The inner and outer compartments of mitochondria are sites of distinct cAMP/PKA signaling dynamics

Konstantinos Lefkimmiatis,1,2 Daniela Leronni,1,2 and Aldebaran M. Hofer1,2

1VA Boston Healthcare System and 2Department of Surgery, Brigham and Women’s Hospital, Harvard Medical School, West Roxbury, MA 02132

Cyclic AMP (cAMP)-dependent phosphorylation has been reported to exert biological effects in both the mitochondrial matrix and outer mitochondrial membrane (OMM). However, the kinetics, targets, and effectors of the cAMP cascade in these organelar domains remain largely undefined. Here we used sensitive FRET-based sensors to monitor cAMP and protein kinase A (PKA) activity in different mitochondrial compartments in real time. We found that cytosolic cAMP did not enter the matrix, except during mitochondrial permeability transition. Bicarbonate treatment (expected to activate matrix-bound soluble adenylyl cyclase) increased intramitochondrial cAMP, but along with membrane-permeant cAMP analogues, failed to induce measurable matrix PKA activity. In contrast, the OMM proved to be a domain of exceptionally persistent cAMP-dependent PKA activity. Although cAMP signaling events measured on the OMM mirrored those of the cytosol, PKA phosphorylation at the OMM endured longer as a consequence of diminished control by local phosphatases. Our findings demonstrate that mitochondria host segregated cAMP cascades with distinct functional and kinetic signatures.

Introduction

The classical second messenger, cAMP, has recently gained attention as a player in the control of mitochondrial function. Typically, cAMP relies on its main effector, protein kinase A (PKA). This enzyme can be free in the cytosol or confined to precise subcellular locations thanks to a family of proteins called A kinase anchoring proteins (AKAPs; Feliciello et al., 2005; Pagliarini and Dixon, 2006; O’Rourke et al., 2011). Specific AKAPs such as AKAP 121 and SPHKAP/SKIP are known to tether PKA to the outer mitochondrial membrane (OMM) and intermembrane space in proximity of local targets (Lieberman et al., 1988; Feliciello et al., 2005; Kovanich et al., 2010; Means et al., 2011). This spatial organization allows timely phosphorylation of mitochondrial proteins that regulate apoptosis (BAD; Harada et al., 1999), mitochondrial shape (Drp1; Cribbs and Strack, 2007; Chang and Blackstone, 2007), and cristae maintenance (ChChd3; Darshli et al., 2011; Means et al., 2011).

Recently, the existence of an independent intramitochondrial cAMP signaling circuit was also reported. This cascade consists of a matrix-localized cAMP source, the bicarbonate-activated soluble adenylyl cyclase (sAC; Wuttke et al., 2001; Zippin et al., 2003), PKA holoenzyme (Sardanelli et al., 2006; Acin-Perez et al., 2011a), and the cAMP-degrading enzyme PDE2A (Acin-Perez et al., 2011b). Matrix-confined PKA was proposed to phosphorylate cytochrome c oxidase (COXIV-I) to enhance oxidative phosphorylation (OXPHOS; Acin-Perez et al., 2011a). However, prior data suggesting that PKA activation inhibits OXPHOS (Bender and Kadenbach, 2000; Robin et al., 2003; Helling et al., 2008) appear to be in striking conflict with this model.

Although the presence of a complete cAMP signaling machinery in the matrix suggests that mitochondria might be able to use a local cAMP messenger system in situ, the dynamic nature of these putative cAMP signals and how they impact downstream effectors is largely unknown. Additionally, the extent to which cAMP produced in the cytoplasm reaches different mitochondrial

Supplemental Material can be found at:
/content/suppl/2013/07/25/jcb.201303159.DC1.html
Figure 1. Cyclic AMP produced in the cytosol does not reach the mitochondrial matrix. (A) Confocal images of HeLa cells expressing mito-EpacH90 and loaded with MitoTracker red suggesting proper localization of mito-EpacH90. (B) cAMP measurements in intact cells expressing cytosolic EpacH90 (black trace) or mito-EpacH90 (red trace; mean of five cells); typical of \( n = 9 \) experiments; 34 mito-H90, 19 Epac-H90 cells. (C) Digitonin-permeabilized...
compartments and the question of how PKA tethered to mitochondria responds to cAMP compared with the enzyme in the bulk cytoplasm have only been partially addressed (DiPilato et al., 2004; Allen and Zhang, 2006; Acin-Perez et al., 2009b; Agnes et al., 2010).

