Interdomain allosteric regulation of Polo kinase by Aurora B and Map205 is required for cytokinesis

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Drosophila melanogaster Polo and its human orthologue Polo-like kinase 1 fulfill essential roles during cell division. Members of the Polo-like kinase (Plk) family contain an N-terminal kinase domain (KD) and a C-terminal Polo-Box domain (PBD), which mediates protein interactions. How Plks are regulated in cytokinesis is poorly understood. Here we show that phosphorylation of Polo by Aurora B is required for cytokinesis. This phosphorylation in the activation loop of the KD promotes the dissociation of Polo from the PBD-bound microtubule-associated protein Map205, which acts as an allosteric inhibitor of Polo kinase activity. This mechanism allows the release of active Polo from microtubules of the central spindle and its recruitment to the site of cytokinesis. Failure in Polo phosphorylation results in both early and late cytokinesis defects. Importantly, the antagonistic regulation of Polo by Aurora B and Map205 in cytokinesis reveals that interdomain allosteric mechanisms can play important roles in controlling the cellular functions of Plks.

Introduction

The cell division cycle is regulated by reversible protein phosphorylation that is spatiotemporally coordinated. The conserved Polo kinase is essential for several events of mitosis and cytokinesis (Archambault and Glover, 2009; Zitouni et al., 2014). Polo-like kinase (Plk) family members are defined by an N-terminal kinase domain (KD) and a C-terminal Polo-Box domain (PBD), which mediates protein interactions (Lowery et al., 2005; Park et al., 2010). In humans, Plk1 is the closest orthologue of Drosophila melanogaster Polo in its essential roles in cell division (Petronczki et al., 2008).

The complex functions of Plks are enabled by several regulatory mechanisms (Archambault and Glover, 2009; Zitouni et al., 2014). The PBD allows Polo to interact with substrates and adaptor proteins that recruit Polo to discrete locations in the cell including centrosomes, centromeres, and the midbody (Archambault and Glover, 2009; Park et al., 2010). The PBD is a phosho-binding module, and many of its interactions are facilitated by prior phosphorylation of the partner (Elia et al., 2003a,b; Park et al., 2010). However, some PBD-dependent partners of Plks do not require phoso-priming. This is the case for Map205, a microtubule-associated protein that binds and stabilizes Polo.

Instead of promoting formation of the complex, phosphorylation of Map205 at a Cdk site in early mitosis negatively regulates its interaction with Polo (Archambault et al., 2008).

Like several kinases, Plks are activated by phosphorylation in their T-loop (Qian et al., 1999; Archambault and Carmena, 2012). In humans, this phosphorylation of Plk1 occurs at Thr210 and is mediated by Aurora A kinase, with its cofactor Bora in G2 (Jang et al., 2002b; Macürék et al., 2008; Seki et al., 2008b). Although Bora is degraded in mitosis, persisting low levels of Aurora A–Bora maintain Plk1 activity until anaphase (Chan et al., 2008; Seki et al., 2008a; Bruinsma et al., 2014). In Drosophila, we have shown that Aurora B kinase, a member of the chromosomal passenger complex (CPC), is required for T-loop phosphorylation of Polo at centromeres in early mitosis (Carmena et al., 2012a,b). Failure of the CPC to activate Polo in prometaphase leads to chromosome alignment and segregation defects (Carmena et al., 2012a).

The function of a Plk is required for cytokinesis from yeasts to humans (Petronczki et al., 2008; Archambault and Glover, 2009). In human cells, Plk1 is targeted to the central spindle via its interaction with PRC1 (Neef et al., 2007). Similarly,
Figure 1. Phosphorylation of Polo by Aurora B regulates its localization in cytokinesis. (A) The localization of pT182-Polo at the midbody depends on Aurora B. Immunofluorescence in D-Mel2 cells is shown. Inhibition of Aurora B with Binucleine 2 reduced the pT182-Polo signal at the midbody (arrowheads). (B) The localization of Polo-GFP at the midbody depends on Aurora B. Binucleine 2 reduced the Polo-GFP signal at the midbody (arrowheads), but not the...
Drosophila Polo requires the PRC1 orthologue Fascetto for its recruitment to the spindle midzone (D’Avino et al., 2007). Plk1 is required for furrow formation by promoting the assembly of the HsCyc-4-Ect2 (RhoGAP-RhoGEF) complex, and subsequent RhoA activation and furrow ingression (Brennan et al., 2007; Burkard et al., 2007; Petronczki et al., 2007; Santamaria et al., 2007; Wolfe et al., 2009). Plk1 has been proposed to regulate additional proteins in cytokinesis, including at the midbody, before abscission (Petronczki et al., 2008; Bruinsma et al., 2012).

