Dyskerin, tRNA genes, and condensin tether pericentric chromatin to the spindle axis in mitosis

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Abbreviations used in this paper: IP, immunoprecipitation; LacO, lactose operon; LTR, long terminal repeat; rDNA, ribosomal DNA; snoRNP, small nuclear RNP; TetO, tetracycline operon; WT, wild type.

Introduction

Condensin is a DNA compaction machine that functions in chromosome architecture, nucleolar organization, and chromosome segregation (Hirano, 2006). Condensin can introduce supercoils and decatenate topologically linked circles. The most prominent sites of condensin localization in the nucleus are the nucleolus, pericentric chromatin, and central axis of condensed metaphase chromosomes. Condensin is also essential for mechanisms of force balance between the pericentromere heterochromatin and spindle microtubules in metaphase (Stephens et al., 2011, 2013a). Condensin interacts with several key transcription factors, including RNA polymerase III transcription factors (TFIIFC and TFIIB; Haeseler et al., 2008). This interaction is responsible for the enrichment of tRNA genes to the nucleolar periphery observed in budding yeast. Recently, it has been shown that the monopolin complex recruits condensin to the pericentromere (Brito et al., 2010; Burrack et al., 2013). The monopolin complex is sequestered in the nucleolus until the onset of anaphase, in which it migrates to the kinetochore (Brito et al., 2010; Burrack et al., 2013). In meiosis, monopolin is thought to cross-link sister kinetochores to ensure that sister chromatids segregate to the same pole. None of the known mechanisms attributed to condensin indicate how it functions in force balance in metaphase.

Budding yeast lacks canonical pericentric heterochromatin observed in most organisms. Centromeres in Saccharomyces cerevisiae are specified in a site-specific manner (point centromere) but share several features of the surrounding pericentric chromatin characteristic of that found in multicellular organisms. Condensin and cohesin are enriched 3× in the 30–50-kb region surrounding the point centromere (Blat and Kleckner, 1999; Megee et al., 1999; D’Ambrosio et al., 2008). tRNA genes, found in the pericentric regions of many organisms (Kuhn et al., 1991; Iwasaki and Noma, 2012), are enriched 1.8× in the pericentromere of budding yeast (32/307 tRNA genes in the 50 kb surrounding the CEN [centromere] sequence in the 16 chromosomes). tRNAs are modified by several factors, including dyskerin. Dyskerin binds to and stabilizes small noncoding RNAs, which together with other components (H/ACA small nuclear RNP [snoRNP]) catalyzes the conversion of uridine to pseudouridine in nascent ribosomal RNA and tRNA (Hoang and Ferré-D’Amaré, 2001). Dyskerin is a component of telomerase and is required for telomere maintenance in humans (Gu et al., 2009; Gardano et al., 2012). Dyskerin has been localized to the
Cbf5 has a C-terminal repeat 10x (KKE/DX) motif, conserved in several microtubule binding proteins (MAP1A and MAP1B). In this report, we demonstrate that Cbf5, together with tRNA genes within the pericentromere, is responsible for condensin accumulation along the spindle axis. This provides a mechanism for the physical segregation of condensin from cohesin in single cells and elucidates the role of condensin in mitotic force balance.

Figure 1. Pericentric condensin enrichment in WT, monopolin, cbf5, and tfc3 mutant cells. Condensin (Smc4-GFP) and spindle pole bodies (Spc29-RFP) were imaged at 24°C or after shift to 37°C for 6 h. Representative cells are shown in the top images. Arrows indicate the rDNA locus. (bottom) Integrated intensity of Smc4-GFP between spindle pole bodies (Spc29-RFP) in metaphase cells at 24°C in WT (n = 36), lrs4Δ (n = 33), and cbf5-AUU (n = 39) cells. WT (n = 32), cbf5-1 (n = 21), and tfc3-tsv115 (n = 55) after 6 h at 37°C are shown. *, P < 0.05, t test; error bars represent standard deviation. Bar, 1 µm.

mitotic spindle in human cells and shown to be required for faithful chromosome segregation (Alawi and Lin, 2013).

