The Localization of Protein Carboxyl-Methylase in Sperm Tails

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ABSTRACT  Protein carboxyl-methylase (PCM), an enzyme known to be involved in exocytotic secretion and chemotaxis, has been studied in rat and rabbit spermatozoa. PCM activity and its substrate methyl acceptor protein(s) (MAP) were demonstrated in the supernate after solubilization of the sperm cell membrane by detergent (Triton X-100). A protein methylesterase that hydrolyzes methyl ester bonds created by PCM was demonstrated in rabbit but not in rat spermatozoa. This enzyme was not solubilized by nonionic detergent.

The specific activities of PCM in rat spermatozoa from Caput and cauda epididymes were similar and lower than that found in testis. By contrast, MAP substrates were low in testis and increased in parallel with sperm maturation in the epididymis. Multiple MAP were demonstrated in spermatozoa by polyacrylamide gel electrophoresis. The pattern of these proteins was similar in spermatozoa from different portions of the reproductive tract. Fractionation of heads and tails of rat spermatozoa on sucrose gradients indicated that PCM was found exclusively in the tail fraction, whereas MAP was detected both in head and tail fractions.

The presence of all the components of the protein carboxyl-methylation system in spermatozoa and the localization of PCM and some of its substrates in the sperm tail are consistent with their involvement in sperm cell motility.

Protein carboxyl-methylase (PCM) is one of several enzymes that produces a reversible posttranslational modification of protein structure that, in turn, leads to altered function. This enzyme methylates the free carboxyl groups of glutamate or aspartate residues in protein substrates (methyl acceptor proteins, MAP) using S-adenosyl methionine (SAM) as a methyl donor (14-16, 18, 23). The methyl ester bonds are hydrolyzed, either enzymatically by a protein methylesterase (PME) (9) or spontaneously in neutral and alkaline pH, to yield methanol. A consequence of these reactions is the reversible neutralization of the negative charge on substrate proteins.

The methanol-forming enzyme was first discovered in 1965 (1) and was later identified as PCM (14, 27). This enzyme has been associated with several cellular functions, including exocytic secretion (4-8, 34) and chemotaxis (19-21, 29-33). Experiments on bacterial chemotaxis showed that methylation of several membrane proteins was stimulated by chemotactic agents in wild-type organisms (19, 20, 31-33), but not in nonchemotactic mutants (20). Furthermore, the relationship between protein carboxyl-methylation and chemotaxis has also been demonstrated in leukocytes, monocytes (29, 30), and macrophages (21).

Although PCM is ubiquitous in mammalian cells, the highest activities are found in endocrine glands and in nervous tissues. Because this enzyme activity is especially high in testis (16), it was of interest to determine whether it was primarily localized in the secretory or germinal elements. A study of testicular cells fractionated on a bovine serum albumin gradient demonstrated that the highest activities of both PCM and MAP were in spermatids with relatively little activity in Sertoli's cells (10). Additional studies showed that PCM activity in the young rat increased in parallel with germinal development and was low in animals with x-ray-induced or genetically determined germ cell deficiency (10).

In view of the observation that protein carboxyl-methylation was localized in germ cells, we postulated that the neutralization of negative charge brought about by this reaction could be
involved in the motility of spermatozoa. If this postulate is correct, then PCM activity should be found in the sperm tail of motile spermatozoa. The present study was conducted to investigate this possibility.

MATERIALS AND METHODS

Materials

S-adenosyl-L-[methyl-3H]homocysteine (SAM), 10-15 Ci/mmol, was obtained from New England Nuclear (Boston, Mass.). Pig skin gelatin, bovine serum albumin, Triton X-100, and cetylpyridinium-chloride (CPC) were purchased from Sigma Chemical Co. (St. Louis, Mo.); sucrose (ultrapure grade) from Schwarz/Mann Div., Becton, Dickinson & Co. (Orangeburg, N. Y.); S-adenosyl-L-homocysteine (SAH) from Boehringer Mannheim Biochemicals (Indianapolis, Ind.); and Coomassie Brilliant Blue R-250 from Bio-Rad Laboratories (Richmond, Calif.).

Tests

Testes were obtained from sexually mature rats (100- to 140-g-old Sprague-Dawley rats from Charles River, Wilmington, Mass.) and mature rabbits (New Zealand white from Marland Farms, Hewitt, N. J.). Before assay, a 10% homogenate was prepared in 0.3 M sucrose with a Polytron homogenizer (Brinkmann Instruments Inc., Westbury, N. Y.).

Isolation of Spermatozoa

Rat spermatozoa were obtained from cauda and cauda epididymis as follows: Cauda epididymis was minced in a petridish containing warm Krebs-Ringer bicarbonate (30°C). The petridish was then placed for 15 min in a humidified air/CO2 (95%/5%) incubator (Heinicke Instruments, Inc., Hollywood, Fla.) at 37°C. After this incubation, all the visible contaminant particles were taken out by filtration through a 50-μm mesh nylon filter. To isolate cauda spermatozoa, the epididymis was dissected free from the interface between the 2.05 and the 2.20 M sucrose, and head pellets were resuspended in 0.3 M sucrose with a Polytron homogenizer (Brinkmann Instruments Inc., Westbury, N. Y.).