How Plk activities are regulated in cytokinesis is poorly understood. Here, we have investigated this question in Drosophila.

Results and discussion

Phosphorylation of Polo by Aurora B regulates its localization during cytokinesis

We showed that Polo is phosphorylated in its T-loop by Aurora B at centromeres in prometaphase (Carmena et al., 2012a). Like Polo, Aurora B localizes to the spindle midzone and midbody, and is required for cytokinesis (Ruchaud et al., 2007). Thus, we hypothesized that Aurora B could be required to activate Polo in cytokinesis.

We began to test this idea using an antibody against Polo phosphorylated at its activation loop site (pT182-Polo; Carmena et al., 2012a). In Drosophila cells, pT182-Polo is detected at the midbody (Fig. 1 A). To inhibit Aurora B, we used Binucleine 2 (Smurnyy et al., 2010). Strikingly, short treatments with Binucleine 2 abrogated the pT182-Polo signal at the midbody (Fig. 1 A). To test if this result reflected the lack of phosphorylation, or a failure to recruit Polo at this site, we used stably transfected cells allowing expression of Polo-GFP at levels close to endogenous Polo levels (Fig. S1 A). Binucleine 2 treatment clearly reduced the localization of Polo-GFP at the midbody (Fig. 1, B and C; and Fig. S1 B). However, the localization of Polo-GFP to microtubules of the central spindle that are adjacent to the midbody was still visible after Aurora B inhibition. This result was confirmed by live cell imaging (Fig. 1 D and Videos 1 and 2). Moreover, Binucleine 2–treated cells failed to complete cytokinesis, as is expected upon Aurora B inhibition (Smurnyy et al., 2010; Carmena et al., 2012b). The failure of Polo to localize to the midbody was not caused by the complete absence of this structure when Aurora B was inhibited, because Binucleine 2 treatment did not disrupt the midbody localization of Aurora B, Deterin-GFP, or Pavarotti-TAP (Fig. S1, C–F). Moreover, addition of Binucleine 2 to cells with a newly formed midbody greatly shortened the retention time of Polo-GFP at that site, which indicates that Aurora B activity is required to maintain Polo at the midbody after its recruitment (Fig. S1 G).

To test if the phosphorylation of Polo was required for its localization to the midbody, we examined the localization of a nonphosphorylatable form of Polo-GFP (PoloT182A-GFP). Strikingly, this mutant protein failed to localize to the midbody (Fig. 1, E and F). Conversely, the phosphomimetic PoloT182D-GFP mutant showed an increased localization at the midbody relative to the adjacent microtubules. Moreover, the T182D substitution largely rescued the localization of Polo-GFP at the midbody in the presence of Binucleine 2 (Fig. 1 C). Altogether, these results suggest that phosphorylation of Polo at Thr182 by Aurora B promotes Polo localization to the midbody in cytokinesis. In contrast, the T-loop state did not grossly affect the localization of Polo-GFP at kinetochores and centrosomes in early mitosis.

Phosphorylation of Polo by Aurora B promotes its dissociation from Map205

The pool of Polo that is localized on microtubules of the central spindle, and not to the midbody, is known to be bound to Map205 (Archambault et al., 2008). To investigate if T-loop phosphorylation of Polo by Aurora B could prevent or disrupt the interaction between Polo and Map205, we tested the copurification between PrA-Map205 and Thr182 mutant forms of Polo-Myc (Fig. 2 A). As expected, the association of the phosphomimetic PoloT182D-Myc mutant with PrA-Map205 was strongly diminished relative to PoloWT-Myc or PoloT182A-Myc.

To test if Aurora B activity could negatively regulate the interaction between Polo and Map205, we overexpressed Aurora B–Myc in cells also expressing Polo-GFP (wild type [WT]) or T182A and PrA-Map205 (Fig. 2 B). Polo-GFP phosphorylation was increased after Aurora B–Myc overexpression, as detected by Western blotting (Fig. 2 B). This signal depended on Aurora B kinase activity because it was abolished by treatment with Binucleine 2 (Fig. S2). Interestingly, overexpression of Aurora B–Myc abrogated the copurification of PoloWT-GFP, but not PoloT182A-GFP, with PrA-Map205 (Fig. 2 B). These results strongly suggest that T-loop phosphorylation of Polo by Aurora B negatively regulates the interaction between Polo and Map205. Introduction of a kinase-dead mutation (K54M; by analogy to the K82M mutation in Plk1; Kachaner et al., 2012) in PoloT182D did not rescue its interaction with Map205 (Fig. 2 A). Therefore, the conformational change itself, induced by T-loop phosphorylation of the KD of Polo, appears to determine the affinity of the PBD for Map205.

