependent integrity of the cilium does not seem to be required because Rpgrip1l stabilizes dishevelled in HEK293T cells that do not form a cilium. We favor an alternative possibility, in which Rpgrip1l would be required for the stability and/or function of a pericentriolar platform involved in dishevelled stabilization. This is suggested by the reduction of the pericentriolar material and associated proteins in MDCK cells upon RPGRIP1L knock-down. Although Rpgrip1l is mainly found at the transition zone in many cell types, it is known to interact with pericentriolar proteins (Coene et al., 2011).

Several ciliopathy proteins, in particular the Rpgrip1l interactors inversin and nephrocystin-4, have been shown to target cytoplasmic dishevelled for proteasomal degradation (Simons et al., 2005; Burcklé et al., 2011; Wallingford and Mitchell, 2011). Here we show that, in contrast, Rpgrip1l protects dishevelled from proteasomal degradation. How is the balance in dishevelled stability achieved, and is the subcellular localization of the complex important for this balance? Our data show that Rpgrip1l does not compete with binding of nephrocystin-4 or inversin to dishevelled. Rpgrip1l could act by anchoring an inversin–nephrocystin-4–dishevelled complex to the basal body. Indeed, in C. elegans neurons, nephrocystin-4 localization at the ciliary transition zone depends on Rpgrip1l (Williams et al., 2011). Rpgrip1l could also modify the activity of nephrocystin-4 and inversin on dishevelled, allowing a precise tuning of dishevelled stability. The stoichiometry or the conformation of the different ciliary proteins in the complex, depending on the cellular context, could modulate the recruitment and/or the activity of proteins involved in proteasomal degradation. For instance, it could modulate the recruitment or activity of the ubiquitin ligase APC-C, which interacts with both dishevelled and inversin and regulates dishevelled stability (Morgan et al., 2002; Ganner et al., 2009). These two different mechanisms, localization of the complex to the pericentriolar region, and modulation of the activity of the complex, are not mutually exclusive and could both participate in fine-tuning dishevelled stability.

In conclusion, we describe here a novel, central function of an Rpgrip1l-dependent nephrocystin complex in stabilizing dishevelled, and we provide compelling evidence that this function is required for planar localization of the basal body. In mice, altered Wnt-PCP signaling results in kidney cysts appearing before birth, associated with CE defects in the elongating renal tubules (Karner et al., 2009). Thus, the function of Rpgrip1l in planar polarity may help interpret defects such as kidney dysfunctions found in mouse Ftm mutants and in humans presenting RPGRIP1L mutations. In this respect, our observation that RPGRIP1L mutations found in Joubert syndrome type B fail to stabilize dishevelled highlights possible physiopathological mechanisms occurring in ciliopathies.

**Materials and methods**

**Mouse and zebrafish strains**

Ftm mutant mice were maintained in a C57B16/J background and geno-typed as described previously [Vierkotten et al., 2007]. Embryonic day 0.5 [E0.5] was defined as noon on the day of vaginal plug detection. We used zebrafish wild-type AB or [FL x AB] hybrid strains and the Tg (β-actin:: Arl13b-GFP) transgenic line, which expresses a GFP fusion protein with the...
cilary protein Actb113b, under the control of the strong ubiquitous β-actin promoter (Barovina et al., 2010). Zebrafish were raised and maintained as described previously (Kimmel et al., 1995). Embryos were staged according to the number of hours (hpf) or days (dpf) postfertilization at 28°C. Alternatively, for staging epiboly-stage embryos we used the percentage of yolk covered by the embryo (% epiboly), for somite-stage embryos, the number of somites was used (Kimmel et al., 1995).

Construction of Rpgrip1l expression plasmids

pCS2-RPGRIP1L (human) was constructed by transferring an EcoRI-XbaI fragment from pSPORT-RPGRIP1L, containing the whole human cDNA (Delous et al., 2007) into pCS2+. The zebrafish Rpgrip1l cdNA sequence was reconstructed from the IRAKp61 G13298 clone and RT-PCR fragments and was cloned into pCS2+. To obtain Myc-tagged zebrafish Rpgrip1l [Rpgrip1l-Hmyc], 5 Myc epitope tags were added in frame in the C terminal, removing the last 11 amino acids of the Rpgrip1l protein and the 3′-UTR.