Here we targeted fluorescence resonance energy transfer (FRET)-based cAMP and PKA activity sensors to the OMM and matrix to visualize changes in mitochondrial cAMP and PKA activity in real time in live cells. Our data demonstrate that mitochondria harbor at least two distinct cAMP microdomains (matrix and OMM) with unique signaling characteristics that separate them functionally from the cytosolic cAMP pathways.

Results & discussion

Among the most important information-bearing molecules, cAMP is known to influence mitochondrial function; however, whether cAMP penetrates the mitochondrial matrix is less clear (DiPilato et al., 2004; Acin-Perez et al., 2009b). To address this issue, we targeted the pH-insensitive FRET- and Epac-based cAMP sensor EpacH90 (van der Krogt et al., 2008) to the mitochondrial matrix. This construct (“mito-EpacH90”) colocalized with the marker MitoTracker red, which is consistent with mitochondrial localization (Fig. 1 A).

HeLa cells expressing mito-EpacH90 or EpacH90 (cytosolic) were mixed and seeded onto glass coverslips so that cells harboring both types of sensor could be visualized in the same microscope field. Cells expressing EpacH90 responded to the cAMP-generating agonist forskolin (FSK) in the presence of the microscope field. Cells expressing EpacH90 responded to the targeted Ca2+ sensor mito-D3cpv (Filippin et al., 2005; Palmer cytosol did not reach the matrix (Fig. 1 B).

We next used mixed populations of digitonin-permeabilized HeLa cells expressing either mito-EpacH90 or the matrix-targeted Ca2+ sensor mito-D3cpv (Filippin et al., 2005; Palmer and Tsien, 2006) together with the marker mCherry (Shaner et al., 2004). Mitochondria displayed the expected Ca2+ uptake (Baughman et al., 2011; De Stefani et al., 2011), indicating successful permeabilization. Meanwhile nearby cells expressing mito-EpacH90 did not respond to exogenous Ca2+ or cAMP (10 µM) but only to the membrane-permeable cAMP analogue 8-bromo-adenosine 3'5'-cyclic monophosphate (8Br-cAMP; Fig. 1 C; see also Fig. S1 A).

Collectively, these data demonstrate that cAMP cannot permeate the inner mitochondrial membrane. This is in agreement with prior studies using indirect biochemical approaches in isolated mitochondria (Acin-Perez et al., 2009b), but contrasts previous cAMP FRET imaging measurements by DiPilato et al. (2004). This discrepancy most likely lies in the localization signal used for our cAMP sensor, which minimized the fraction of reporter mislocalized to the cytosol.

Next, we investigated whether conditions exist that permit cytosol and matrix to exchange information via cAMP. We reasoned that this communication might be most prominent during the mitochondrial permeability transition (MPT; Newmeyer and Ferguson-Miller, 2003; Hajnóczky et al., 2006). We induced MPT in digitonin-permeabilized HeLa cells using Ca2+ overload (Bopassa et al., 2005) while monitoring mitochondrial Ca2+ and cAMP contemporaneously. Upon MPT, exogenous cAMP rapidly entered the matrix (Fig. 1 D). Treatment with 10 µM Ca2+ in the presence of 10 µM cAMP did not alter the matrix cAMP content; however, addition of 100 µM Ca2+ induced massive cAMP entry after roughly 60 s (Fig. 1 E). Our findings suggest that during MPT, cAMP is free to diffuse into the matrix, intermingling these two otherwise independent cascades.