The KD inhibitor BI 2536 destabilizes the Polo-Map205 complex

To test the possibility that the shift from microtubules to the midbody observed with the PoloT182D mutant was dependent on its increased kinase activity, we tested the effect of BI 2536, a widely used Plk1 ATP-competitive inhibitor, on Polo localization in cytokinesis (Steegmaier et al., 2007). Intriguingly, short treatments with BI 2536 strongly reduced the microtubules/midbody...
Figure 2.  T-loop phosphorylation of Polo by Aurora B or interference with the interdomain interaction in Polo promotes its dissociation from Map205.  

(A) The phosphomimetic mutation T182D of Polo reduces its interaction with Map205. Cells were transfected as indicated and PrA-Map205 was purified. Purification products and whole cell extracts (WCE) were analyzed by Western blotting. 

(B) Overexpression of Aurora B results in hyperphosphorylation on July 9, 2017 jcb.rupress.org Downloaded from
localization ratio of Polo-GFP (Fig. 2 C). Moreover, BI 2536 strongly abrogated the copurification of PoloWT-GFP with PrA-Map205 (Fig. 2 D). As the effect of BI 2536 was similar to that of the T182D activating mutation in Polo, we hypothesized that it could be caused by the induction of a conformational change, and not to kinase inhibition per se. To test this hypothesis, we devised an in vitro experiment. Recombinant GST-Map205254-416 immobilized on Sepharose was added to cell extracts to bind Polo, the resin was then washed, and the purified complex was incubated with or without BI 2536 in an ATP-free buffer. Strikingly, BI 2536 destabilized the Polo–Map205 complex (Fig. 2 E). Therefore, the effect of BI 2536 on the Polo–Map205 interaction is not caused by the inhibition of Polo kinase activity. Addition of ATP to the Polo–Map205 complex had no effect.

A cocrystal structure has recently been published between the KD of zebrafish Plk1, its PBD, and a fragment of Drosophila Map205 (Xu et al., 2013). We used structural alignment to compare the conformation of the KD in this structure to that of the KD of human Plk1 bound to BI 2536 (Kothe et al., 2007). Only subtle differences were observed between the conformations of the KDs alone. However, we noticed a steric clash between BI 2536 itself and a conserved region of the PBD including Pro394, which mediates the PBD–KD contacts in zPlk1 (Fig. 2 F; Xu et al., 2013). Thus, binding of BI 2536 in the KD catalytic cleft likely disrupts its intramolecular contact with the PBD. Together with our finding that BI 2536 induces the dissociation of Map205 from Plk1, our results suggest that the KD contact with the PBD stabilizes its Map205-interacting conformation.

T-loop phosphorylation of human Plk1 has also been reported to abrogate the KD–PBD interaction (Jang et al., 2002a). Thus, the conformational change induced by T-loop phosphorylation of Polo likely leads to the dissociation of Map205 from the PBD by weakening the PBD–KD contact (Fig. 2, A and B). Loss of the interdomain interaction in Polo would then favor the alternative conformation of the PBD, which is competent for binding to canonical phosphorylated targets (Elia et al., 2003b). Crystal structures of full-length Polo/Plk1 in their inactive and active states will be needed to fully visualize how this interdomain allosteric regulation is achieved. Activating phosphorylation of Ser137 in the PBD-contacting region of the KD of human Plk1 has also been proposed to promote the dissociation between the KD and PBD (Xu et al., 2013). However, we have never detected phosphorylation at the equivalent site (Ser109) in Drosophila Polo, and a phosphomimetic mutation of this residue did not increase Polo kinase activity (unpublished data).

Map205 sequesters unphosphorylated Polo on microtubules and inhibits its kinase activity

If the model we proposed is correct, then removal of Map205 should facilitate the recruitment of Polo to the midbody. As previously reported, RNAi depletion of Map205 resulted in the loss of PoloWT-GFP on microtubules but did not reduce its localization at the midbody (Fig. 3 A; Archambault et al., 2008). Strikingly, depletion of Map205 restored the localization of PoloT182A-GFP at the midbody in a majority of cells. These results confirm that when bound to Map205, Polo requires phosphorylation for its localization to the midbody because this phosphorylation induces the dissociation of Polo from Map205, and not because it promotes its kinase activation.