Separation of Sperm Heads and Tails

Sperm heads and tails were separated by equilibrium density gradient centrifugation according to Calvin (2). Motile cauda spermatozoa were isolated from rats and washed in 0.02 M Na-phosphate buffer, pH 6.0 (buffer P), and then centrifuged at 1,000g for 15 min. The washing procedure was repeated.

Electrophoretic Analysis of the MAP

Sperm cell proteins were methylated with [methyl-3H]SAM by purified PCM and electrophoresed under acidic conditions in the presence of the detergent CPC in 10% acrylamide gels as previously described (7). The gels were stained with 0.25% Coomassie Brilliant Blue, destained, and sliced in 1-mm fractions. Each slice was incubated overnight with 10 ml of Liquifluor-toluene mixture containing 3% of Protosol (New England Nuclear, Boston, Mass.). The radioactivity was measured in a liquid scintillation spectrometer.

RESULTS

Effect of the Solubilization of Sperm Cell Membrane by Detergent

Because PCM has been associated with cell motility and because a large fraction of the PCM activity and MAP capacity in testis is in spermatozoa, we first determined whether this enzyme was present in spermatozoa. A nonionic detergent was used in the assays because it can dissolve sperm membranes without altering the machinery of movement (13). After treatment of intact rat and rabbit sperm with detergent (Triton X-100 at a final concentration of 0.1%), PCM activity and MAP capacity were predominant in the soluble fraction (Table I). The results obtained with detergent-solubilized sperm were the same as those after sonication.

PME Activity

Because PME is a newly identified component of the protein carboxyl-methylation system in mammals, this enzyme activity
TABLE I
Effect of Treatment by Triton X-100 on PCM Activity, MAP Capacity, and PME Activity in Rat and Rabbit Sperm

<table>
<thead>
<tr>
<th></th>
<th>Soluble fraction</th>
<th>Insoluble fraction</th>
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</thead>
<tbody>
<tr>
<td></td>
<td>pmol/mg protein</td>
<td>pmol/mg protein</td>
</tr>
<tr>
<td>PCM activity</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Rat caudal sperm</td>
<td>28.5 ± 8.3*</td>
<td>8.8 ± 3.0</td>
</tr>
<tr>
<td>Rabbit ejaculated</td>
<td>4.4 ± 0.9</td>
<td>2.1 ± 0.4</td>
</tr>
<tr>
<td>MAP capacity</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Rat caudal sperm</td>
<td>50.3 ± 16.9</td>
<td>19.5 ± 4.4</td>
</tr>
<tr>
<td>Rabbit ejaculated</td>
<td>67.1 ± 10.6</td>
<td>33.3 ± 4.5</td>
</tr>
<tr>
<td>PME activity</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Rat caudal sperm</td>
<td>1.0</td>
<td>N.D. §</td>
</tr>
<tr>
<td>Rabbit ejaculated</td>
<td>10.0</td>
<td>N.D.</td>
</tr>
</tbody>
</table>

* Mean ± SE.
† Number of experiments.
§ N.D., not detectable.

Table I:

Rat and rabbit sperm were treated by detergent (Triton X-100, 0.1%) and centrifuged at 10,000 g for 30 min to separate the soluble fraction from the insoluble. The pellet was resuspended in 0.5 ml of 0.3 M sucrose. PCM activity, MAP capacity, and PME activity were determined as described in Materials and Methods.

was examined in spermatozoa (Fig. 1). The PME activity in rabbit spermatozoa was high (33 pmol/10 min per milligram of protein) relative to that reported for other tissues (9), but was undetectable in the rat sperm. Although this enzyme was present in both rat and rabbit testes, the activities were low (1 and 2 pmol per milligram of protein, respectively). The presence of an inhibitor of PME activity in rabbit testis was excluded by an experiment in which samples of testis and sperm were mixed. After treatment by detergent, PME activity was located predominantly in the nonsoluble fraction (Table I).

PCM Activity and MAP Capacity in Rat Testis and Epididymis

The PCM activity and MAP capacity in testis are compared in Table II with those of spermatozoa from different portions of the epididymis. Because PCM and MAP are predominantly in the detergent-soluble fraction, assays were performed both on solubilized spermatozoa (Triton X-100, 0.1%) and on high-speed-produced supernate obtained from these detergent-treated spermatozoa. The specific activity of PCM was much lower in rat spermatozoa than in the testis. Enzyme-specific activities in spermatozoa from different portions of the epididymis were in the same range with slightly higher values in those from caput. By contrast, the MAP capacity in the high-speed-produced supernate increased between testis and cauda epididymis in parallel with sperm maturation (Table II).