Zebrafish microinjection

1 mL RNA- and/or morpholino-containing Danaua medium was injected into 1- to 4-cell stage embryos. Rpgrip1l translation-blocking morpholinos (Gene Tools) were designed to target either a region located at 1-24 bp downstream (Mo-1: 5′-AGTTTCACTACGAGCAGAAACAATG-3′) or 55-31 bp upstream (Mo-55: 5′-GACCACAGCGAGCTTGCTAAAGA-3′) of the translation start site. For functional interaction studies, morpholinos targeting Vangl2 (5′-TGTACCAGCTATTCCATGC-3′; Otto et al., 2003), Prickle1 (5′-GCCACCCTGATTTCTCAGGATATCT-3′; Carreño-Barbosa et al., 2003), the zebrafish Dvl3 (Dvl3: 5′-TATAAATATCCGTCGATGCATC-3′; Sály et al., 2006) and Dvl2-like (5′-GGAATATGTTCCCATGACA-3′; Ségal et al., 2010), and inv (splice morpholino, 5′-GGAACATAGATCCATCTGC-3′; Sayer et al., 2006) were used. mRNA was synthesized in vitro from pCS2+ (or pSP6 T7-11), plasmid containing the mMessage mMachine kit (Ambion) and injected at the following doses: Dvl2-GFP (Drosophila dishelled, 10 pg for subcellular localization, 12 pg for rescue experiments), Ras-GFP (mbGFP: membrane localization signal of Ras protein fused to eGFP, 60 pg; Ségal et al., 2010), mCherry (two membrane localization signals fused to GFP, 25 pg; Megason and Fraser, 2003; Megason, 2009), GFP-centrin1 (human centrin1; 15 pg), mouse GFP-Vangl2 (mouse Vangl2; 25 pg), Myc-Dvl2 (Xenopus Dvl2; 15 pg), human Rpgrip1l (100 pg), rpgrip1l-myc (7 pg), and NLSlacZ (60 pg; Smith and Harland, 1991). All fused proteins are full length unless otherwise stated, and the position of the tag in the name of the plasmid reflects the position in the fusion protein.