Having found that modulation of the KD could impact the Polo–Map205 interaction, we asked if, in turn, Map205 could modulate Polo kinase activity. To test it, we used a small fragment of Map205 (aa 254–416) sufficient for Polo binding (Archambault et al., 2008). GST-Map205254-416 could pull down PoloWT-Myc or PoloT182D-Myc more efficiently than PoloT182A-Myc, which is consistent with the previous results (Fig. 3 B). Strikingly, addition of GST-Map205254-416 (but not GST alone) inhibited Polo kinase activity in vitro on casein and on itself (Figs. 3 C and S3, A and B). This is consistent with the recent crystal structure between a fragment of Map205 and both domains of zebrafish Plk1. This revealed that Map205 stabilizes a closed conformation of the PBD, which in turn interacts with the KD in its inactive conformation (Xu et al., 2013). It seems reasonable to assume that Map205 inhibits Drosophila Polo enzymatic activity by the same mechanism.

To test if T-loop phosphorylation of Polo regulates its susceptibility to inhibition by Map205, we immunoprecipitated Polo WT, T182A, or T182D, and tested the effect of GST-Map205254-416 on their kinase activities (Fig. 3 D). Although PoloWT-Myc and PoloT182A-Myc could be efficiently inhibited by GST-Map205254-416 PoloT182D-Myc retained significant activity. These results suggest that Map205 preferentially binds and inhibits unphosphorylated Polo. This is consistent with the absence of pT182-Polo antibody signal on microtubules, where Polo is bound to Map205 (Fig. S1 B).

Overall, these findings suggest an interesting molecular model for Polo regulation during cytokinesis where (1) Map205 sequesters Polo on central spindle microtubules, (2) phosphorylation of Polo by Aurora B abrogates the Map205–Polo interaction, and (3), when freed from Map205, Polo is recruited to the midbody (Fig. 3 E).
Figure 3. **Map205 sequesters unphosphorylated Polo on microtubules and inhibits its kinase activity.** (A) Silencing of Map205 rescues the midbody localization of PoloT182A-GFP. Cells were treated for 3 d with dsRNA against Map205 or the bacterial kanamycin resistance gene (control). (A, left) Immunofluorescence showing the localization of PoloWT-GFP or PoloT182A-GFP during cytokinesis. Arrowheads, midbody. Bar, 5 µm. (A, top right) RNAi depletion of Map205 on Polo at the midbody.

Figure 3B shows the GST pull-down experiment with different constructs. The table indicates the molar ratio (Map205/Polo) at various concentrations.

Figure 3C demonstrates the autoradiograph and Western blot results for the pull-down experiment. The blots are shown for Polo, GST, and Casein.

Figure 3D presents the IP experiment with different constructs. The blots are shown for Myc, GST, and IgG.

Figure 3E explains the mechanism of inactivated Polo activation by Aurora B and the role of Map205 in microtubule and midbody dynamics.
Activating phosphorylation of Polo is required for cytokinesis

Although Polo is known to be required for cytokinesis and Polo’s full activation is known to require T-loop phosphorylation, whether Polo phosphorylation is required for cytokinesis had never been tested. Unphosphorylated Polo shows a basal kinase activity (Fig. 3 D), which we have previously shown to be sufficient for bipolar spindle assembly (Carmena et al., 2012a). Depletion of Polo in control cells by transfection of a dsRNA targeting the 3’ UTR of the endogenous transcript resulted in the accumulation of defective mitotic cells and cell death (Carmena et al., 2012a). Expression of PoloWT-GFP largely rescued cell viability and the ability of these cells to divide properly (n = 276; Fig. 4, B and C; and Video 3). Expression of PoloT182A-GFP failed to rescue the viability of Polo-depleted cells, and many cells developed early mitotic defects (Carmena et al., 2012a). Nevertheless, many cells entered anaphase and attempted cytokinesis. Of them, >80% failed cytokinesis (n = 309; Fig. 4, B–D). This failure could occur either at the onset of cytokinesis, with incomplete or no furrow ingression (45% of these cells; n = 115; Video 4), or later, as cells failed to form a stable intercellular bridge (55% of the cells; n = 142; Video 5). In both cases, failure in cytokinesis resulted in binucleation. Replacement of endogenous Polo with PoloT182A-GFP, similar to PoloWT-GFP, allowed cytokinesis to proceed in most cells (Fig. 4, B and C; and Video 6). These results indicate that T-loop phosphorylation of Polo is required for cytokinesis.

Polo phosphorylation could be required in cytokinesis for two reasons: (1) to activate Polo’s intrinsic kinase activity and/or (2) to free Polo from Map205 and allow its localization to the midbody. We have shown that Map205 depletion restored PoloT182A-GFP localization at the midbody (Fig. 3 A). To try to discriminate between the two possibilities, we tested if the RNAi depletion of Map205 could rescue cytokinesis in PoloT182A-GFP–expressing cells depleted of endogenous Polo. As a result, cytokinesis was largely rescued (Fig. 4 E). Although this is a complicated experiment, which should be interpreted cautiously, the result suggests that the basal activity of unphosphorylatable Polo is sufficient for cytokinesis if it can be recruited to the midbody.