The yeast homologue of dyskerin, CBF5 was discovered in a biochemical assay for centromere binding factors (Jiang et al., 1993). Overexpression of CBF5 can partially suppress a temperature-sensitive mutation in cbf2/ndc10 encoding the 110-kD subunit of the Cbf3 kinetochore complex (Jiang et al., 1993). Furthermore, Cbf5 has a C-terminal repeat 10x (KKE/DX) motif, conserved in several microtubule binding proteins (MAP1A and MAP1B). In this report, we demonstrate that Cbf5, together with tRNA genes within the pericentromere, is responsible for condensin accumulation along the spindle axis. This provides a mechanism for the physical segregation of condensin from cohesin in single cells and elucidates the role of condensin in mitotic force balance.
Results and discussion

Enrichment of condensin along the metaphase spindle axis requires both monopolin and dyskerin

In metaphase, condensin (Smc4-GFP) is localized to both ribosomal DNA (rDNA; nucleolus) and the pericentric chromatin surrounding the spindle microtubules in mitosis. The rDNA is organized as a chromatin loop, often juxtaposed to the nuclear envelope (Fig. 1, arrows). Pericentric condensin appears as a focus or line of fluorescence between the spindle poles. In the absence of the major subunit of monopolin (lrs4Δ), there is a 20% reduction in the level of Smc4 associated with the pericentric chromatin. The remaining condensin appears heterogeneous along the spindle axis or radially dispersed (Fig. 1, lrs4Δ left bottom images).

Because tRNA genes are enriched in the pericentromere in budding yeast, we asked whether regulators of tRNA expression or modification enzymes, such as Cbf5, might recruit condensin to the metaphase spindle. Depletion of CBF5 results in a 50% reduction of Smc4 associated with the pericentric chromatin (Fig. 1, cbf5-1 top right images). In contrast to lrs4Δ mutants, condensin is not depleted from the rDNA in cbf5-1 (Fig. S1 C). To separate the essential tRNA modification function from Cbf5’s role in condensin localization, we used a nonessential mutation in the first AUG codon to AUU previously identified to alleviate repression by tRNA genes (art1-1; Kendall et al., 2000). This mutant affects Cbf5 expression levels and disrupts the nucleolar localization of tRNA genes (Kendall et al., 2000).

The concentration of condensin along the spindle axis in metaphase was reduced 60% in the cbf5-AUU mutant (Fig. 1, bottom). The localization of condensin in the nucleolus was unperturbed (Fig. S1, C and D). Because Cbf5 is part of the H/ACA box snoRNP complex (Nop10, Nhp2, and Gar1), we asked whether their depletion would affect spindle condensin. We placed Nop10 and Nhp2 under the control of the GALL promoter (Mumberg et al., 1994) and quantitated Smc4-GFP intensity along the spindle axis (Fig. S1). Smc4-GFP is reduced to ~50% of the levels after repression of either Nop10 or Nhp2 (Fig. S1 A). Thus, Cbf5 and the H/ACA box snoRNP complex are responsible for the majority (60%) of condensin recruitment to the mitotic spindle in metaphase.

To test whether RNA polymerase III transcription factors are involved in condensin recruitment, we used a mutation in a polymerase III transcription initiation factor (TFIIC), TFC3. There was a 60% reduction in Smc4-GFP associated with the spindle in a tfc3-tsv115 mutant (Fig. 1, bottom right bar). As with cbf5-AUU, the localization of condensin in the nucleolus was unperturbed in tfc3-tsv115 (Fig. S1 C). Loss of TFIIC leads to the reduction of condensin binding to these sites and therefore depletion in the pericentromere.

