ASSEMBLY OF LIPIDS INTO MEMBRANES IN
ACANTHAMOEBA PALESTINENSIS

I. Observations on the Specificity and Stability of Choline-\(^{14}\)C
and Glycerol-\(^{3}\)H as Labels for Membrane Phospholipids

FRANCIS J. CHLAPOWSKI and R. NEAL BAND

From the Department of Zoology, Michigan State University, East Lansing, Michigan 48823.
Dr. Chlapowski's present address is the Department of Anatomy, University of Massachusetts
Medical School, Worcester, Massachusetts 01604

ABSTRACT

In order to determine the feasibility of using radioactive precursors as markers for membrane
phospholipids in Acanthamoeba palestinensis, the characteristics of phospholipids labeled with
choline-\(^{14}\)C and glycerol-\(^{3}\)H were examined. Choline-\(^{14}\)C was found to be a specific label for
phosphatidylcholine. There was a turnover of the radioactive moiety of phosphatidyl chol-
line at a rate that varied with the concentration of nonradioactive choline added to the
growth medium. Radioactivity was lost from labeled phosphatidyl choline into the acid-
soluble intracellular pool and from the pool into the extracellular medium. This loss of
radioactivity from cells leveled off and an equilibrium was reached between the label in the
cells and in the medium. Radioactive choline was incorporated into phosphatidyl choline
by cell-free microsomal suspensions. This incorporation leveled off with the attainment
of an equilibrium between the choline-\(^{14}\)C in the reaction mixture and the choline-\(^{14}\)C moiety
of phosphatidyl choline in the microsomal membranes. Therefore, a choline exchange reac-
tion may occur in cell-free membranes, as well as living A. palestinensis. In contrast to cho-
line-\(^{14}\)C, the apparent turnover of glycerol-\(^{3}\)H-labeled phospholipids was not affected by
large concentrations of nonradioactive choline or glycerol in the medium. The radioactivity
in lipids labeled with glycerol-\(^{3}\)H consisted of 33\% neutral lipids and 67\% phospholipids.
Phospholipids labeled with glycerol-\(^{3}\)H turned over slowly, with a concomitant increase in
the percentage of label in neutral lipids, indicating a conversion of phospholipids to neutral
lipids. Because most (~96\%) of the glycerol-\(^{3}\)H recovered from microsomal membranes
was in phospholipids, whereas only a minor component (~2\%) of the glycerol-\(^{3}\)H was in
the phospholipids isolated from nonmembrane lipids, glycerol-\(^{3}\)H was judged to be a specific
marker for membrane phospholipids.

INTRODUCTION

Because phospholipids are a ubiquitous component of biological membranes, radioactive phospho-
lipid precursors are often used to label membranes (1, 2, 3, 4, 5, 6, 7). As a prelude to studies of
membrane phospholipids in the soil ameba, Acan-
thamoeba (=Mayorella) palestinensis (8, 9), we ex-
examined the specificity and stability of choline-\(^{14}\)C as a marker for phosphatidyl choline and glycerol-
\(^{2}\)H as a marker for phospholipids in membranes. Other investigators have reported exchange reactions of choline (10), serine (11, 12), ethanolamine (13, 14, 15), and inositol (16) in cell-free systems, but the existence of these reactions in vivo has been questioned (7, 17). Recently, new evidence has been obtained in vivo for an exchange of bases between choline and phosphatidyl choline in liver (18). Data presented in this report indicate that the choline moiety of phosphatidyl choline may undergo an exchange reaction in living *A. palestinensis*, as well as in cell-free microsomal membrane suspensions. The results also indicate that glycerol-\(^{3}\)H is a specific and relatively stable label for membrane phospholipids.

Portions of this work have been reported briefly (19, 20).

**METHODS**

**Cell Culture**

*A. palestinensis* was cultured axenically in silicone-coated flasks containing proteose peptone-glucose medium (21). Cells were incubated at 29°C with agitation (100 rpm) on a rotary shaker (22) and grew exponentially with a doubling time of 27 hr. Growth was monitored by direct counting in all experiments.