Establishment of lentiviral cell lines and transient transfections

shRNA constructs targeting the coiled-coiled domain 3 (sh1 and 5) and sh2) of the canine RPGRIP1L mRNA sequences (sh1: 5′-GGATAAGTAAAATCTTCGATGCACCA-3′) were designed and cloned into the lentiviral pSICOR vector, lentiviruses were produced in HEK293T cells, and MDCK type II cells were infected, as described previously (Delous et al., 2009). MDCK control and Rpgrip1l-KD cells were cotransfected with human full-length NPHP4-V5 C-terminal probe and diluted 1:400. After a final overnight wash at 4°C, coelacanth were detached from the vesicle with fine forceps, after removing nervous connections and underlying mesenchyme. Coelacanth were then transferred on a glass coverslip, mounted in Vectashield (Vector Laboratories) and imaged using a confocal microscope (model TCS SPE II; Leica) using an HCX Plan Apochromat 63×/1.40-0.60 oil objective at room temperature. All channels were acquired by sequential scanning on LASSAF software (Leica). For scanning electron microscopy (SEM), inner ears were removed from E18.5 heads in 1.22x PBS (pH 7.4) and fixed overnight with 2% glutaraldehyde in 0.61x PBS (pH 7.4) at 4°C. They were washed several times in cold PBS and pinned with pinza on a dissection dish. Cartilages surrounding the cochlea were removed with fine forceps. The Reissner’s membrane was then peeled off to expose the sensory epithelium. Coelacanth still attached to the vestibule were then transferred to a 2-mL Eppendorf tube (Montcouquiol et al., 2008). After a permeabilization step of 15 min in PBS, 0.1% Triton X-100 at 4°C (link 80°C), all samples were fixed overnight in 1% Triton X-100, 10% PBS, 0.1% Triton X-100, 10% normal goat serum. Coelacanth were incubated overnight with the following primary antibodies: mouse anti-acetylated tubulin (catalog no. T6793, 1:250; Sigma-Aldrich), mouse anti-γ-tubulin (catalog no. GTU-88, 1:1,000; Sigma-Aldrich), or rabbit anti-Rpgrip1l (Vierkotten et al., 2007), Phalloidin–Alexa Fluor 488 (Molecular Probes) was added to stain F-actin. All secondary antibodies were washed twice in PBS and diluted 1:400. After a final overnight wash at 4°C, coelacanth were detached from the vestibule with fine forceps, after removing nervous connections and underlying mesenchyme. Coelacanth were then transferred on a glass coverslip, mounted in Vectashield (Vector Laboratories) and imaged using a confocal microscope (model TCS SPE II; Leica) using an HCX Plan Apochromat 63×/1.40-0.60 oil objective at room temperature. All channels were acquired by sequential scanning on LASSAF software (Leica). For scanning electron microscopy (SEM), inner ears were removed from E18.5 heads in 1.22x PBS (pH 7.4) and fixed overnight with 2% glutaraldehyde in 0.61x PBS (pH 7.4) at 4°C. They were washed several times in 1.22x PBS, and coelacanth were dissected as indicated above and postfixed for 15 min in 1.22x PBS containing 1% Os4. Fixed samples were then prepared for SEM as described previously (Besse et al., 2010). Samples were observed under a scanning electron microscope (model S260; Cambridge) at 10 keV.
2% goat serum), and incubated overnight with primary antibodies and 2 h with secondary antibodies. Primary antibodies were mouse anti-GFP (catalog no. 1181440001, 1:100; Rocken), polyclonal anti-GFP (1:150; Molecular Probes), mouse anti-acetylated tubulin (catalog no. T6793, 1:250; Sigma-Aldrich), mouse anti-α-tubulin (catalog no. GTU-88, 1:1,000; Sigma-Aldrich), rabbit anti-β-galactosidase (catalog no. 55976, 1:500; Cappel), mouse anti-P53 (catalog no. 2276, 1:200; Cell Signaling Technology), rabbit anti-Vanich (Montavjial et al., 2006), rabbit anti-DS Red (catalog no. 632496, 1:500; Takara Bio Inc.), rabbit anti-MKS1 (catalog no. HPA021812, 1:500; Sigma-Aldrich), and rabbit anti-Cep290 (catalog no. ab58470, 1:100; Abcam). All secondary antibodies were from Molecular Probes and diluted 1:400. Nuclei were stained with DAPI (Invitrogen). Zebrafish embryos were flat-mounted in Vectashield mounting medium (Vector Laboratories) and imaged under a motorized microscope (model DM6000B, Leica). Confocal images were captured using a confocal microscope (model TCS SP5 II; Leica) using an HCX Plan Apochromat 63x/1.40–0.60 oil objective (3x zoom) at room temperature. All channels were acquired by sequential scanning on LAS-AF software (Leica). For basal body localization, we acquired lateral views of the whole region corresponding to the floor-plate blastomeres. We then selected 18- to 14-s embryos previously injected with RNAs encoding a membrane-bound GFP. Each cell was divided into three regions (anterior, median, posterior) and the position of basal bodies was determined in each floor plate cell. Basal bodies and cell membrane confocal planes were merged using the position of basal body in each floor plate cell determined in stock. Images shown correspond to one optic section of 0.29 µm. Contrast and brightness of images were adjusted using Photoshop software (Adobe).

RT-PCR and in situ hybridization

For RT-PCR analysis, sets of primers specific to canine RT-PCR and in situ hybridization were designed. Whole-mount in situ hybridization was performed according to standard protocols and imaged on a stereomicroscope (model MZ16; Leica) equipped with a camera (model DFC 425; Leica).

Online supplemental material

Fig. S1 shows zebrafish rpgrip1l gene expression. Fig. S2 shows no apparent defect in dorso–ventral (DV) and antero–posterior (AP) patterning and in ciliature and basal body integrity, in rpgrip1l morphants. Fig. S3 shows interaction of rpgrip1l with PCP genes and with inversin. Fig. S4 shows ciliature and basal body integrity in MDCX cells upon Rpgrip1l depletion. Fig. S5 shows Rpgrip1l depletion in MDCX cells: rescue and dishevelled localization at the membrane.