Antagonistic regulation of Polo by Aurora B and Map205 in cytokinesis

Here, we showed that Aurora B and Map205 control Polo activity during cytokinesis. We had previously shown that Map205 sequesters and stabilizes Polo on microtubules in interphase, and that overexpression of Map205 in embryos interfered with Polo function (Archambault et al., 2008). In addition, Map205 inhibits Polo enzymatic activity (Fig. 3; Xu et al., 2013). This effect is the opposite of that of phosphorylated peptides derived from canonical PBD targets, which increase Plk1 kinase activity (Elia et al., 2003b). Our findings also indicate that Aurora B–dependent phosphorylation on Thr182 releases a fraction of Polo from its sequestration by Map205, and allows its recruitment to the midbody. Aurora B localizes to the midzone in anaphase and to the midbody, but it does not colocalize with the microtubule-bound pool of Polo. However, a gradient of Aurora B activity is established around the midzone in anaphase in human cells (Fuller et al., 2008). Diffusion of Aurora B from the midzone/midbody could allow it to reach and phosphorylate Polo on microtubules to promote its recruitment to the midzone/midbody. Phosphorylation of Polo by Aurora B in early mitosis likely also contributes to its dissociation from Map205, a mechanism that would collaborate with the phosphorylation of Map205 by Cdk1 (Archambault et al., 2008; Archambault and Carmena, 2012; Carmena et al., 2012a). As map205 is not an essential gene (Pereira et al., 1992; Archambault et al., 2008), control of Polo by Map205 is not essential, but our work indicates that the inhibition of Polo by Map205 imposes a need for a mechanism that allows Polo to dissociate from Map205. Indeed, the Aurora B–dependent release of Polo from Map205 is required for cytokinesis.

The requirement for Plk1 activation during cytokinesis in human cells has never been investigated. Further work will be required to determine whether human Aurora A, Aurora B, or another kinase is acting upstream of Plk1 to activate it in cytokinesis. The absence of a clear Map205 orthologue and the absence of a strong localization of Plk1 to microtubules suggest that the mechanism reported here is not strictly conserved. However, the fact that Plk1 is competent for interaction with, and inhibition by, Drosophila Map205 suggests that physiological interaction partners may regulate Plk1 in a similar way (Fig. S3 C; Archambault et al., 2008).

Importantly, our findings demonstrate for the first time that a Plk (Polo) is subject to regulation by reciprocal interdomain mechanisms for its cellular functions (Fig. 5). Our results reveal this mechanism at work in the context of a physiological process: cytokinesis. While Map205 interaction with the PBD of Polo induces inhibition of its KD, phosphorylation of the activation loop in its KD induces dissociation of Map205 from the PBD, and facilitates PBD interactions with other targets, including at the midbody. The effects of various PBD-dependent interactors on Polo kinase activity and the impact of Polo KD
Figure 4. Activating phosphorylation of Polo is required for cytokinesis. (A) Expression of PoloWT-GFP, PoloT182A-GFP, and PoloT182D-GFP was induced with CuSO₄ (300 µM), and cells were transfected with Polo 3′ UTR dsRNA (or control KAN dsRNA). The next day, protein extracts were analyzed by Western blots. (B) Time-lapse imaging of cells depleted of endogenous Polo and expressing PoloWT-GFP, PoloT182A-GFP, or PoloT182D-GFP. Note the early cytokinesis failure. (C) % of cytokinesis failure. (D) % of cytokinesis failure. (E) % of cytokinesis failure.
activation on these interactions should be examined in the context of other functions in cell division. A good candidate as an additional allosteric inhibitor of Polo is Matrimony, a PBD-dependent interactor that genetically behaves as an antagonist of Polo during female meiosis (Xiang et al., 2007). Moreover, the importance of interdomain regulatory mechanisms for the functions of other Plk family members should be evaluated. A more precise understanding of how Plk1 intramolecular switching is regulated by interaction partners, kinases, phosphatases, and small molecules could inform the development of alternative or pathway-specific Plk1 inhibitors.

Materials and methods

DNA constructs

The expression vectors were generated by Gateway recombination (Invitrogen). Coding sequences were first cloned into the pDONR221 entry vector. They were then recombined into the relevant destination vectors for expression from copper-inducible (pMT) or constitutive (pAC5) promoters.

