Isolation and Partial Characterization of a Cage of Filaments That Surrounds the Mammalian Mitotic Spindle

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ABSTRACT Mitotic cells have been detergent extracted under conditions that support microtubule assembly. When HeLa cells are lysed in the presence of brain tubulin, mitotic-arrested cells nucleate large asters and true metaphase cells yield spindles that remain enclosed within a roughly spherical cage of filamentous material. Detergent-extracted mitotic Chinese hamster ovary (CHO) cells show a similar, insoluble cage but the mitotic apparatus is only occasionally stabilized. In later stages of mitosis, HeLa cages are observed in elongated and furrowed configurations. In the terminal stages of cell division, two daughter filamentous networks are connected by the intercellular bridge. When observed in the electron microscope the cages include fibers 7-11 nm in diameter. The polypeptide composition of cages isolated from mitotic HeLa cells is complex, but the major polypeptides are a group with mol wt ranging from 43,000-60,000 daltons and a high molecular weight polypeptide. CHO cells contain a subset of these proteins which includes a major 58,000-dalton and a high molecular weight polypeptide. Two different antisera directed against the vimentin-containing intermediate filaments bind to polypeptides in the electrophoretic profiles of isolated HeLa and CHO cages and stain the cages, as visualized by indirect immunofluorescence. These results suggest that the HeLa and CHO cages include intermediate filaments of the vimentin type. The polypeptide composition of HeLa cages suggests that they also contain tonofilaments. The cages apparently form as the cells enter mitosis. We propose that these filamentous cages maintain the structural continuity of the cytoplasm while the cell is in mitosis.

Mitosis in mammalian cells is accompanied by a dramatic reorganization of cytoplasmic and nuclear components. The cytoskeleton of interphase is largely dissolved during prophase, and the distribution of cytoplasmic membranes is grossly perturbed with the breakdown of the nuclear envelope (12, 16, 42, 43, 23). Even overall shape is altered at mitosis, given the tendency of most cells to alter their physical attachment to neighboring surfaces and round up (28, 57). Within the nucleus the nucleolus dissociates and the chromatin condenses into chromosomes. The actions of the mitotic spindle on the organization of the chromosomes and the subsequent segregation of the chromatids are perhaps the best-studied rearrangements in the mitotic cell. Cytoplasmic reorganization, on the other hand, is a less well characterized process. It is clear that the interphase organization breaks down during mitosis, and it is equally clear that the organization is reestablished after telophase with the postmitotic reentry into interphase. Several stimulating papers have recently shown that there is often a similarity between the two daughters of a cell division, suggesting that some factors are inherited that help to set up cytoplasmic organization in the next generation (1, 51). What these components might be and how the cells might remember their interphase organization are at present questions requiring further investigation.

A previously successful approach to the study of factors important in cell architecture has been that of lysing and extracting cells with conditions that preserve the fibrous elements of the cytoplasm (10, 36, 25, 20). There are several methods of lysis applicable to mitotic cells which preserve some or all of the mitotic spindle or the cleavage furrow (37, 39, 41, 48, 58), but little attention has yet been paid to other components of the mitotic cell.
In this paper and the accompanying one (26), we present the result of a search for new lysis conditions that would extract most of the protein of a mitotic mammalian cell, yet stabilize the microtubule components of the mitotic apparatus (MA). We have found a buffer that extracts >80% of the cell's protein, yet leaves the stabilized spindle enclosed within a fibrous cage of detergent-insoluble proteins. This cage appears to be deformable, as we have isolated it in a sequence of shapes that reflects the geometry of the dividing cell. By microscopic, biochemical, and serological criteria we demonstrate that the cages include 10-nm filaments of the vimentin type (18, 30) in both HeLa and Chinese hamster ovary (CHO) cells. These isolated structures are most likely related to the cages of intermediate filaments that have recently been observed to form around the developing mitotic spindle, using indirect immunofluorescence (6, 18, 30). The cages represent fibrous elements of the cytoplasm that persist throughout mitosis, and we suggest that such a structure might serve as a framework for the reestablishment of interphase organization after cell division.