**Radioactive Labeling**

Choline-Me-\(^{14}\)C (7.6 \(\mu\)Ci/\(\mu\)mole) and glycerol-
\(^{2}\)H (500 \(\mu\)Ci/\(\mu\)mole) were purchased from New England Nuclear Corp., Boston, Mass. Cells were grown during incorporation experiments, at an initial concentration of \(~10^6\) cells per milliliter in fresh medium containing either 1 \(\mu\)Ci choline-\(^{14}\)C/ml (132 nmoles/ml) or 5 \(\mu\)Ci glycerol-\(^{2}\)H/ml (10 nmoles/ml). For turnover experiments, cells were grown for 12 hr in medium supplemented with either 0.25 \(\mu\)Ci choline-\(^{14}\)C/ml (33 nmoles/ml) or 2 \(\mu\)Ci glycerol-
\(^{2}\)H/ml (4 nmoles/ml). At the end of the labeling period the cells were collected and washed twice in unlabeled medium by centrifugation at 125 \(g\) for 5 min. The washed cells were either prepared for radioactivity determinations, as described below, or were brought to 1 ml (original volume) with ice-cold 10% trichloroacetic acid (TCA), and stored for at least 2 hr at 4°C. The acid-insoluble fraction was centrifuged for 10 min at 16,000 \(g\) in a Sorvall SS-34 fixed-angle rotor. Portions of the clarified, acid-soluble fraction were removed and counted directly in scintillation fluid. The acid-insoluble material was washed twice in ice-cold 9% TCA and once in distilled water by centrifugation at 16,000 \(g\) for 10 min to remove residual soluble radioactivity (1). The TCA-insoluble precipitate was extracted for lipids, as will be described, or was dissolved completely in 98% formic acid and transferred to scintillation vials for counting. In this manner, the acid-soluble and acid-insoluble radioactivity of cells could be related directly to the number of cells per milliliter of growth medium.

Cell radioactivity was determined by restoring the pellets of washed cells recovered from 1 ml of medium to a volume of 1 ml with distilled water, resuspending the cells and counting portions in scintillation fluid. The total radioactivity of 1 ml of medium, from which the cells had been removed by centrifugation, was ascertained by the counting of portions of that medium in scintillation fluid.

**Cell Fractionation**

In experiments to determine the percentage of lipids labeled with glycerol-\(^{2}\)H in microsomes and lipid droplets, an isolation medium containing 0.25 \(m\) sucrose, 0.005 \(m\) Tris-HCl, and 0.002 \(m\) MgSO\(_4\) (pH 7.5) was used. To isolate microsomal fractions from unlabeled cells for cell-free reaction mixtures, the isolation medium contained 0.25 \(m\) sucrose and 0.1 \(m\) phosphate buffer at a pH of 7.4 (13).

Labeled or unlabeled cells from 150 to 200 ml of medium were harvested and washed twice in ice-cold isolation medium by centrifugation at 125 \(g\) for 5 min in a Sorvall HB-8 swinging-bucket rotor. Thereafter, all steps were carried out at 4°C.

The wet, packed cells were resuspended in 15 ml of isolation medium and homogenized with seven strokes of a 30 ml Potter-Elvehjem glass-Teflon grinder (Arthur H. Thomas Co., Philadelphia, Pa.)
at 2500 rpm. The homogenate was centrifuged at 10,000 g for 25 min in a Sorvall SS-34 fixed-angle rotor to remove nuclei, mitochondria, and cell debris. The supernatant was decanted with essentially all of the floating lipids and centrifuged for 1.5 hr at 100,000 g in a Spinco 30 fixed-angle rotor to yield the microsomal and postmicrosomal fractions. The top one-fourth of the postmicrosomal supernatant was decanted, taking care to recover all of the floating lipids. Presumably the floating lipids were derived mainly from lipid droplets seen in fixed cells. The remaining postmicrosomal supernatant was discarded. The lipid droplets and the microsomal pellet were resuspended separately in isolation medium and washed by centrifugation at 100,000 g for 1.5 hr.

In experiments where the cells had been labeled with glycerol-3H, the washed microsomes and lipid droplets (floating lipid layer and top one-fourth of the postmicrosomal supernatant) were precipitated with 10% (final concentration) TCA and allowed to stand for a minimum of 2 hr at 4°C. Before lipid extraction the acid-insoluble material was washed as previously described.