Stable cell lines expressing pMT-PoloWT-GFP, pMT-PoloT182A-GFP, and pMT-PoloT182D-GFP were obtained by cotransfection with the pCoBlast and selection in medium containing 20 µg/ml blasticidin as described previously (Archambault et al., 2008). Stable cell lines expressing pAC5-Deterin-GFP, pAC5-Pavarotti-TAP, and pAC5-Aurora B-Myc were obtained by cotransfection with pCoBlast and selection in medium (Invitrogen) supplemented with glutamine, penicillin, and streptomycin. Stable cell lines expressing pAC5-PoloWT-Myc, pAC5-PoloT182A-Myc, pAC5-PoloT182D-Myc, pAC5-PrA-Map205, pAC5-Aurora B-Myc, pAC5-Aurora B-GFP were obtained by cotransfection with the pCoBlast and selection in medium (Invitrogen). Stably transfected cells were sorted by FACS for the desired expression level of the GFP fusion proteins. Stable cell lines allowing the copper-inducible expression of Polo-GFP (WT, T182A, and T182D) were induced with CuSO4 (300 µM) for at least 1 d. For Aurora B inhibition, cells were treated with either DMSO or 20 µM Binucleine 2 (EMD Millipore) for 10 min before being processed for immunostaining. For pT182-Polo signal detection, cells were treated with 100 nM okadaic acid (BioShop Canada Inc.) for 1 h before lysis.

Immunofluorescence and Western blotting

Primary antibodies in immunofluorescence and Western blotting were anti–ß-tubulin DM1A from mouse (Sigma-Aldrich), anti-Myc 9E10 from mouse (Santa Cruz Biotechnology, Inc.), mouse monoclonal anti-Polo MA294 (a gift of D. Glover, University of Cambridge, Cambridge, England, UK), anti-pT210 Plk1 2A3 from mouse (recognizes Drosophila pT182 Polo; Abcam), rabbit anti-Map205 (a gift from A. Pereira, University of Massachusetts Medical School, Worcester, MA), anti-GFP from rabbit (#A6455; Invitrogen), anti-GST from rabbit (#2622; Cell Signaling Technology), and peroxidase-conjugated ChromPure rabbit IgG (for PrA detection; Jackson ImmunoResearch Laboratories, Inc.).

For immunofluorescence, cells were fixed with 4% formaldehyde for 10 min, then permeabilized and blocked in PBS containing 0.1% Triton X-100 and 1% BSA (PBSTB). Cells were incubated with primary antibodies diluted in PBSTB for 2 h at RT, washed three times in PBS, and incubated with secondary antibodies diluted in PBSTB for 1 h at RT. Cells were washed three times in PBS before being mounted in Vectashield medium with DAPI (Vector Laboratories).

Protein affinity purification and immunoprecipitation

Protein A affinity purifications were performed essentially as described previously (D’Avino et al., 2009). In brief, around 10 million cells were harvested and resuspended in 400 µl of lysis buffer (50 mM Tris-HCl, pH 7.5, 150 mM NaCl, 0.2% Triton X-100, 1 mM EDTA, and 10% glycerol) supplemented with protease inhibitors (1 mM PMSF, 10 µg/ml leupeptin, and 10 µg/ml aprotinin). Lysates were clarified by centrifugation at 14,000 rpm for 10 min in a tabletop centrifuge at 4°C. Supernatants were incubated for 1 h at 4°C with 20 µl of DynaBeads (Invitrogen) that we previously conjugated to rabbit IgG. Beads were washed five times with 1 ml of lysis buffer for 5 min at 4°C. Purification products were eluted by heating at 95°C.
for 5 min in 20 µl of Lømøml buffer twice (Sigma-Aldrich) and analyzed by Western blotting.

For immunoprecipitation of Polo-Myc (WT, T182A, and T182D), extracts were prepared as for Protein A affinity purifications and lysates were incubated with anti-Myc antibodies for 1 h at 4°C, then incubated with 20 µl of Protein G–conjugated Dynabeads (Life Technologies) for 45 min at 4°C, before being washed in lysis buffer.

**Transient transfections**

Transfections of D-Mel2 cells with plasmids were performed using Xtreme-GENE HP DNA Transfection Reagent (Roche) according to the manufacturer’s instructions. For RNA interference, cells were transfected in 6-well plates with 20 µg of dsRNA (Polo 3’ UTR dsRNA or Map205 dsRNA) using Transfast reagent (Promega). The control dsRNA was generated against the sequence of the bacterial kanamycin resistance gene. Cells were analyzed between 24 h and 72 h later by immunoblotting, immunofluorescence, or live cell imaging.