The reduction of condensin within the pericentromere in monopolin and dyskerin mutants results in a slight increase in spindle length (~1.8 µm) relative to wild type (WT; ~1.5 µm) but not as dramatic as the complete loss of condensin in brn1-9 mutants (2.3 µm; Table 1). To differentiate the role of monopolin and CBF5 in condensin localization within the pericentromere, line scans were drawn between the spindle poles in cells expressing Smc4-GFP in WT, lrs4Δ, or cbf5-AUU mutants (Fig. 2). The peak intensity of each line scan was binned into two zones, representing pole proximal (Fig. 2 B, green) and the central spindle axis (Fig. 2 B, blue). In the absence of monopolin, residual condensin localizes to the midspindle 83% of the time compared with 78% in WT (Fig. 2 B). In the cbf5-AUU mutant, condensin midspindle localization decreased, and pole-proximal localization increased significantly (Fig. 2 B). Representative images indicate the central spindle axis localization in monopolin mutants and a single focus that is proximal to the pole in cbf5-AUU (Fig. 2 A). Monopolin contributes disproportionally to the fraction of condensin proximal to the pole (only 50% central zone localization in cbf5-AUU mutant), whereas Cbf5 is responsible for condensin’s distribution along the spindle axis in metaphase (83% central axis in lrs4Δ mutant).

In live cells, Cbf5 is concentrated in the nucleolus (Fig. S2 A). To test whether Cbf5 is involved in condensin recruitment to the pericentromere, we performed chromatin immunoprecipitation (IP) in cells containing Cbf5-GFP (Fig. 2 C). There is a 2.4-fold enrichment of Cbf5 at the pericentromere (CEN3) versus an arm locus (HTB1; Fig. 2 C, left). Likewise, there is a significant loss of condensin from the pericentromere in the cbf5-1 mutation (Fig. 2 C, right). Thus, Cbf5 physically associates with pericentric chromatin and is responsible for aggregation of condensin to the spindle axis.

Proper distribution of condensin is required for kinetochore clustering in metaphase

Kinetochore from each of the 16 chromosomes are clustered into sister foci in a condensin-dependent fashion (Stephens et al., 2011, 2013a). Reducing the concentration of condensin in monopolin and dyskerin mutants might have a similar effect on kinetochore clustering. In ~47% of mutant cells, the outer kinetochore protein NuF2-GFP was no longer clustered into a close to diffraction focus (Fig. 3, A and C). Likewise, in greater than half of cells, the inner kinetochore protein Cse4-GFP was no longer clustered along the spindle axis (Fig. 3, B and D). The kinetochore were either declustered (as defined by multiple peaks of kinetochore proteins in line scans through the spindle}

Table 1. Spindle length increases in monopolin and cbf5 mutants, and spindle height increases in the cbf5 mutant

<table>
<thead>
<tr>
<th>Strain</th>
<th>Mean ± Standard Deviation</th>
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<tbody>
<tr>
<td>Spindle length (µm)</td>
<td></td>
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</tr>
<tr>
<td>WT</td>
<td>1.4 ± 0.21</td>
<td>47</td>
</tr>
<tr>
<td>lrs4Δ</td>
<td>1.8 ± 0.52</td>
<td>81</td>
</tr>
<tr>
<td>cbf5-1 3 h TS</td>
<td>1.9 ± 0.41</td>
<td>121</td>
</tr>
<tr>
<td>cbf5-1 6 h TS</td>
<td>2.4 ± 0.55</td>
<td>84</td>
</tr>
<tr>
<td>brn1-9</td>
<td>2.3 ± 0.40</td>
<td>80</td>
</tr>
<tr>
<td>Spindle height (nm)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>WT</td>
<td>517 ± 39</td>
<td>103</td>
</tr>
<tr>
<td>cbf5-1 6 h TS</td>
<td>583 ± 77</td>
<td>63</td>
</tr>
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were 10% wider in the cbf5-1 mutant (583 vs. 517 nm WT; Table 1). We estimate the increase in radius to be 50 nm in cbf5-1 versus WT. The kinetochore is radially displaced as a result of a structural change of the pericentric chromatin, rather than direct alteration of the kinetochore. Thus, condensin recruitment through monopolin and dyskerin is integral to kinetochore structure (Fig. 3) and spindle length regulation (Table 1).

