New Problems in the Functional Activity of the Pacinian Corpuscle. By T. Andrew Quilliam. (From the Department of Anatomy, University College, London, England.)*

During the last decade, investigations into the mode of function of the Pacinian corpuscle have been focussed almost exclusively on recording the activity of its nervous components, and some of the more recent histological studies (Quilliam and Sato, 1955; Pease and Quilliam, 1957) have contributed to interpreting the results so obtained. Lately, interesting structural modifications have come to light in end-organs of the Pacinian type found in various sites in different species. Thus, the presence of a system of intracorpuscular capillaries draining into nearby extracorpuscular glomerular arteriovenous anastomoses has been reported in human digital material (Cauna and Mannan, 1958), and the hitherto unsuspectedly complex morphology of the inner core of cat's mesenteric corpuscles has also been demonstrated by Pease and Quilliam (1957). These findings, together with the results obtained from employing micromechanical methods (Hubbard, 1956; Loewenstein and Rathkamp, 1958), now emphasise the desirability of considering the role played by the various non-nervous elements of this type of sense organ in its task of monitoring seemingly different processes taking place in diverse situations.

The outstanding feature characteristic of all Pacinian type corpuscles is their lamellated structure, and recently Hubbard (1958) has shown that the lamellae of the outer zone of the cat's mesenteric corpuscle (together with the fluid-filled spaces which intervene between them) act as a mechanical high pass filter, in that they effectively insulate the inner core and the nerve terminal within it from all the heterogeneous environmental stimuli, save those possessing high frequency components. The importance of this finding is that it is now unnecessary to ascribe the receptor's well known characteristic of quick adaptation to a maintained pressure stimulus solely to an inherent property of the nerve terminal itself. It also follows that, during effective stimulation, a radially directed force is momentarily applied to the inner core. As the nerve terminal is bare both of myelin and Schwann cell sheaths (Pease and Quilliam, 1957), this force might conceivably distort its surface membrane. The possibility that such a deformation may be involved in the production of a receptor potential has, together with other theories, already been briefly discussed by Gray (1956), but the part played by the complex inner core in channelling or otherwise modifying the circumferentially applied force is as yet obscure.

Another interesting problem requiring clarification emerges from the realization that the distribution of both true and pseudocholinesterase in cat's pancreatic corpuscles (Hebb and Hill, 1955; Portuguese, 1955; Coupland, 1958), and in human digital material (Beckett, Bourne, and Montagna, 1956) is restricted almost exclusively to the inner core. It may be that this distribution is related to the extraordinarily large surface area of the closely packed cells of the inner core in relation to their cytoplasmic volume. The presence inside the axially located nerve terminal of a peripheral palisade of large mitochondria and of structures similar to synaptic vesicles (Pease and Quilliam, 1957) also emphasises the probability that special metabolic processes take place at the nerve terminal.

Since they lie in close proximity to collagen fibrils, the lamellae of the outer zone are almost certainly highly modified fibrocytes, and they probably correspond to the outer sheath cells of the amphibian muscle spindle as described by Robertson (1956). Their putative origin is thus in keeping with their specialized "protective" type of function. Whether the cells of the inner core are also basically of a similar cell type is not so certain, as little is known about either their derivation or their function. Comparison of the appearance of the cytoplasm of these cells with that of the cells of the outer zone shows similarities in their strikingly flattened outlines and in the organelles which both contain, but discrepancies appear when the interrelationships of the peripheral parts of neighbouring cells in the two regions come to be considered and when the profusion and orientation of the fibrils associated with them are studied.

The intimate relationship which the cells of the inner core bear to the nerve terminal suggests the possibility that they may be related to Schwann cells, but there is, as yet, no clear evidence to support such a theory. No continuity between these cells and undoubted periaxonal Schwann cells found further proximally has yet been traced. Whilst the cleft between the two cellular groupings of the inner core is reminiscent of the gaps

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identified between neighbouring Schwann cells at
nodes of Ranvier (Robertson, 1957; Uzman and
Nogueira-Graf, 1957), in the latter situation there
is no multiplicity of cytoplasmic layers, as is the
case in the former. Since the cells of the inner core
bear some resemblance to both fibrocytes and
Schwann cells, it might be supposed that they
would best be provisionally classified as 'neural
fibroblasts' (Denny-Brown, 1946). However, until
a description of "neural fibroblasts" as seen with
the aid of the electron microscope is available, the
allocation of inner core cells to such a cell type
must be regarded as tentative only.