An intramitochondrial sAC has been proposed to generate cAMP in response to metabolically produced HCO3-/CO2 (Acin-Perez et al., 2009b). As shown in Fig. 1 F, mito-EpacH90 responded with an increase of the FRET ratio when we switched acutely from Hepes-buffered media to HCO3-/CO2-buffered solutions. These data are consistent with the idea that bicarbonate anion is capable of eliciting cAMP signals in the matrix by allowing sAC to independently generate cAMP (Acin-Perez et al., 2009b, 2011a).

We next asked whether cAMP produced via HCO3-/CO2 would stimulate the previously reported matrix-resident PKA (Sardanelli et al., 2006; Acin-Perez et al., 2009b; Agnes et al., 2010). PKA activity was monitored using matrix-targeted versions of the FRET-based sensors AKAR3 and AKAR4 (Allen and Zhang, 2006; Depry et al., 2011; “mito-AKAR3; mito-AKAR4”); Fig. 2 A). HCO3-/CO2 caused the predicted increase in matrix cAMP content; however, contrary to our expectations, there was no increase in PKA activity in neighboring cells expressing mito-AKAR4 (identified by the marker mCherry; Fig. 2 B).

As controls we generated sensors targeted to the OMM (“OMM-AKAR3” and “OMM-AKAR4”). As shown in Fig. 2 C, in mixed populations of HeLa cells expressing mito-AKAR3 or OMM-AKAR3, mito-AKAR3 (red traces) did not respond to cytosolic cAMP generated by FSK/IBMX, whereas OMM-AKAR3 in neighboring cells (black traces) did. This result was expected based on the finding that cytosolic cAMP does not.

Hela cells expressing mito-D3cpv (black traces) or mito-EpacH90 (red trace; n = 6 experiments; 10 mito-D3cpv cells; 13 mito-EpacH90 cells; red trace indicates the mean of three cells). [D] Mixed populations of permeabilized Hela cells expressing mito-D3cpv (black trace) or mito-EpacH90 (red traces; n = 3 experiments; 5 mito-D3cpv cells; 8 mito-EpacH90 cells). After Ca2+ pulses, mitochondria changed morphology and failed to retain Ca2+, (hallmarks of MPT), coincident with responses of mito-EpacH90 to exogenous cAMP. [E] Permeabilized Hela cells expressing mito-EpacH90 (n = 5 experiments; 20 cells) subjected to increasing [Ca2+] in the presence of cAMP (10 µM). 100 µM Ca2+ induced a dramatic increase in cAMP measured by mito-EpacH90. [F] Cells expressing mito-EpacH90 bathed in Hepes-buffered normal Ringer’s solution (continuous perfusion) then switched to CO2/HCO3-buffered Krebs-Ringer’s, inducing an apparent cAMP rise (presumably due to matrix sAC activation). Data from two representative cells are shown (n = 6 repeats; 16 mito-EpacH90 cells). Error bars indicate mean ± SD.
Figure 2. PKA activity in the mitochondrial matrix measured using targeted FRET-based reporters. (A) Confocal images of HeLa cells expressing mito-AKAR4 loaded with MitoTracker red. (B) Mixed populations of HeLa cells transfected with mito-EpacH90 (black trace; mean of three cells) or mito-AKAR4 (red trace; mean of four cells) were initially bathed in Hepes-buffered solution and then switched to CO$_2$/HCO$_3^-$-buffered solution ($n=5$ experiments; on June 28, 2017 Downloaded from Published July 29, 2013
reach the matrix. However, 8-(4-chloro-phenylthio)-adenosine-3',5'-cyclic monophosphate (8CPT-cAMP; a potent activator of PKA; see Fig. S1B) also failed to elicit PKA activity in the matrix as measured by mito-AKAR3 (Fig. 2 D; red traces; see also Fig. S1 C). 8CPT-cAMP rapidly reached the mitochondrial matrix (Fig. 2 E), but failed to elicit detectable PKA activity. In HEK cells, OMM-AKAR4 responded to cytosolic cAMP elevations induced by prostaglandin E2 (PGE2) and FSK/IBMX. Addition of 8CPT-cAMP did not elicit additional elevation of the ratio (likely because of saturation of the probe), whereas the PKA inhibitor N-[2-[p-Bromocinnamylamino]ethyl]-5-isouquinolinesulfonamide (H89) reversed the FRET signal. However, neighboring cells expressing mito-AKAR4 again showed no ratio changes during any of these treatments (Fig. 2 F).