Condensin is required for sister chromatin biorientation along the spindle axis
To test whether condensin functions in sister chromatin biorientation, we examined the geometry of centromere-linked lactose operon (LacO) spots in metaphase. The LacO arrays generally appear as a single focus (Fig. 4 B, WT) but separate into two foci in longer spindles (Fig. 4 B; Pearson et al., 2001). Sister LacO arrays biorient along the spindle axis in both WT and cells depleted of pericentric cohesin (>98%; Fig. 4 A). In the
Pericentric chromatin motion is perturbed in cbf5-AUU mutants

Pericentromeres of different chromosomes move coordinately in a condensin-dependent fashion (Stephens et al., 2013c). To determine whether dyskerin or monopolin was necessary for coordinated behavior, we imaged LacO and tetracycline operator (TetO) arrays linked to CEN15 and CEN11 (Fig. 4 D). These arrays were introduced into condensin (brn1-9), cohesin (mcm21Δ), dyskerin (cbf5-AUU), and monopolin (irs4Δ) mutants in the absence of condensin (brn1-9), 27% of the LacO spots become misaligned and appear perpendicular to the spindle axis. Similarly, sister LacO arrays orient perpendicular to the spindle axis in 31% of cases in cbf5-AUU mutants and 23% of cases in monopolin mutants (Fig. 4 C). The separation between sister foci does not necessitate biorientation between kinetochore microtubules from opposite spindle poles. Recruitment to the microtubule spindle axis is dependent on the axial distribution of condensin, and Cbf5 plays a dominant role in this function.
Like condensin, pericentric Tfc1 is heterogeneous and appears along the spindle or slightly displaced in $\geq 50\%$ of cells (Fig. 5 A, images). To quantitate the distribution, we measured the width of a line scan through the axis perpendicular to the spindle. In WT cells, the peak-to-peak distance is 470 nm. There is a 20% increase in the peak-to-peak distance of the Tfc1 bilobed structure in the $cbf5-1$ mutant (604 nm), indicative of radial expansion of Tfc1 localization. Thus, Cbf5 is necessary for maintenance of TFIIIC proximal to the spindle axis.

CBF5 could cluster condensin around the spindle axis in one of two ways. First, Cbf5 is reported to bind microtubules, which would present a viable mechanism for the axial positioning of condensin. Second, condensin could cross-link tRNA genes from different chromosomes. Based on the radial geometry of pericentric chromatin, cross-linking would gather strands (Fig. 4 E). The movements in the same half-spindle were compared using cross-correlation analysis (Stephens et al., 2013c). We find that correlated motion is decreased in the $cbf5-AUU$ mutant to the same extent as found in $brn1-9$ mutants (Fig. 4 E). In contrast, there is no significant reduction in correlated motion in $lrs4$ mutants in which condensin remains along the spindle axis (Fig. S2 C). Thus, Cbf5-dependent condensin along the spindle axis is required for the behavior and coordination of pericentric chromatin motion in metaphase.

**Role of pericentric tRNA genes in pericentric organization**

To determine how pericentric tRNA genes promote coordinated movement between different pericentromeres, we analyzed Tfc1-GFP. In WT cells, Tfc1 localizes to the pericentromere between the poles in metaphase (Fig. 5 A). Like condensin, pericentric Tfc1 is heterogeneous and appears along the spindle or slightly displaced in $\sim 50\%$ of cells (Fig. 5 A, images). To quantify the distribution, we measured the width of a line scan through the axis perpendicular to the spindle. In WT cells, the peak-to-peak distance is 470 nm. There is a 20% increase in the peak-to-peak distance of the Tfc1 bilobed structure in the $cbf5-1$ mutant (604 nm), indicative of radial expansion of Tfc1 localization. Thus, Cbf5 is necessary for maintenance of TFIIIC proximal to the spindle axis.