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An Electron Microscope Study of the Cells Lining the Small Blood Vessels of Helix pomatia
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INTRODUCTION
In the course of another investigation narrow
channels were observed in association with the
esophageal ganglia of Helix pomatia. The channels
are lined with flattened cells and are identified as
small blood vessels. The Golgi complex, mito-
chondria, and other cell inclusions are visible in
electron micrographs of these cells. Since little
work has been carried out on the ultrastructure
of the cells of invertebrate animals, we consider it
desirable to give a brief account of our observa-
tions.

Materials and Methods
Pieces of tissue surrounding the esophageal ganglia
of Helix pomatia were fixed in 1 per cent buffered
osmium tetroxide (pH 8.0) and embedded in a mixture
of N-butyl and methyl methacrylate.

Material for observation with the light microscope
was fixed in Bouin's picroformol (picric acid, saturated
aqueous solution 75 cc., formol 25 cc., acetic acid 5
cc.) and in Kolatchev solution (equal parts of 6 per
cent potassium dichromate, 1 per cent chromic acid,
and 2 per cent osmium tetroxide; followed by post-
osmication). Sections were subsequently cut at 5 µ
and 7.5 µ thickness.

OBSERVATIONS
The Golgi complex of the cells lining a small
blood vessel lies some little distance from the
nucleus in an area between the latter and the
lumen of the vessel. In electron micrographs of
relatively low resolution it is visible as three or
more groups of dense structures (Fig. 1). Large
dense deeply osmiophilic bodies are present in this
part of the cell and in the cytoplasm between the
Golgi complex and the lumen. A small number of bodies of similar appearance may occur in the cytoplasm close to the opposite pole of the nucleus. The nature of these bodies was not determined, but presumably they are composed of nutritive or storage material. A small number of large clear spaces, sometimes occurring singly and sometimes in contact with one another, are scattered through the cell but are not present in the vicinity of the Golgi complex. Each of these spaces appears to be enclosed by a single membrane; they possibly represent vacuoles the contents of which was dissolved out during the preparation of the tissue. When viewed with the light microscope, similar spaces are visible in cells fixed in Bouin's picroformol and in Kolatchev. In these preparations they are often larger and more numerous than in the cells shown in electron micrographs.

The examination of Kolatchev preparations with the light microscope is rendered difficult owing to the large amount of osmiophilic material present. Some of this material corresponds to the large dense bodies of electron micrographs. A number of curved rods and filaments are, however, frequently visible in the Golgi zone. In some cells they extend laterally round the anterior pole of the nucleus. We conclude that the elements of the Golgi complex revealed by the electron microscope are represented by the osmiophilic rods and filaments of cells in Kolatchev preparations.

An examination of electron micrographs of high resolution shows that the Golgi complex is composed of three components—closely applied flattened vesicles or saccules, vacuoles containing little dense material, and minute vesicles or granules (Fig. 2). The walls of the flattened vesicles are dense and their interiors comparatively clear. Frequently, a group of vesicles is intimately associated with one side of a Golgi vacuole, or almost completely surrounds a vacuole. Some of the flattened vesicles are continued from one group of vesicles to another so that, sometimes at least, neighbouring bundles are connected (Fig. 2).

The greater part of a Golgi vacuole (Fig. 2) appears in an electron micrograph as a clear space but a small amount of dense material is usually scattered through its interior. Some of this dense material seems to be membranous in form. Granules or vesicles with dense walls are scattered in the cytoplasm around the flattened Golgi vesicles and vacuoles. These, we believe, correspond to the small Golgi vesicles described by other workers.

While the present investigation was undertaken primarily with the object of determining the ultrastructure of the Golgi complex, three other components of the cytoplasm are worthy of mention. The cell or plasma membrane (Figs. 1 and 3) is clearly shown to possess intracellular folds or invaginations. Membranes composed of external dense components separated by a less dense area are present in the cytoplasm (Figs. 2 and 3). These differ in appearance from the profiles of the flattened Golgi vesicles in that they are not arranged in bundles and that the less dense inner region is considerably wider than the interior of a Golgi vesicle. We identify these structures as cytoplasmic membranes which constitute the endoplasmic reticulum. Sometimes the elements of the endoplasmic reticulum occur in the neighbourhood of the Golgi complex but do not appear to be continuous with the latter. Invaginations of the plasma membrane are frequently in close topographical relationship with the endoplasmic reticulum. It is probable that some of the latter are greatly extended invaginations of the cell membrane, but it is not possible in our material to determine with certainty whether such a connection does in fact exist.