**GST pull-down assay**

For pull-down assays, pellets cells transfected with Polo-Myc (WT, T182A, and T182D) from confluent 75-cm² flasks were resuspended in lysis buffer (75 mM K·Hepes, pH 7.5, 150 mM KCl, 2 mM EGTA, 2 mM MgCl₂, 5% glycerol, 0.2% Triton X-100, 1 mM DTT, 1 mM PMSF, 10 µg/ml aprotonin, and 10 µg/ml leupeptin) and centrifuged for 15 min at 4°C. Clarified lysates were incubated with GST-Map205 (544-416), GST-Map205 (544-375), or GST Sepharose beads for 2 h at 4°C. Beads were washed five times with lysis buffer before SDS-PAGE and immunoblotting.

**Polo kinase expression and purification**

The coding region of *Drosophila* Polo was amplified by PCR and cloned into pFastbac-Hb vector. To express Polo protein, a bacmid was produced and transfected into SF-9 cells to generate the baculovirus according to the manufacturer’s protocol (Bac-to-Bac; Invitrogen). The virus was subsequently amplified to infect a larger quantity of cells (400 ml) in a spinner flask. 72 h after infection, cells were collected, washed with 1× PBS, and frozen until purification. To purify the recombinant protein, the SF-9 cell pellet was first lysed in lysis buffer (50 mM Tris, pH 7.5, 300 mM KCl, 10 mM imidazole, 0.5% Triton X-100, 10 mM β-mercaptoethanol, 1 mM PMSF, 10 µg/ml of leupeptin, pepstatin A, and chymostatin). The lysate was then spun at 40,000 rpm in a rotor (R-865; Sorvall) at 4°C for 40 min. The supernatant was recovered and allowed to bind to Talon resins (Takara Bio Inc.), pre-equilibrated with lysis buffer for 1 h at 4°C. Affinity resin-bound proteins were washed thoroughly with lysis buffer with 600 mM KCl and eluted with imidazole-containing buffer (50 mM Tris, pH 7.5, 250 mM imidazole, 300 mM KCl, and 1 mM DTT). Purified Polo was then dialyzed into storage buffer (50 mM Tris, pH 7.5, 300 mM KCl, 1 mM DTT, and 20% glycerol) overnight before flash freezing in small aliquots in liquid nitrogen.

**In vitro binding assay and kinase assays**

Purified GST-Map205 (544-416) were incubated with active purified Polo or hPkl1 or immunoprecipitated Polo-Myc (WT, T182A, and T182D) in binding buffer (75 mM K·Hepes, pH 7.5, 150 mM KCl, 2 mM EGTA, 2 mM MgCl₂, 5% glycerol, 0.2% Triton X-100, 1 mM DTT, 1 mM PMSF, 10 µg/ml aprotonin, and 10 µg/ml leupeptin) for 1 h at 4°C.

For kinase assays, reactions were performed in kinase buffer (20 mM K·Hepes, pH 7.5, 2 mM MgCl₂, and 1 mM DTT) with 0.5 µM ATP, [γ-³²P]-ATP, and 1 µg casein at 30°C for 15 min. For Polo inhibition in the kinase assay, BI 2536 was added at 300 nM. Reactions were stopped with the addition of the laemmli buffer and heating at 95°C for 2 min. Samples were separated by SDS-PAGE and transferred onto nitrocellulose for autoradiography and Western blotting.

**Microscopy**

Images of fixed cells were acquired on an inverted microscope (AxioImager) with a 40× objective lens (NA 1.4), and an Orca-R2 charge-coupled device (CCD) camera (Hamamatsu Photonics) with 2 × 2 binning, using Velocity 6.0 software (PerkinElmer). Binucleate 2 was added at anaphase onset. Live imaging shown in Fig. 4 was performed using a DeltaVision elite microscope (Applied Precision) and a 60x oil objective lens (NA 1.42) in a temperature-controlled environment (25°C). Images were acquired using a CoolSnap HQ2 camera (Photometrics). For each condition in this experiment, a minimum of three independent experiments was performed, a minimum of 91 cell divisions was recorded, and cytokinesis defects were counted manually. Images were treated using SoftWoRx (Applied Precision), ImageJ, and Photoshop (Adobe) software.

**Online supplemental material**

Fig. S1 shows additional experiments and controls accompanying Fig. 1. Fig. S2 shows that polo hyperphosphorylation at Thr182 induced by Aurora B overexpression depends on Aurora B kinase activity. Fig. S3 shows that Map205 inhibits the activity of Drosophila Polo and human Pkl1. Video 1 shows time-lapse imaging of a Polo-GFP–expressing cell in mitosis. Video 2 shows time-lapse imaging of a Binucleine 2–treated Polo-GFP–expressing cell in mitosis. Video 3 shows time-lapse imaging of a cell depleted of endogenous Polo and expressing Polo2566-GFP. Video 4 shows time-lapse imaging of a cell depleted of endogenous Polo and expressing PoloT182D-GFP showing an early cytokinesis failure. Video 5 shows time-lapse imaging of a cell depleted of endogenous Polo and expressing Polo1820-GFP. Online supplemental material is available at http://www.jcb.org/cgi/content/full/jcb.201408081/DC1.