Submitted: 2 November 2011
Accepted: 6 August 2012

References


Figure S1.  **Zebrafish rpgrip1l gene expression.** (A) RT-PCR analysis of rpgrip1l expression in zebrafish embryos at different stages post-fertilization. (B–D') In situ hybridization with rpgrip1l antisense (B, C, and D) or sense (B', C', and D') probes at the 10-s (14 hpf; B and B'), 14-s (16 hpf; C and C'), and 24-hpf (D and D') stages. Rpgrip1l is expressed widely and weakly in 10-s and 14-s stage embryos, and at higher levels in the otic vesicle, the floor plate and the notochord. At 24 hpf, rpgrip1l is enriched in the central nervous system. Methods: RT-PCR analysis of rpgrip1l was performed in standard conditions, using forward 5'-GCTTTAATCGACTAAACGATG-3' and reverse 5'-TGCCTGAGAGCTGCTTTCG-3' primers. Control PCR primers for zebrafish ef1α were: ef1α-forward 5'-CTTCTCAGGCTGACTGTGC-3' and ef1α-reverse 5'-CCGCTAGCATTACCCTCC-3'. For in situ hybridization, probes spanning two regions of the rpgrip1l cDNA, nucleotides 1004–2480 and 2348–4020, gave similar results. Bars, 250 mm. bp, base pairs.
Figure S2. No apparent defect in dorso-ventral (DV) and antero-posterior (AP) patterning, and in cilium and basal body integrity, in rpgrip1l morphants. (A and B) Animal views of noninjected control (A) or Mo-Rpgrip1l–injected (B) embryos at the 50% epiboly stage after in situ hybridization (ISH) with a *chordin* (*chd*) probe. (C) Diagram illustrating the mean length of the *chd* domain (portion of the embryo perimeter presenting *chd* expression; arbitrary units) in control and Mo-Rpgrip1l–injected embryos. 5 control embryos and 17 Mo-Rpgrip1l–injected embryos were measured. No significant difference in length was found (Student’s t-test, \( t > 0.1; \) ns, non significant). *Chd* is expressed in a dorsal mesodermal domain of similar size and shape in controls and rpgrip1l morphants, showing that early dorso-ventral patterning is normal. (D and E) Dorsal views of flat-mounted noninjected control (D) or Mo-Rpgrip1l–injected (E) embryos at the 12-s stage after in situ hybridization with *krox20/egr2b*, *pax2a*, and *myoD* probes. *Krox20* is expressed in two hindbrain transverse domains, r3 and r5; *pax2a* expression marks the midbrain–hindbrain boundary and the optic stalk, and *myoD* is expressed in the somites and adaxial cells. Together, these three gene expression patterns provide landmarks to assess CE and AP patterning defects. (F) Diagram illustrating the mean length-to-width ratio (ratio of the AP length of the adaxial domain to the embryo width at the level of the fifth somite) of control and rpgrip1l morphant embryos. In these morphants, all three expression domains are present and at the right place. However, all domains are wider along the DV axis and shorter along the AP axis in rpgrip1l morphants as compared with controls. The different AP landmarks of the neural plate: optic stalk, midbrain–hindbrain boundary, and rhombomeres 3 and 5 are also closer to each other. A widened and shorter embryo is indicative of CE defects and is a feature of the knock-down of PCP genes (Carreira-Barbosa et al., 2003) and of several ciliary genes (Ross et al., 2005; Gerdes et al., 2007). Together, these data show that early DV and AP patterning are not affected and that CE is perturbed upon Rpgrip1l depletion. 