Sections of mitochondria are shown in some of our electron micrographs (Figs. 1 and 3). Their external membranes are not well defined and their interiors are dense. In many cases the internal membranes are not visible while in others they are seen as poorly defined structures. Owing to the dense nature of the intermembranous region of a mitochondrion and the poor definition of the internal membranes, we are unable to determine the arrangement of the latter.

**DISCUSSION**

It is now widely accepted that the Golgi complex of the cells of vertebrate animals is composed of flattened vesicles, granules or small vesicles, and frequently large vacuoles (3, 8, 10). Further, it is claimed that the dictyosomes of germ cells and of somatic cells of invertebrates basically resemble in their fine structure the Golgi complex of vertebrate somatic cells (2-7). We believe that the Golgi complex of the cells forming the wall of the small blood vessels of *Helix* is made up of flattened vesicles, large vacuoles, and minute vesicles, and that it, therefore, closely resembles the complex of the somatic cells of vertebrates. It is of interest that in this animal neighbouring bundles of flattened Golgi vesicles are sometimes connected by...
a few vesicles. A somewhat similar condition was previously observed in the neurons of *Patella vulgata* (7, 9). According to Lacy (7) the Golgi lamellae of *Patella* branch and anastomose to form a network. In *Helix* branching appears to be less marked than in *Patella* and it is doubtful whether the condition in the former could be described as a true network.

Dalton and Felix (3) believe that the structures seen in electron micrographs, and formerly described as Golgi membranes, are in reality "profiles of flattened sacs." A similar claim is made by Grassé and Carasso (6). Our conclusions support this view.

The dense nature of the mitochondria of the cells lining the small blood vessels of *Helix* is rather surprising. It is likely that the large dense bodies present in the cytoplasm arise from mitochondria, but there is no direct evidence in support of this suggestion. Reference to the relevant literature indicates that the fine structure of the mitochondria of invertebrates varies considerably according to the type of cell in which they are resident. Beams and Tahmisian (1), for example, claim that the mitochondria of the male germ cells of *Helix* differ from those of other cells examined prior to their study. They comment that "it may be that further investigation will establish the difference as due to a variation in structure within the mitochondria of certain animal species." More recently Dalton and Felix (3), with reference to the mitochondria of vertebrate animals and of some unicellular organisms, remark that "the number, spacing and orientation of the internal membranes . . . appeared to be characteristic for each cell type."

It is worthy of note that the elements of the endoplasmic reticulum shown in our electron micrographs are few in number and are not associated with granules. Beams, Tahmisian, Devine, and Anderson (2) consider that a "Golgi lamella" of the male germ cells of *Nemobius sp.* is sometimes extended as "a string of relatively small bead-like vacuoles . . . in close approximation to elements of the endoplasmic reticulum." In some of our electron micrographs the interiors of some of the flattened vesicles at the end of a bundle are wider than elsewhere, and consequently present an appearance somewhat similar to that described by Beams et al. There is, however, no evidence that the flattened Golgi vesicles of *Helix* are directly associated with cytoplasmic membranes.

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**EXPLANATION OF PLATE 192**

**FIG. 1.** Electron micrograph of part of two cells from a small blood vessel of *Helix pomatia*. Large dense bodies (B.) Clear spaces (S.). Part of the nucleus (N.), the nuclear membrane (N.M.), and the cell membrane (P.M.) are shown. Small clear spaces at the periphery are surrounded by invaginations of the cell membrane. A group of flattened Golgi vesicles (G.M.) is present between the nucleus and the lumen. M., mitochondria. X 9,273.

**FIG. 2.** Electron micrograph. Flattened Golgi vesicles (G.M.). Golgi vacuoles (G.V.). Small Golgi vesicles or granules (G.G.). At (A) the connections between adjacent bundles of flattened Golgi vesicles are shown. Cytoplasmic membranes (endoplasmic reticulum) (C.M.) are cut at various angles. X 85,000.

**FIG. 3.** Electron micrograph of small part of peripheral region of a cell. The cell membrane (P.M.) and intracellular invaginations of the cell membrane (I.P.M.) are shown. Cytoplasmic membranes (endoplasmic reticulum) (C.M.) are cut at various angles. M., mitochondrion. X 38,636.
(.Threadgold and Gresson: Cells of Helix small blood vessels)