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the LacO containing chromosome III, the diameter of LacO distribution expands to 271 ± 195 nm with a greater mean distance from the pole of 372 ± 234 nm (Table 2).

Cbf5 is enriched 1.5–3.5-fold across seven tDNAs on chromosome III as determined by chromatin IP–quantitative PCR (Fig. S3). Thus, tDNA contributes to pericentric chromatin organization by acting as a binding site for condensin, TFIIIC, and Cbf5. Together, the tRNA gene machinery promotes condensin localization to the spindle axis, therefore contributing to spindle-proximal constraint of pericentric chromatin.

Convergence of point and regional centromeres

The budding yeast centromere shares features with regional centromeres found in fission yeast Schizosaccharomyces pombe and perhaps multicellular eukaryotes. The major repeated DNA sequences in yeast are the rDNA, subtelomere repeats, long terminal repeats (LTRs; 300-400 bp bracketing retrotransposons, 429 total), and tDNA genes (307 total). LTRs and tDNA genes are enriched 1.8× in the 50-kb region surrounding the centromere, relative to the remainder of the genome. tRNA genes, as well as repeat elements, are enriched in fission yeast centromeres. In from different chromosomes to a central position. To distinguish these hypotheses, we deleted the C-terminal microtubule binding domain of Cbf5. Condensin intensity and distribution along the pericentric region was indistinguishable from WT (Fig. S1 B). If the transcription and modification enzymes function at the centromere–proximal tRNA genes, removal of the tRNA genes from one chromosome should lead to displacement of the pericentromere from the spindle axis. A strain was constructed in which all tDNA genes were removed from chromosome III with essential genes incorporated elsewhere in the genome.

We analyzed the location of LacI-GFP bound to LacO in WT and the tDNA− strain. The LacO-GFP signal from individual cells is the same in WT and the tDNA− strain. Color-coded heat maps were created to represent the probability of LacO position relative to the spindle pole body (Fig. 5 B). The decrease in intensity reflects the change in distribution of LacO in WT versus tDNAΔ strains. The hot spots are reduced in the tDNA strain, indicating a reduced probability of localization of LacO. Furthermore, there is an increased radial displacement in tDNAΔ, indicative of increased randomness. In WT cells, the LacO position has a diameter of 240 ± 166 nm and a mean distance of 332 ± 200 nm from the spindle pole. Without tDNA on the LacO containing chromosome III, the diameter of LacO distribution expands to 271 ± 195 nm with a greater mean distance from the pole of 372 ± 234 nm (Table 2).

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Chromomatos pallidivittatus, a SINE (short interspersed repeat) element, Cp1, is enriched in the centromeres (Rovira and Edström, 1996; Liao et al., 1998). Cp1 has a polymerase III box, typical of tRNA genes, and is found in the range of 5–10 copies per centromere (Liao et al., 1998). Repeat sequences and tRNA genes have been proposed to prevent heterochromatin spreading into the centromere (Donze et al., 1999; Scott et al., 2007; Raab et al., 2012) as well as contributing to heterochromatin clustering as loading sites for structural maintenance of chromosome proteins (Kirkland and Kamakaka, 2013). In this study, we identify a novel function for tRNA genes that form the basis for condensin’s role in the chromatin spring.