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Figure S1. Additional experiments and controls accompanying Fig. 1. (A) Expression of Polo^{WT}-GFP, Polo^{T182A}-GFP, and Polo^{T182D}-GFP was induced with CuSO_{4} (300 µM). The next day, protein extracts were analyzed by immunoblotting with an anti-Polo monoclonal antibody. (B) The localization of Polo-GFP and pT182-Polo at the midbody depends on Aurora B. Cells expressing Polo-GFP were treated with DMSO or Binucleine 2, and stained for pT182-Polo (red). DNA is stained with DAPI, and colocalization of GFP (green) and pT182 Polo (red) appears in yellow. Arrowheads indicate the midbody. Bar, 5 µm. (C–F) Inhibition of Aurora B does not affect the localization at the midbody of Aurora B, Pavarotti, and Deterin. Cells expressing Polo^{WT}-GFP, Aurora B-GFP, Pavarotti-TAP, or Deterin-GFP were treated with DMSO or Binucleine 2, and examined by immunofluorescence as indicated. DNA is stained with DAPI. Arrowheads indicate the midbody. Bars, 5 µm. (G) The activity of Aurora B is required to maintain Polo-GFP at the midbody during cytokinesis. Polo-GFP–expressing cells having just formed a midbody were treated with Binucleine 2 or DMSO and imaged by time-lapse microscopy. The retention time of Polo-GFP was measured for at least 18 cells in each condition.
Figure S2. **Polo hyperphosphorylation at Thr182 induced by Aurora B overexpression depends on Aurora B kinase activity.** Cells expressing Polo^WT^-GFP or Polo^T182A^-GFP were transfected with Aurora B–Myc and treated with Binucleine 2 as indicated. 24 h after transfection, cells were treated for 1 h with okadaic acid (100 nM) and protein extracts were analyzed by Western blotting for pT182 Polo, GFP, Myc, and α-Tubulin.
Figure S3. Map205 inhibits the activity of Drosophila Polo and human Plk1. (A) In vitro kinase assays, using casein as a substrate, were performed in the presence of purified Polo (100 ng) mixed with purified GST-Map205[254-416] (100 ng) or GST alone (control). Protein phosphorylation was analyzed by autoradiography. Western blots were performed with anti-Polo and anti-GST antibodies. Casein was stained with amido black. (B) Control experiment for results shown in Fig. 3 C. In vitro kinase assays with Polo (100 ng) and casein as a substrate are shown. Increasing amounts of GST were added. Reactions were analyzed by autoradiography, Western blots, and amido black (total protein). No Polo kinase inhibition was observed. (C) Kinase assays were performed in the presence of purified active Plk1 (100 ng) and casein as a substrate. Increasing amounts of purified GST-Map205[254-416] were added as indicated. Phosphorylation of Plk1, Casein, and GST-Map205[254-416] was analyzed by autoradiography. Western blots were performed with anti-Plk1 and anti-GST antibodies. Casein was stained with amido black.
Video 1. **Time-lapse imaging of a Polo-GFP–expressing D-Mel cell in mitosis.** DMSO was added at anaphase onset. Images were acquired every 1 min on a spinning-disk confocal microscope. Bar, 3 µm.

Video 2. **Time-lapse imaging of a Binucleine 2–treated Polo-GFP–expressing D-Mel cell in mitosis.** Binucleine 2 was added at anaphase onset. Images were acquired every 1 min on a spinning-disk confocal microscope. Bar, 3 µm.

Video 3. **Time-lapse imaging of a D-Mel cell depleted of endogenous Polo and expressing Polo<sup>WT</sup>-GFP.** Images were acquired every 4 min on a DeltaVision microscope (Applied Precision).

Video 4. **Time-lapse imaging of a D-Mel cell depleted of endogenous Polo and expressing Polo<sup>T182A-GFP</sup> showing an early cytokinesis failure.** Images were acquired every 7 min on a DeltaVision microscope (Applied Precision).

Video 5. **Time-lapse imaging of a D-Mel cell depleted of endogenous Polo and expressing Polo<sup>T182A-GFP</sup> showing a late cytokinesis failure.** Images were acquired every 4 min on a DeltaVision microscope (Applied Precision).

Video 6. **Time-lapse imaging of a D-Mel cell depleted of endogenous Polo and expressing Polo<sup>T182D-GFP</sup>.** This cell successfully completes cytokinesis. Images were acquired every 7 min on a DeltaVision microscope (Applied Precision).