Although the preceding findings would imply that the matrix does not contain an endogenous PKA holoenzyme capable of responding to intramitochondrial cAMP signals, we also considered the trivial possibility that the AKAR sensors simply do not perform properly in the milieu of the matrix. As shown in Fig. S1 D, reduction of the bulky targeting signal (4Cox8) on the AKAR reporter did not alter our results, nor did we see FRET changes after delivery of exogenous cAMP to the matrix during MPT (Fig. S1 E). We also confirmed that the AKAR sensors were functional in the relatively alkaline environment of the matrix (Fig. S1 F).

As a further control, we prepared matrix-targeted versions of mCherry-tagged constitutively active PKA “mito-PKACat-mCherry” (based on the catalytic subunit α; Day et al., 2011) and the potent protein kinase inhibitor PKI (isoform α), also tagged with mCherry (“mito-PKI-mCherry”). The mitochondrial localization of these constructs was ascertained by confocal microscopy (Fig. S2, A and B).

As detailed in Fig. S2 (C and D), the cytosolic parent constructs were extremely effective in activating (cyto-PKACat-mCherry) or inhibiting (cyto-PKI-mCherry) PKA. In addition, the fact that cyto-PKACat-mCherry did not affect the FRET signal of mito-AKAR4 and that PKI targeted to the matrix did not influence cytosolic or OMM PKA activity (Fig. S2, E–H) attests to the fidelity of our targeting strategy and demonstrates that we can independently manipulate PKA activity in these different compartments.

As summarized in Fig. 3 A, mito-AKAR4 consistently reported significantly higher starting FRET ratios when co-expressed with mito-PKACat-mCherry compared with neighboring controls. This effect was completely reversed by mito-PKI-mCherry, and remained unaltered in the presence of cytosolic PKI. Importantly, matrix-targeted PKI had no effect on the initial mito-AKAR4 ratio, which suggests a lack of basal phosphorylation of the matrix-targeted sensor under resting conditions.

Collectively, these data indicate that mito-AKAR4 is able to report PKA activity, but the endogenous PKA is either not

![Figure 3](https://example.com/fig3.png)

**Figure 3.** Overexpression of PKA in the matrix is detected by AKAR sensors, and induces specific phosphorylation patterns. (A) Summary of the starting mito-AKAR4 ratios in the presence of mito-PKACat-mCherry, mito-PKI-mCherry, and cyto-PKI-mCherry (***, P < 0.0002 with respect to control). Error bars indicate mean ± SD. (B) Phosphorylation status of cytosolic and mitochondria-enriched fractions of HEK cells treated with cAMP-generating agonists or cell-permeant cAMP analogues, or transfected with mito-PKACat-mCherry. A phospho-(Ser/Thr) PKA substrate antibody unveiled mito-PKACat-mCherry–dependent mitochondria-specific phosphorylation bands (arrows). The purity of mitochondria was tested using an antibody cocktail against the human OXPHOS subunits, whereas GAPDH was used to detect any cytosolic contamination (typical of n = 3 independent experiments).
mito-GO-ATeam2 (Fig. S3D). In a recent paper, Lu et al. (2013) found that the mitochondrial transcription factor TFAM can be phosphorylated by overexpression of catalytically active PKA in the matrix of HEK cells (Lu et al., 2013). However, we interpret our data above and those of Lu et al. (2013) cautiously; PKA is a very strong kinase, and its expression in restricted domains such as the matrix might result in promiscuous phosphorylation of proteins with less than optimal PKA consensus sites (Ubersax and Ferrell, 2007).