The mechanism for restricting condensin to the spindle axis and repulsion of cohesin from the spindle axis had not been known. The enrichment of tRNA genes in the pericentric region led us to examine the role of tRNA transcription and modification factors as potential regulators. The transcription factor Tfc3 and modification enzyme, Cbf5 recruit condensin to the pericentromere and are responsible for the axial localization. Binding of tDNA transcription factor brings condensin proximal to the coding tDNA, which through aggregation of multiple tDNAs, results in the gathering of pericentromeres of different chromosomes. This will naturally lead to the centration of condensin along the spindle axis (Fig. 5 C). The removal of tDNA from chromosome III results in “puffing” of the pericentromere and an increase in the amount of space it can occupy (Fig. 5, B and C). The loss of tDNA results in the inability to secure the pericentromere of this chromosome to its neighbors, resulting in its expanded distance from the spindle axis.

The human homologue of Cbf5, dyskerin, functions in chromosome segregation fidelity in several organisms. Depletion of dyskerin results in mitotic defects, including delayed cell cycle progression, an increase in lagging chromosomes in anaphase, and activation of the spindle assembly checkpoint (Alawi and Lin, 2013). Upon depletion of cbf5-1 in budding yeast, we witness perturbation of kinetochore clustering and an increase in spindle length. We attribute these defects to a disruption of the pericentromere caused by loss of condensin along the spindle axis. A characteristic of mammalian cells depleted of condensin is the loss of ability of the kinetochore to sense tension (Ribeiro et al., 2009), metaphase arrest, and missegregation of chromosomes when the spindle checkpoint is inactivated (Yong-Gonzalez et al., 2007). Thus, mitotic dyskerin phenotypes in mammalian cells may be a consequence of reduced condensin levels within the spindle.

We propose that the functional consequence of condensin and Cbf5 binding to TFIIIC sites in tRNA genes is the building of a chromosome tether. A major mechanism for regulating chromosome dynamics is through tethering the centromere to a spindle pole or telomeres to the nuclear envelope (Verdaasdonk et al., 2013). Our tenet is that tethers throughout the chromosome dictate local dynamics that facilitate or restrict movement optimal for a given DNA metabolic process, such as DNA repair or mitotic force balance. In the pericentromere, the tethers are critical for force balance. It is currently thought that cohesin, by promoting sister chromatid cohesion is the basis for resisting microtubule forces from opposite spindle poles. However, pericentric cohesin is radially displaced from the spindle axis and therefore mechanically decoupled from direct microtubule-based force. In contrast, condensin is colinear with the kinetochore microtubules and likely to be involved in this process. The finding herein provides a mechanism for how the extensional forces are resisted. Condensin is cross-linked to different chromosomes to resist extensional forces through the network.

This provides a framework for integrating several recent findings regarding the distribution of TFIIIC binding sites. Incomplete TFIIIC sites are found throughout the genome (Moqtaderi et al., 2010). Condensin recruitment to these sites via Tfc3 could act in a similar fashion as condensin bound to tRNA genes in the pericentromere and for topological chromatin domains in Drosophila melanogaster (Van Borlte et al., 2014).

Upon anaphase onset, cohesin is degraded, and the pericentric chromatin rapidly migrates to the pole. Condensin is no longer required for force balance at this juncture. Instead, condensin has been shown to facilitate the recoil of chromosome arms to the spindle poles (Renshaw et al., 2010). This function is temporally correlated with the release of monopolar from the nucleolus. Monopolar is situated in a pole proximal position (Fig. 2), where its role in chromosome condensation is commensurate with a biological role in anaphase compaction (Fig. 5 C). Condensin’s critical role in the mitotic spindle is thus spatially and temporally partitioned by dyskerin for force balance in metaphase and monopolar for compaction in anaphase.

Materials and methods

Cell preparation
Cells were grown in YPD media (2% glucose, 2% peptone, and 1% yeast extract) at 24°C for WT, lrs4, and cbf5-AUU strains. Temperature-sensitive strains [cbf5-1 and ifc3-1sv115] were grown at 24°C and then shifted to 37°C 6 h before imaging. Temperature-sensitive strains containing cbf5-1 were provided by J. Carbon (University of California, Santa Barbara, Santa Barbara, CA), and ifc3-1sv115 was given by F. Uhlmann (London Research Institute, London, England, UK).