**G** and **H**. Lateral view of the floor plate of control (**G**) and Mo-Rpgrip1l–injected (**H**) Arl13bGFP transgenic embryos stained with antibodies specific for GFP and Ac-Tub and counterstained with DAPI. This transgenic line expresses an Arl13b-GFP fusion protein in all zebrafish embryonic cells and thereby permits to visualize the shaft of primary cilia, in which the small GTPase Arl13b trafficks (Borovina et al., 2010). Accumulation of Arl13bGFP in the cilium of floor plate cells appears similar in control (**G**, \( n = 12 \)) and rpgrip1l morphant (**H**, \( n = 12 \)) embryos. (I–L) Control (**I** and **J**) and Mo-Rpgrip1l–injected (**K** and **L**) embryos were stained by immunofluorescence with antibodies specific for MKS1 (I and J), CEP290 (K and L), \( \gamma \)-Tub, and Ac-Tub. In zebrafish floor plate cells, Mks1 and Cep290 are present in granules along the cilium shaft. This localization is not significantly modified in rpgrip1l morphants. We counted the number of cilia presenting Mks1 or Cep290 staining. Mks1 granules were found on 79% of floor plate cilia in controls (\( n = 71 \) cells in 4 embryos) and on 75% of floor plate cilia in morpholino-injected embryos (\( n = 80 \) cells in 4 embryos). Cep290 granules were found on 48% of floor plate cilia in controls (\( n = 72 \) cells in 3 embryos) and on 60% of floor plate cilia in Mo-Rpgrip1l–injected embryos (\( n = 74 \) cells in 4 embryos). (M and N) Centrin2-GFP (cm2-GFP) mRNA injected control (**M**, \( n = 4 \)) and rpgrip1l morphant (**N**, \( n = 4 \)) embryos were subjected to immunofluorescence for GFP (centrioles) and Ac-Tub (cilia). Neither defects in centriole engagement nor an abnormal number of centrioles per cell were detected (71 cells in 4 control embryos and 80 cells in 4 rpgrip1l morphants were analyzed). (O–Q) Cm2GFP RNA (15 pg) was co-injected with mb-Cherry (membrane-associated mCherry) RNA (22 pg) in control (**O**) or rpgrip1l morphant (**P** and **Q**) embryos in order to detect cell membranes and to correlate defects in asymmetric basal body positioning with eventual centriolar disengagement. The results show that centrioles are always in pairs and correctly assembled in close proximity to the apical membrane, even in cells that display anterior or medial basal bodies (**P** and **Q**). All pictures in **G**–**Q** are lateral views of the floor plate of flat-mounted 18-s embryos. All antibody stainings are indicated in the figure and color coded. Nuclei were stained with DAPI when indicated. Bars: [A, B, D, and E] 250 µm; [G–Q] 10 µm.
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[Figure S3. Interaction of rpgrip1l with PCP genes and with inversin. (A–C) Diagrams illustrating the functional interaction between rpgrip1l and the vangl2 (A), prickle1 (B), and inv (C) genes in axis elongation. Co-injection of MoRpgrip1l with morpholinos specific for vangl2 (A, Mo-vangl2), prickle1 (B, Mo-pk1), or inv (C, Mo-inv) led to an enhancement of the morphant phenotype. For each lane, the injected morpholinos are indicated on the right. In A, n = 25 control, 21 Mo-Rpgrip1l–injected, 17 Mo-Vangl2–injected, and 22 co-injected embryos, from a representative experiment out of 4 repeats. In B, n = 11 control, 19 Mo-Rpgrip1l–injected, 22 Mo-Pk1–injected, and 20 co-injected embryos, from a representative experiment out of 4 repeats. In C, four independent experiments are represented. n = 151 control, 178 Mo-Rpgrip1l–injected, 126 Mo-inv–injected and 134 co-injected embryos. In A–C, the differences in the repartition in classes between batches of embryos injected with one morpholino only and the batch of co-injected embryos are significant (\( \alpha < 0.001 \), Khi2 test). (D) Diagram illustrating the percentage of floor plate cells with a basal body in posterior (P), medial (M), or anterior (A) position in 18-s stage embryos, either uninjected or injected with a mixture of morpholinos targeting each of the 3 zebrafish dvl genes (Mo3Dvl; 0.2 mM each). The diagram corresponds to a total of 5 control embryos (252 cells analyzed) and 5 Mo3Dvl-injected embryos (204 cells analyzed), from a single representative experiment out of 3 repeats. The double asterisk means that the differences are statistically significant (Khi2 test: \( \alpha < 0.001 \)). This experiment shows that the posterior localization of basal bodies in floor plate cells depends on the concentration of functional Dvl proteins and suggests that the reduction in Dvl amounts in rpgrip1l morphants may be the cause of basal body localization defects. (E–J) Immunofluorescence with an anti-GFP antibody to reveal the GFP tagged Dsh protein and with an antiaetylated \( \alpha \)-tubulin antibody (Ac Tub) to label cilia in embryos injected with DshGFP RNA (10 pg/embryo) without morpholino (E) or with Mo-1 (F), Mo-inv (G and H), or both (I and J). In control embryos, DshGFP accumulated at the cilium base in notochord and floor plate cells (E). In embryos co-injected with Mo-Rpgrip1l and DshGFP, DshGFP was severely reduced at the cilium base (F). Mo-inv–injected embryos showed increased Dsh-GFP levels in the cytoplasm as compared with controls, whereas the basal body pool appeared unaltered (G and H). In double morphants, the pool of dishevelled at the cilium base was severely reduced or absent, and the cytoplasmic pool was reduced compared with inversin morphants, but higher than in controls (I and J). Insets in E–J show higher magnifications in the floor plate region. These experiments show that Rpgrip1l and inversin interact positively for PCP and suggest that each protein acts preferentially on a specific cellular pool of dishevelled: inversin mainly destabilizes the cytoplasmic pool, whereas Rpgrip1l mainly stabilizes the basal body pool. Bars: (E–J and insets) 5 \( \mu \)m. (K–R) Immunofluorescence on control (K–N) and rpgrip1l morphant (O–R) embryos at the 14-s stage co-injected with vangl2-GFP (25 pg) and mb-Cherry (22 pg) mRNAs. Lateral view of a few floor plate cells of flat-mounted embryos are shown. The epitopes detected by the antibodies are indicated on the top of the picture. In the merge picture, cilia (Ac Tub) are blue, membranes (mCherry) are red, and Vangl2 (GFP) is green. The Vangl2-GFP protein is present in the membrane of floor plate cells without any obvious anterior or posterior enhancement, and no obvious modification of this pattern is observed in rpgrip1l morphants. However, an asymmetric accumulation of Vangl2 could be missed in these overexpression experiments. Bar: (K–R) 10 \( \mu \)m.]