During the course of our studies, we consistently observed that OMM-AKAR4 responses were very robust compared with the cytosol, which was also noted by Allen and Zhang (2006) using AKAR3 targeted to the OMM via a different targeting strategy. As shown in Fig. 4A, in HEK cells challenged with the cAMP-generating agonist isoproterenol (ISO; 10 nM), both OMM-AKAR4 and AKAR4 responded strongly. However, upon ISO removal, OMM-AKAR4 exhibited strikingly slower termination kinetics. In HEK expressing either OMM-EpacH90 or EpacH90, all cells responded similarly to ISO, suggesting that cAMP rises and is degraded similarly in cytosol and OMM.

Interestingly, when we expressed mito-PKACat-mCherry and examined PKA-dependent phosphorylation patterns by Western blotting, at least two mitochondrial-specific bands were observed (Fig. 3 B). Meanwhile, activation of endogenous PKA through FSK (which generates extra-mitochondrial cAMP only) or 8CPT-cAMP (which potentially influences both the matrix and cytosol) did not yield this same mitochondrial-specific phosphorylation pattern. It is perhaps noteworthy that our measurements using a matrix-localized ATP sensor, mito-GO-ATeam2 (Nakano et al., 2011), were not consistent with an effect of matrix-specific cAMP signals on ATP production (Fig. S3, A–C). Meanwhile, expression of mito-PKACat-mCherry did alter the resting mitochondrial ATP content as measured by mito-GO-ATeam2 (Fig. S3 D). In a recent paper, Lu et al. (2013) found that the mitochondrial transcription factor TFAM can be phosphorylated by overexpression of catalytically active PKA in the matrix of HEK cells (Lu et al., 2013). However, we interpret our data above and those of Lu et al. (2013) cautiously; PKA is a very strong kinase, and its expression in restricted domains such as the matrix might result in promiscuous phosphorylation of proteins with less than optimal PKA consensus sites (Ubersax and Ferrell, 2007).

During the course of our studies, we consistently observed that OMM-AKAR4 responses were very robust compared with the cytosol, which was also noted by Allen and Zhang (2006) using AKAR3 targeted to the OMM via a different targeting strategy. As shown in Fig. 4A, in HEK cells challenged with the cAMP-generating agonist isoproterenol (ISO; 10 nM), both OMM-AKAR4 and AKAR4 responded strongly. However, upon ISO removal, OMM-AKAR4 exhibited strikingly slower termination kinetics. In HEK expressing either OMM-EpacH90 or EpacH90, all cells responded similarly to ISO, suggesting that cAMP rises and is degraded similarly in cytosol and OMM.
PDEs are major players in the generation of cAMP microdomains (Zaccolo, 2011). However, their role in the generation of persistent PKA activity at the OMM appeared to be limited. We therefore considered the role of local Ser/Thr protein phosphatases, known constituents of macromolecular complexes coordinated by AKAPs (Feliciello et al., 2005; Carlucci et al., 2008), in this process. As shown in Fig. 5 A, the potent Ser/Thr phosphatase inhibitor calyculin A attenuated the H89-induced...
recovery of both AKAR4 and OMM-AKAR4 in HeLa cells stimulated with FSK/IBMX. These data suggest that the termini of PKA activity at the OMM depends largely on calyculin A-sensitive phosphatases.

In HEK cells challenged with 10 nM ISO, the onset of PKA activation was similar in cytosol and OMM (Fig. 5 B, right). However, the termination of the response after PKA inhibition by H89, which reflects the local phosphatase activity, was again significantly slower at the OMM (P < 0.0002; Fig. 5 B, right). Finally AKAR4 and OMM-AKAR4 responded differently to H89 in cells expressing the catalytically active PKA-Cat-mCherry. As expected, both sensors displayed a much higher starting ratio than that of AKAR4 (Fig. 5 C, right), which indicates that these differences did not depend entirely on local PKA activity.

Collectively, these data indicate that the OMM is a privileged cAMP signaling microdomain with high PKA activity. This microenvironment promotes cAMP-triggered activity not only by concentrating PKA in situ via specific AKAPs (Feliciello et al., 2005), but also, as demonstrated by our data, by hosting a less active Ser/Thr phosphatase population. We propose that this pro-PKA OMM maintains the phosphorylation state of local targets such Drp1, BAD, and Ca2+-release channels at ER–OMM junctions (Rizzuto et al., 1998; Harada et al., 1999; Cribs and Strack, 2007; Chang and Blackstone, 2007; Csordás et al., 2010; Merrill et al., 2011) even in the face of waning cytosolic PKA activity.