Microscopy
Images were acquired with a microscope stand (Eclipse TE2000-U; Nikon) with a 100× Plan Apochromat, 1.4 NA 100× digital interference contrast.
oil immersion lens with a camera (ORCA-AE; Hamamatsu Photonics) at room temperature (25°C). MetaMorph 7.1 (Molecular Devices) was used to acquire unblinded images of in a seven-step z series with a 300-nm step size of Smc4-GFP, kinetochore proteins Nu2-GFP and Cse4-2x-GFP, 12.5 kb LacO, T(

The number of occurrences of a LacO occupying a position within one linear fluorescent signal (aspect ratio > 1.2). Stretching events were determined as one focus and another

{\text{area}} \times \text{(area of inner region)} 

F_{\text{integrated intensity of outer region}} = \text{FSmc4} \times \text{area}{\text{of inner region}} + \text{FI} \times \text{area}{\text{of outer region}}. 

FSmc4 = \frac{\text{FI} \times \text{area}{\text{of inner region}}}{\text{area}{\text{of inner region}} + \text{FI} \times \text{area}{\text{of outer region}}}. 

Cells were grown to an OD_{600} of 0.4 in YPD. HCHO was added to a

HCHO was added to a

Chromatin IP

within <25% or >75% of spindle length (localized to the kinetochore and

quadrants were classified as within the middle 50% of spindle length, a

divided into four quadrants. Localization was determined as

to draw line scans in MetaMorph 7.1 were then logged to Excel and graphed

as a function of pixel intensity versus spindle length. The length of the spindle

was then divided into four quadrants. Localization was determined as

at the pole if any line scan peaks occurred in the outer two quadrants,

peaks occurring in the two inner quadrants were classified as

center of the axis.

peaks occurring in the two inner quadrants were classified as within the middle 50% of spindle length, whereas peaks falling within the outer two quadrants were classified as within <25% or >75% of spindle length (localized to the kinetochore and spindle pole region).

Chromatin IP

Cells were grown to an OD_{600} of 0.4 in YPD. HCHO was added to a final concentration of 1% for 20 min at room temperature followed by the adding of glycine to a final concentration of 125 mM. Pellets were resuspended in FA (fixation) lysis buffer (50 mM Hepes-KOH, pH 7.5, 300 mM NaCl, 1 mM EDTA, 1% Triton X-100, 0.1% sodium deoxycholate, and protease inhibitor cocktail) and vortexed with acid-washed glass beads for 1 h at 4°C. After sonication of the lysate, protein concentration of the whole cell extract was assayed using the protein assay kit (Dye Reagent Concentrate; Bio-Rad Laboratories). For IP reactions, whole cell extract was added to make final protein concentration 3 mg with 12 µl anti-GFP rabbit IgG fraction antibody (Life Technologies) and FA-lysis buffer to bring the final volume to 500 µl. IP reactions were allowed to rock at 4°C overnight before 12 µl FA-lysis buffer–washed Dynabeads Protein A (Life Technologies) was added to each IP and allowed to rock an additional 3 h. Anti- GFPs were eluted from the beads with 1% SDS and 0.1 M NaOH after sequential washing of the beads with FA-lysis buffer, FA-lysis buffer with 500 mM NaCl, LiCl solution, TE (10 mM Tris-HCl and 1 mM EDTA, pH 8.0), pH 8.0, and TE, pH 8.0, containing 20 µg RNAse A. 16 µl of 5 M NaCl was added to elute and input samples and allowed to incubate at 65°C overnight to reverse cross-links. DNA purification was completed using the PCR purification kit protocol (QIAGEN). PCR products run on 1% agarose gels were analyzed using MetaMorph 7.1. Integrated intensity of the bands on the gel was analyzed by drawing a rectangle to fit the band on the gel and a second rectangle around the first. Intensity was then calculated using the following formula: 

\[
F_{\text{band}} = F_{\text{band} - \text{background}}, \text{ in which } F_{\text{background}} = (F_{\text{inner}} - F_{\text{outer}}) \times (\text{area of inner region} - \text{area between perimeter of inner and outer regions}), \quad F_{\text{inner}} = \text{integrated intensity of inner region, and} \quad F_{\text{outer}} = \text{integrated intensity of outer region.}
\]