**Cell-Free Reaction Mixture**

The cell-free system was prepared according to the method of Vandor and Richardson (13). Microsomal fractions, isolated as described above, were diluted to 1 mg protein/ml as determined by the Folin-Ciocalteau test (23), with a standard solution containing 0.821 mM CaCl₂ and 0.1 mM phosphate buffer (pH 7.4). Either 2 or 4 ml of the suspension was brought to 5.25 ml with the standard solution to yield a mixture containing 2 or 4 mg of microsomal protein. The cell-free suspensions were allowed to warm from 4°C to 29°C in a water bath. After temperature equilibrium (~5 min), 1 µCi choline-14C (132 nmoles) or 5 µCi glycerol-3H (10 nmoles) was introduced into the reaction mixture in 0.25 ml of the standard solution. The complete reaction mixture contained 2 or 4 mg of microsomal protein, 132 nmoles choline-14C or 10 nmoles glycerol-3H, and 5 µmoles CaCl₂, made up to a total volume of 5.5 ml with 0.1 mM phosphate buffer (pH 7.4). After incubation at 29°C, with agitation, the reactions were terminated at specified time intervals by adding equal volumes of ice-cold, 20% TCA. The TCA suspensions were stored for a minimum of 2 hr at 4°C before washing and extraction of lipids.

**Lipid Isolation**

The total lipids of washed TCA precipitates of cells or cell fractions were extracted twice with 10 volumes of chloroform:methanol (2:1) for 2 hr at room temperature. The extracts were washed with 0.1 N HCl to remove residual protein (24). Separate portions were used for chromatographic analysis, lipid phosphorus determination, and scintillation counting.

The pooled chloroform:methanol extracts were spotted on activated silica gel plates and developed in chloroform:methanol:acetic acid:water (23:15:4:2) (25). The separated phospholipids were detected by iodine vapors and scraped, together with the associated silica gel, into scintillation vials for counting. Alternatively, the extracts were evaporated to dryness, and the lipids were dissolved in chloroform and separated into neutral lipids and phospholipids by column chromatography with silicic acid (26). Lipid phosphorus was determined according to Bartlett (27). Portions of lipid extract were evaporated to dryness in counting vials before the addition of scintillation fluid.

**Liquid Scintillation Counting**

Radioactive counting of aqueous solutions, as well as of lipid extracts evaporated to dryness in scintillation vials, was done in 15 ml of P-dioxane scintillation fluid containing 7.84 g/liter 2,5-diphenyloxazole (PPO), 0.16 g/liter bis-MSB and 120 g/liter naphthalene (New England Nuclear Corp.). A Packard Tri-Carb liquid scintillation counter was used to determine counting rates, which were corrected for background and quenching to disintegrations per minute (dpm).

**RESULTS**

**Specificity of Labeling**

When cells were grown in medium containing initial concentrations of 1 µCi choline-14C/ml (132 nmoles/ml) or 5 µCi glycerol-3H/ml (10 nmoles/ml) essentially all of the acid-insoluble radioactivity of cells was extractable in the total lipid fraction. That is, approximately 98% of the 14C-radioactivity and ~95% of the 3H-radioactivity was recoverable in chloroform: methanol (2:1) extracts when samples were analyzed at 1, 8, 24, and 72 hr after introduction of isotope into the medium. Apparently little, if any, choline-14C or glycerol-3H was converted to a nonlipid, acid-insoluble form by cells growing in proteose-peptone glucose medium.

In thin-layer chromatography (25) of the lipids of cells grown for 12 hr in medium containing 1 µCi choline-14C, all of the radioactivity was localized in phosphatidyl choline. The radioactive lipids of cells labeled for 12 hr with 2 µCi glycerol-3H/ml medium consisted of 33% neutral lipids and 67% phospholipids, as separated by column
Therefore, direct counting of TCA precipitates could be used to measure the amount of choline-14C in phosphatidyl choline or the amount of glycerol-3H in phospholipids plus neutral glycerides.