The PCM activity and MAP capacity (131.9 and 11.7 pmol per milligram of protein, respectively) in the high-speed-produced supernate of rabbit testis were similar to those of rat. In rabbit sperm as in rat, the MAP capacity was also higher (38.3 pmol per milligram of protein) and the PCM activity lower (6 pmol per milligram of protein) than in testis cytosol.

Analysis of the MAP by Polyacrylamide Gel Electrophoresis (PAGE)

Rat spermatozoa from caput and cauda epididymis were first treated by detergent (Triton X-100, 0.1%) and then reacted with [methyl-³H]SAM in the presence of excess PCM; the methylated proteins were fractionated on CPC-PAGE (Fig. 2). The patterns of methylated proteins were similar, regardless of the origin of the rat sperm. In addition, the pattern of methylated proteins in rabbit spermatozoa was similar to that of rat.

Localization of the Enzymatic System

Having demonstrated PCM and MAP in spermatozoa, we next determined the distribution of these activities in fractionated heads and tails. The head and tail fractions were highly purified and the cellular components were well preserved, as judged by light and transmission electron microscopy (Figs. 3 and 4). In particular, the acrosomes were present and generally intact, even though the plasma membrane was absent. Structures of the tail, including dense fibers, mitochondria, flagellar tubules, and fibrous sheath, appeared intact.

>95% of the recovered PCM activity was in the tail fractions (12.6 ± 1.7 pmol per milligram of protein; n = 5). Little or no activity was recovered in the head fraction. For each experiment a sample of nonfractionated sperm from the same animal was assayed, and the PCM specific activity was found to be similar to that of the tail fragments (16.8 ± 5 pmol per milligram of protein; n = 4). By contrast, the MAP capacity was higher in the head (61.5 ± 9.8 pmol per milligram of protein; n = 3) than in the tail fraction (21.1 ± 4.3, n = 3).

DISCUSSION

Rapid and transient modification of the protein charge brought about by the addition of methyl groups to the free carboxyl...
FIGURE 2 PAGE analysis of rat sperm MAP. Rat and rabbit sperm proteins were methylated with [methyl-3H]SAM by PCM and electrophilized at pH 2.2 in the presence of the detergent CPC in 10% acrylamide gels. The gels were stained with 0.25% Coomassie Brilliant Blue and then sliced for measurement of radioactivity by liquid scintillation counting. Cytochrome c (Cyt c) was added to the samples as a marker (7).

In the present study, these latter experiments were extended by demonstrating the components of the carboxyl-methylation system in mammalian spermatozoa. In the rat, maturation of spermatozoa in the epididymis seems correlated with an increase in MAP capacity. The ratio of MAP/PCM increases 10-fold from testis to cauda epididymal spermatozoa. The presence of PCM and its substrates in the soluble extract after treatment by detergent is in agreement with studies in other tissues (5, 11). By contrast, in rabbit the MAP/PCM ratio increases 60-fold between testis and ejaculated spermatozoa. Rabbit spermatozoa were also distinguished from those of rat by the presence of PME activity. The significance of these species differences must await further study.

The localization of the PCM activity in sperm tail suggests its involvement in motility. This hypothesis was prompted by the fact that PCM is involved in chemotaxis of bacteria and leukocytes (19, 20, 29, 30-33). This postulate was subsequently strengthened by the observations that patients with immotile spermatozoa have very low PCM activity compared with normal controls (12). Although the exact function of PCM in the sperm tail remains to be elucidated, the rapidity with which the reaction occurs and its reversibility suggest that it could act as a “switch” for reversal of the beat. Alternatively, protein carboxyl-methylation could be involved in sperm chemotaxis. Although chemotaxis has not been demonstrated in mammalian spermatozoa, it has been clearly demonstrated in spermatozoa of some marine invertebrates (25, 26).

Certainly protein carboxyl-methylation plays a part in the control of behavior at the cellular level in many systems (11, 33). One possible approach to understanding these mechanisms is the characterization of the specific substrates. It has been shown here that several proteins are substrates in spermatozoa. The analysis and exact location of these proteins should help to clarify the role of protein carboxyl-methylation in signal transduction. It is of note that several substrates for PCM were observed in sperm heads. Although no enzyme activity was recovered in this fraction, its presence in sperm heads cannot be excluded, because the plasma membrane was lost during purification of heads and tails.

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FIGURE 3  (a) The head fraction contains only very slight contamination with tail elements and no observable contamination with cell debris. × 4,000. (b) Acrosomes appear intact by morphological criteria. The plasma membrane is missing. × 30,000.
**Figure 4**

(a) The tail fraction is highly pure. × 4,000. (b) The midpiece appears intact except for the missing plasma membrane. × 25,000. (c) The fibrous sheath maintains the segmented appearance although the principal piece is fragmented by the sonication. × 18,000.
REFERENCES