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Figure S4. Cilium and basal body integrity in MDCK cells upon RPGRIP1L depletion. (A) Quantitative RT-PCR and Western blot analysis of the canine RPGRIP1L gene and protein product expression levels in polyclonal MDCK cell lines depleted for RPGRIP1L (RPGRIP1L-KD-sh1/sh2) and control cells (pSICOR). (B) Immunostaining of acetylated α-tubulin (ac-α-tub) performed on pSICOR and RPGRIP1L-KD cells after PFA fixation. The diagrams present the percentage of ciliated cells and cillum length in RPGRIP1L-KD versus control cells (n = 1,500). (C) Pericentrin (red) staining after PFA/Pipes fixation was reduced in RPGRIP1L-depleted cells compared with control pSICOR cells, suggesting a partial loss of the pericentriolar material. (D) The percentage of cells exhibiting a γ-tubulin staining after PFA fixation was reduced in RPGRIP1L-KD-sh1/2 MDCK cell lines versus control pSICOR. While inversin or nephrocystin-3 were present in most of the γ-tubulin–positive RPGRIP1L-KD cells, Dvl2 or Dvl3 staining were lost in half of these cells (200–300 cells analyzed). (E) A specific affinity-purified rabbit antibody raised against the human nephrocystin-3 protein (aa 301–577; APN3) detected both the endogenous protein (113 kD) and the overexpressed truncated protein nephrocystin-3-Nter-Myc (aa 1–619; 72 kD) in HEK293T cells. No specific staining was observed with the preimmune serum (preIm). (F and G) Co-immunostaining of CEP290 (F) or nephrocystin-3 (G, red) with γ-tubulin (green) on RPGRIP1L-KD MDCK cells was not affected compared with control cells (pSICOR). Nuclei were stained with Hoechst. Bars, 10 μm.
Figure S5. **RPGRIP1L depletion in MDCK cells: rescue and dishevelled localization at the membrane.** (A–C) Levels of Dvl3 proteins at the basal body are rescued by reexpression of RPGRIP1L. (A) Co-staining of Dvl3 (red) with γ-tubulin (green) after PFA fixation in RPGRIP1L-KD-sh1 MDCK cell lines with (bottom) or without (top) stable expression of RPGRIP1L-Myc demonstrates a rescue of Dvl3 accumulation at the centrosome/basal body in RPGRIP1L-reexpressing cells. (B) Quantitative RT-PCR analysis showing RPGRIP1L-Myc expression. (C) Western blot analysis reveals that expression of RPGRIP1L-Myc in RPGRIP1L-KD cells restores global Dvl3 protein levels. (D) Presence of Dvl2 and Dvl3 at the membrane of MDCK cells. Co-staining of Dvl2 or Dvl3 (red) with γ-tubulin (green) after methanol fixation in RPGRIP1L-KD-sh1/2 MDCK cell lines and control pSICOR. Methanol fixation permits the detection of dishevelled at the cell membrane and in faint dots at the basal body, but not in the pericentriolar material. Dvl2 and Dvl3 staining is present at the membrane in both control and RPGRIP1L-depleted cells, despite the wavy appearance of the cell junctions. A weaker staining was noted only for Dvl3 in RPGRIP1L-KD-sh1 cells. Bars, 10 µm.

References