**Characteristics of Incorporation of Radioactive Precursors into Cellular Lipids**

Incorporation of choline-14C into the acid-insoluble fraction of cells reached its maximum rate about 45 min after introduction of radioactive choline into the medium (Fig. 1 A, one of three identical experiments). Coinciding with the beginning of rapid incorporation of labeled choline into phosphatidyl choline (acid-insoluble fraction), the rate of increase in specific activity of the soluble intracellular pool slowed down, but did not level off. Thus, a large and increasing intracellular pool of soluble radioactivity was maintained throughout incorporation.

In contrast to choline-14C, incorporation of glycerol-3H into phospholipids and neutral glycerides (acid-insoluble fraction) was evident immediately after exposure of the cells to the radioactive label (Fig. 1 B, one of two identical experiments). Furthermore, no appreciable soluble pool of radioactive precursors accumulated, demonstrating that essentially all of the glycerol-3H which entered the cells was quickly incorporated into lipids.

**Characteristics of Turnover of Radioactive Labels in Cellular Lipids**

When different concentrations of nonradioactive choline (0.033, 3.3, and 16.5 µmole/ml) were added to the fresh medium of three identical, labeled cell cultures, the measurable rates of turnover of the choline-14C in phosphatidyl choline (acid-insoluble fraction) increased with increasing concentrations of nonradioactive choline (Fig. 2, one of three identical experiments). The highest concentration (16.5 µmoles nonradioactive choline/ml medium) inhibited growth and induced encystation. However, no effect on growth was observed with the lower concentrations (i.e., 0.33 and 3.3 µmoles/ml). The soluble intracellular pool of radioactive molecules did not increase enough to account for the loss of radioactivity from phosphatidyl choline (acid-insoluble fraction) in any of the turnover experiments (see Fig. 2); therefore, radioactivity must have been lost from the soluble cell pool to the medium.

Indication that this was the case was obtained when labeled cells were grown in medium supplemented with 3.3 µmoles nonradioactive choline/ml. Then the loss of radioactivity to the medium was measurable for ~70 hr, after which an equilibrium was reached between radioactivity in the cells and that in the medium (Fig. 3, one of two identical experiments). At each of the time points shown in Fig. 3, radioactive medium was cleared of cells by low-speed centrifugation. Scintillation counting of the acid-insoluble fractions of the medium revealed no radioactivity. Therefore, the molecules that escaped into the medium probably were free choline-14C, unless a soluble form of phosphorylated choline, such as phosphorylcholine or cytidinediphosphatecholine, had passed into the medium.
In summary, addition of nonradioactive choline to the medium caused a loss of radioactivity from phosphatidyl choline into the soluble cell pool, which, in turn, equilibrated with the medium. Apparently, nonradioactive choline either induced an increased turnover of the entire phosphatidyl choline molecule or caused an "exchange" of the labeled choline moiety of phosphatidyl choline.

To distinguish between these two alternative explanations, the characteristics of glycerol-3H-labeled phospholipids were examined. No turnover of glycerol-3H-labeled, acid-insoluble material (total cellular lipids) was apparent, and essentially no acid-soluble pool of radioactive molecules could be detected in cells chased with 3.3 µmoles nonradioactive choline/ml (Fig. 4, one of three identical experiments). The results were identical if the medium had been supplemented
TABLE I
**Turnover of Glycerol-3H-Labeled Phospholipids in the Presence of Large Concentrations of Nonradioactive Choline or Glycerol**

<table>
<thead>
<tr>
<th>Concentration of chase</th>
<th>Experiment No.</th>
<th>Time in chase medium</th>
<th>0 hr</th>
<th>24 hr</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>dpm/µmole lipid P</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>16.5 µmoles glycerol/ml</td>
<td>1</td>
<td>508,171</td>
<td>246,500</td>
<td>(456,000)*</td>
</tr>
<tr>
<td></td>
<td>2</td>
<td>406,895</td>
<td>182,140</td>
<td>(355,085)*</td>
</tr>
<tr>
<td>16.5 µmoles choline/ml</td>
<td>1</td>
<td>460,018</td>
<td>451,000</td>
<td>(451,000)*</td>
</tr>
<tr>
<td></td>
<td>2</td>
<td>629,030</td>
<td>584,830</td>
<td>(584,830)*</td>
</tr>
</tbody>
</table>

Cells from four exponentially growing cultures were suspended at a concentration of ~10^6 cells per milliliter in fresh medium containing 2 µCi glycerol-3H/ml (4 nmoles/ml). After a 12 hr labeling period, the cells were washed and resuspended to a concentration of ~10^6 cells per milliliter in fresh medium containing the indicated concentrations of nonradioactive glycerol or choline. Duplicate samples were taken immediately (0 hr) or 24 hr later, and analyzed for the radioactivity in phospholipids.