In this work we found that the mitochondrial matrix of HEK and HeLa cells contains low or null PKA activity. However, this raises the question of how mitochondrial proteins that contain PKA consensus can be phosphorylated in the matrix (Covian and Balaban, 2012). Our finding that the OMM is rich in PKA activity together with the observation that the intermembrane space contains a functional AKAP (Means et al., 2011) raises the possibility that PKA targets might be phosphorylated during their transition into the matrix.

Materials and methods

Reagents
8Br-cAMP, 8CPT-cAMP, 8CPT-OME-cAMP, and N6-Benzoyladenosine-3’, 5’-cyclic monophosphate (6Bnz-cAMP-AM) were purchased from Biolog Inc. and EMD Millipore. H89 was purchased from Enzo Life Sciences. All restriction enzymes were obtained from New England Biolabs, Inc. Custom-made primers and MitoTracker red/green were purchased from Invitrogen. Unless otherwise noted, all other reagents were obtained from Sigma-Aldrich.

Generation of mitochondrial-targeted FRET-based sensors

The FRET-based CAMP sensor EpacH90 was a generous gift of Kees Jalink (Netherlands Cancer Institute, Amsterdam, Netherlands). AKAR3 and AKAR4 were provided by Jin Zhang (John Hopkins University School of Medicine, Baltimore, MD). Hiromi Imamura (The Hakubi Project, Kyoto University, Tokyo, Japan) provided us with the mitochondrial ATP sensors, mito-GO-Ateam2/3. The matrix targeting signal 4Cox8 was extracted from mito-D3cpv (a gift of Giulietta Di Benedetto and Tullio Pozzan, University of Padova, and Venetian Institute of Molecular Medicine, Padua, Italy) using HindIII and introduced to the N terminus of EpacH90 (pcDNA3). AKAR3, and AKAR4 (pcDNA3). The OMM targeting peptide yTom70 (derived from yOMmCherry, provided by Gyorgy Hajnoczky, Thomas Jefferson University, Philadelphia, PA) was introduced into these plasmids using custom-made primers encoding this fragment and pre-designed restriction enzyme sites for HindIII. The protein kinase A inhibitor isoform alpha (purchased from Origene Technologies, Rockville, MD) was amplified with primers bearing BamHI sites and subcloned in-frame with mCherry in pcDNA3. PKAcatal-mCherry (PKA catalytic subunit alpha from mouse) was a gift from Susan S. Taylor (University of California at San Diego, La Jolla, CA) and Roger Y. Tsien (University of California at San Diego and Howard Hughes Medical Institute, La Jolla, CA). Matrix targeting for PK1-mCherry and PKA-Cat-mCherry was achieved using the 4cox8 sequence. All constructs were tested functionally and verified by sequencing (Dana Farber DNA Resource Core, Boston MA).

Cell culture and transfection

HEK and HEK293 cells were obtained from the American Type Culture Collection and were grown in DMEM + 10% FBS. Both cell lines were maintained in a humidified CO2 (5%) incubator at 37°C and were split every 2–3 d after reaching 80% confluence. All plasmids were transfected using Effectene transfection reagent (Qiagen).

Confocal imaging

HEK cells were seeded onto glass coverslips and transfected with sensors the next day. 24 h after transfection, the cells were loaded with 10 nM MitoTracker red or 25 nM MitoTracker green for 15 min at 37°C. Subsequently, cells were rinsed four times with Hepes-buffered Ringer’s solution containing (in mM): 125 NaCl, 25 Hepes, 10 glucose, 5 K2HPO4, 1 MgSO4, and 1 CaCl2, pH 7.40. Coverslips were mounted onto a home-built flow-through perfusion chamber, and cells were bathed in Hepes-buffered Ringer’s solution at room temperature and imaged under a 60× Plan-Apochromat (NA 1.40) oil immersion objective lens using a confocal microscope (Eclipse C1si; Nikon). Images were collected on live cells with Kalman filtering (n = 6) using the EZ-C1 software (Nikon).