Kinetochore declustering and Tfc1 analysis

Population images were taken of Spc29-RFP and GFP-tagged kinetochore components (Nu2-GFP and Cse4-2x-GFP). Cse4-2x-GFP was provided by R. Baker (University of Massachusetts Medical School, Worcester, MA). Using MetaMorph 7.1, line scans were drawn through each sister kinetochore along the spindle axis (axial) or perpendicular to the spindle axis (radial) to determine whether the kinetochore was clustered (one peak) or declustered (multiple peaks). Similarly, population images of Tfc1-GFP and Spc29-RFP were rotated and aligned relative to the spindle axis using MATLAB. Line scans drawn through Tfc1 signal perpendicular to the spindle axis were used to determine inclusive peak-to-peak measurements.

LacO array analysis

Population images of metaphase cells were acquired of strains with 12.5 kb LacO arrays in WT, GalH3, mcm21α, bra1-9, and cb5-AUU mutants. Orientation relative to the spindle axis was determined as axial if both foci lie parallel to the spindle axis, whereas perpendicular orientation was determined by foci lying at a 90° angle to the axis. The 12.5 kb LacO array is a 32-mer repeat of the Lac operator (1.2 kb in length) introduced 12.7 kb from the centromere on chromosome XI (Pearson et al., 2001). The coordinates relative to CEN11 (440,246–440,129) are 427,362–427,412 on chromosome XI.

To determine the cross-correlation of pericentric movement (Fig. 4, D and E), strains containing a 10-kb LacO/LacI-GFP 1.8 kb from CEN15 (centrid of 6.8 kb, Gushima and Yangada, 2000), 8 kb Teo/TetR CFP 0.4 kb from CEN11 (centrod of 4.5 kb), and Spc29-RFP were built (Stephens et al., 2013c). The Teo/TetR arrays were amplified from the plasmid backbone described in Raftern et al. (2008). The coordinates relative to CEN11 (440,246–440,129) were 439,444–439,412 on chromosome XI. Images were acquired in single planes (binned 2x) over time at 15- or 30-s intervals. Each image at every time point was aligned horizontally with regard to the spindle axis using Spc29-RFP. Distance in nanometers (x axis) from LacO and Teo/TetR to the pole was measured at each time point, and correlation of these distances over time was calculated using the CORREL function in Excel.

Coordinated stretching was measured using the strain described in the previous paragraph by taking population images in 10 200-nm z steps (binned 2x). Stretching events were determined as one focus and another linear fluorescent signal (aspect ratio > 1.2). Stretching events were defined as uncoordinated if one array (either LacO or Teo/TetR) is stretched in the previous paragraph by taking population images in 10 200-nm z steps (binned 2x). Stretching events were determined as one focus and another linear fluorescent signal (aspect ratio > 1.2). Stretching events were defined as uncoordinated if one array (either LacO or Teo/TetR) is stretched in the
This information was transferred to MATLAB, in which an image was generated using the black body radiation spectrum to depict probability of the LaC0 position.

Online supplemental material

Fig. S1 shows that box H/ACA snoRNP complex members are necessary for condensin enrichment of the pericentromere. Fig. S2 shows localization and function of CBF5 and monopolin. Fig. S3 shows the concentration of CBF5-GFP determined by chromatin IP at sites of the genome where the RNA genes on chromosome III were deleted. Table S1 provides a list of the strains used in this study. Online supplemental material is available at http://www.jcb.org/cgi/content/full/jcb.201405028/DC1.

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