* The specific activities corrected for the observed growth dilution are given in parenthesis. No growth was observed when 16.5 µmoles choline/ml was added to the medium, whereas growth was normal (i.e., 27 hr doubling time) when 16.5 µmoles glycerol/ml was used.

Inasmuch as no turnover of glycerol-3H was apparent in the total cellular lipids (Fig. 4), although some turnover was observed in isolated phospholipids (Table I), it seemed likely that some of the glycerol-3H in phospholipids had been converted to neutral lipid. To test this possibility, the percentage of total lipid radioactivity in neutral lipids was analyzed in cells grown for 12 hr in medium containing 2 µCi glycerol-3H/ml (4 nmoles/ml) as well as in cells subsequently cultured for 24 hr in chase medium containing 16.5 µmoles nonradioactive glycerol/ml. In two identical experiments the radioactivity of total cellular neutral lipids increased from about 33% during incorporation to ~38% after 24 hr in medium containing nonradioactive glycerol.

**Cell-Free Incorporation and Turnover**

In order to determine whether an “exchange” reaction was operational in vitro, cell-free mixtures of microsomal membranes were incubated with choline-14C. Incorporation of choline-14C into the acid-insoluble fraction of the microsomes increased as the protein concentration of the fraction was increased (Fig. 5, one of three identical experiments). Chromatography demonstrated that all of the acid-insoluble radioactivity was in phosphatidylcholine. A divalent cation requirement was suggested by the inhibition of incorporation in the presence of ethylenediaminetetraacetic acid, (EDTA) (Fig. 5). Glycerol-3H was not incorporated by this cell-free system when substituted for choline-14C in the reaction mixture. As much as 132 µmoles of nonradioactive choline, added to the reaction mixture along with the radioactive glycerol, did not induce any measurable exchange of glycerol. Thus, as with living cells, only the choline moiety of phosphatidyl choline seemed to be involved in this phenomenon.

Net incorporation of choline-14C into phosphatidylcholine by cell-free microsomes was initially linear as a function of time, but stopped after 30–45 min (Fig. 6, one of two identical experiments). The addition of concentrated nonradioactive choline, either before or after this leveling off, not only stopped measurable incorporation of choline-14C but effected a rapid turnover of label in the microsomal phosphatidyl choline as if free and bound choline were in equilibrium.
Cell-free reaction mixtures containing 2 or 4 mg microsomal protein were incubated with 1 µCi choline-14C (132 nmoles/ml) for 60 min of incorporation at 29°C, when the reactions were terminated by the addition of 10% (final concentration) TCA.

- microsomes; ▲ microsomes + 0.05 µmoles disodium EDTA.

Incorporation of choline-14C into cell-free microsomal suspensions and the effect of chasing with nonradioactive choline. 2-mg samples of microsomal protein were incubated as for Fig. 5. In the reactions terminated at 60 and 120 min, nonradioactive choline (9----9) or buffer alone (0---0) had been added at 15 and 60 min, respectively. (0---0: 132 µmoles nonradioactive choline in 0.1 ml of 0.1 M phosphate buffer [pH 7.4]; •---•: 0.1 ml of 0.1 M phosphate buffer [pH 7.4].)

**DISCUSSION**

As observed by Nagley and Hallinan (2) in rat liver cell fractions and by Plagemann (4, 5) in rat hepatoma cells, essentially all of the acid-insoluble choline-14C was localized in the phosphatidyl choline of growing *A. palestinensis*.