Cell permeabilization

Cells were briefly rinsed with an intracellular-like buffer (125 mM KCl, 25 mM NaCl, 10 mM Hepes, 0.5 mM MgCl2, 1 mM ATP, 500 μM EGTA, and 211 μM CaCl2, pH 7.25) complemented with 5 mM succinate and 5 mM glutamate. Consequently, cells were treated with intracellular buffer containing digitonin (5–10 μg/ml). Cellular permeabilization was determined on stage by visual evaluation of the plasma membrane morphology under bright-field illumination.

Western blotting

HEK cells were transfected with mito-PKAC (a) mCherry or left untransfected. 24 h later, proteins were extracted from the transfected populations, while the nontransfected cells were treated with cAMP analogues (8CPT-cAMP, 8Br-cAMP, or 6Bnz-cAMP) or TSK in the presence or absence of the PKA inhibitor H89 for 1 h at 37°C. Subsequently, cells were lysed using RIPA buffer (Sigma-Aldrich) complemented with a protease and phosphatase inhibitor cocktail (Thermo Fisher Scientific). Cell lysates were sonicated and insoluble material was removed by centrifugation at 14,000 g for 10 min at 4°C. Mitochondria were extracted using a commercial mitochondria isolation kit (Thermo Fisher Scientific) according to the manufacturer’s instructions. Total cell lysates or mitochondria-enriched fractions were resolved on 5–20% Tris-glycine SDS-PAGE gels (Wako Chemicals, Inc.) and electroblotted onto polyvinylidene fluoride (PVDF) membranes (Hybond-P, GE Healthcare). After transfer, PVDF membranes were blocked for 3 h at room temperature in 5% BSA–Tris-buffered saline/ Tween 20 (TBS; 10 mM Tris HCl, pH 8.0/0.150 mM NaCl/0.1% Tween 20). The membranes were incubated overnight at 4°C with a phospho-(Ser/Thr) PKA substrate antibody diluted 1:700 in 5% BSA-TBST. The day after, the membranes were washed four times with TBS and incubated at room temperature for 1 h with a peroxidase-conjugated secondary antibody (1:2,000; Santa Cruz Biotechnology, Inc.). Peroxidase activity was detected with enhanced chemiluminescence (ECL Advance Western blotting detection kit, GE Healthcare). GAPDH antibody (1:4,000; Santa Cruz Biotechnology, Inc.) was used as a loading control for the total cell lysates and to detect cytosolic protein contamination in the mitochondrial enriched samples, whereas a total OXPHOS Human WB antibody cocktail against complexes I–V (1:700; Abcam) was used to determine mitochondrial enrichment.

Ratio imaging

Cells were mounted onto a home-built perfusion chamber and bathed in Hepes-buffered Ringer’s solution containing (in mM): 125 NaCl, 25 Hepes,
Krebs-Henseleit solution was gassed continuously with 95% O2/5% CO2.

ments of sAC activation, we acutely switched the superfusion from Hepes

Acin-Perez, R., D.L. Gatti, Y. Bai, and G. Manfredi. 2011a. Protein phosphory-


Bender, E., and B. Kadenbach. 2000. The allosteric ATP-inhibition of cyto-

chrome c oxidase activity is reversibly switched on by cAMP-dependent phosphorylation. FERS. Lett. 466:130–134. http://dx.doi.org/10.1016/S0001-5793(99)01773-1


Carlucci, A., L. Lignito, and A. Feliciello. 2008. Control of mitochondria dy-


Cribbs, J.T., and S. Strack. 2007. Reversible phosphorylation of Drp1 by cy-

clic AMP-dependent protein kinase and calcium regulates mitochondrial fission and cell death. EMBO Rep. 8:939–944. http://dx.doi.org/10.1038/embor.7410862