In the ameba, the choline moiety of phosphatidyl choline was assumed to be in equilibrium with free choline in the cell and in the extracellular medium (Figs. 2 and 3). The addition of large concentrations of nonradioactive choline induced a turnover of the radioactivity already incorporated into phosphatidyl choline.

In contrast, the apparent rate of turnover of phospholipids labeled with glycerol-3H was slow and was not affected by the addition of nonradioactive choline or glycerol (Table I). Phosphatidyl choline has been reported to comprise 45% of the phospholipids in a similar soil ameba (28); thus, any change in its rate of turnover should have been reflected in measurements of the total phospholipid fraction. Therefore, it seems likely that the observed turnover of choline-14C involved only the choline moiety of phosphatidyl choline and not the entire phospholipid molecule. In other words, the phenomenon probably was due to an exchange reaction rather than to the degradation and *de novo* synthesis of phosphatidyl choline.

Neither the soluble form of choline involved nor the mechanism of exchange in these cells is known. In growing rat hepatoma cells, where the possibility of a choline exchange reaction has been suggested by Plagemann (5), only phosphorylcholine was found in the soluble cell pool. On the other hand, cytidinediphosphatecholine has been reported to be present and in equilibrium with...
phosphatidyl choline in both normal (29) and choline-deficient (17) rat liver. In choline-deficient liver, an increased synthesis and secretion of plasma lecithin and not an exchange of choline reportedly results from the administration of large doses of choline (7, 30). However, the latter results have been challenged by Treble et al. (18) who presented evidence that a base exchange of choline to phosphatidyl choline occurs in vivo in liver at four times the rate at which phosphoryl choline is converted to lecithin.

The rapid attainment of an equilibrium between free choline-\(^{14}\)C and the radioactive moiety of phosphatidyl choline in cell-free microsomes (Fig. 6) was in sharp contrast to the slower kinetics of this reaction in living cells (Figs. 2 and 3) which perhaps was limited by a slow rate of transfer of choline in and out of the cells. Consistent with this, Plagemann and Roth (31) have reported that transport is the rate-limiting step in the incorporation of choline into the acid-insoluble pool of rat hepatoma cells in vitro. The enzyme(s) or cofactor(s) needed for the exchange reaction must have been bound to membranes, since only inorganic ions and choline were added to the microsomal fraction. Cell-free exchange reactions, similar to the phenomenon described here, have been reported for choline (10), serine (11, 12), ethanolamine (13, 14, 15), and inositol (16).

Under normal growth conditions, no exogenous choline is required by these cells (21). However, cell growth and morphology appeared normal in medium supplemented with up to 3.3 \(\mu\)moles nonradioactive choline/ml, a concentration at which the choline-\(^{14}\)C moiety of phosphatidyl choline was assumed to be exchanging (Fig. 2 B). While it is hard to envision exchange reactions in equilibrium with the natural environment as being biologically significant to soil ameba, the observations of this study could represent the amplification of a local and normally occurring intracellular process. Inasmuch as phospholipids are important components of cellular membranes, it is conceivable that exchange reactions play some role in membrane function.

The implication of these experiments on the use of choline-\(^{14}\)C as a marker for membrane phosphatidyl choline in \(A.\) \textit{palestinensis} is obvious; it is an unsuitable label if large chase concentrations of nonradioactive choline are used, because of the resulting instability of the label.

In contrast, glycerol-\(^{3}\)H was found to be a relatively stable marker of cellular phospholipids. It also appears to be a specific marker for membrane phospholipids. Although \(~33\%\) of the incorporated glycerol-\(^{3}\)H was recovered from cellular neutral lipids, most of these neutral lipids were confined to lipid droplets, whereas the glycerol-\(^{3}\)H in membranes was almost entirely phospholipid (Table II).

The apparent rate of turnover of total cellular phospholipids labeled with glycerol-\(^{3}\)H was not affected by large chase concentrations of nonradioactive choline or glycerol (Table I). During the course of turnover the amount of label in cellular neutral lipids relative to cellular phospholipids increased slightly. Because little, if any, turnover of total cellular lipids (acid-insoluble material)
was apparent (Fig. 4), it seems likely that some membrane phospholipids were slowly converted to neutral lipids and transferred to lipid droplets.

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