MATERIALS AND METHODS

Cell Culture

HeLa cells were grown as monolayer cultures in Dulbecco's modified Eagle's medium (Grand Island Biological Co. [GIBCO], Grand Island, New York) supplemented with 7% calf serum (GIBCO). Monolayers of CHO cells were maintained in Ham's F12 (GIBCO) supplemented with 10% fetal calf serum (GIBCO). In some preparations mitotic cells were detached from the monolayer by gently shaking. In others, cells were partially synchronized with a single thymidine block and release (9), followed by accumulation in 0.04 μM nocodazole and shaking-off as previously described (62). Populations of late stage mitotic cells were obtained by releasing Nocodazole-accumulated cells into drug-free medium for 50 min in the case of HeLa cells and 12 min for CHO cells (62).

Cell Fractionation

All steps of the fractionation procedure were carried out at 37°C. Mitotic cells were collected by centrifugation at 200 g for 3 min, resuspended in a hypotonic buffer (1 mM PIPES, pH 6.9, 2 mM EGTA, 1 mM MgCl₂, 5% dimethyl sulfoxide [DMSO], 95% D₂O) for 2 min, pelleted by centrifugation at 200 g for 2 min, and lysed in microtubule (MT) assembly buffer (0.5 M PIPES, pH 6.9, 1 mM MgCl₂, 1 mM EGTA, 5% DMSO, 1% Triton-X 165, 0.5% sodium deoxycholate, 0.2% SDS, 1 mM GTP, 2 mM phenylmethylsulfonyl fluoride [PMSF], 1% Aprotinin [Sigma Chemical Co., St. Louis, Mo.] (26)) containing 2 mg/ml of porcin brain microtubule protein (MTP). The MTP was prepared by the method of Weingarten et al. (59). After the fourth cycle of polymerization, the protein was resuspended at a concentration of 4 mg/ml in 0.5 M PIPES, 1 mM EDTA, depolymerized at 0°C, and centrifuged for 4 h at 225,000 g at 4°C to produce MTP which showed a lag in spontaneous assembly (2).

Lysed cells were digested with 20 μg/ml of DNase I (Worthington Biochemical Corp., Freehold, N. J.) to reduce viscosity. Some preparations were further extracted in 1 M KCl, 10 mM Tris, pH 7.5, 1 mM EDTA for 1 h, collected at 1000 g for 4 min, and then resuspended in 1 M KCl, 100 mM Tris, pH 8.7, 1 mM EGTA, 100 mM β-mercaptoethanol for an additional hour before use (34).

Microscopy

For light microscopy, preparations were sealed under coverslips and observed with a Zeiss Photomicroscope II using phase, differential interference contrast (DIC) and polarization optics. All photographs were taken on Kodak SO 115 film (ASA 100) developed with Dufine.

Whole mounts of the lysed cell preparations were prepared for electron microscopy by fixation for 10 min in 2% glutaraldehyde with 2% MgCl₂ in 0.1 M PIPES buffer, pH 6.9. Immediately after fixation, lysed cells were centrifuged at 100 g for 3 min on a sucrose (95%, w/v) cushion and then resuspended in 0.1 M Na cacodylate, pH 7.4, 10% sucrose, stained with 1% osmium in the same buffer for 2 min, and then rinsed in distilled water, dehydrated with a concentration series of acetone or 2-methoxyethanol, and dried by passing through the critical point of CO₂ (3). The specimens were then carbon coated on both sides and examined in a JEM 1000. Thin sections of intact cells were prepared as previously described (40).

SDS Polyacrylamide Electrophoresis

Material to be analyzed by SDS polyacrylamide electrophoresis was dialyzed against 0.1% SDS, precipitated with 10 vol of acetone, and then collected by centrifugation at 20,000 g for 10 min. Samples were resuspended in sample buffer and electrophoresed according to the procedure of Laemmli (33). Molecular weight determinations were calculated relative to known polypeptide standard by assuming that mobility is proportional to the log of the molecular weight.

Immunological Analysis

Antigens were identified in SDS polyacrylamide gels by the procedure of Burridge (11). After electrophoresis and fixation without staining, gels were equilibrated with buffer at neutral pH and overlaid with a 1:15 dilution of antisera for 16-24 h followed by 6 d of rinsing. Gel lanes were then overlaid with Staph A protein (Pharmacia Inc., Piscataway, N. J.) labeled with 125I (29) for 16-24 h and further rinsed for 5 d. The gel slices were then stained with Coomassie Blue, dried, and analyzed by autoradiography.

Indirect immunofluorescence of intact cells was performed as previously described (12). Lysed cells were fixed for immunofluorescence in 0.1 M HEPES, pH 6.9, 3 mM MgSO₄, 1.2% formaldehyde, and 0.1% glutaraldehyde for 5 min. Fixed preparations were then collected by centrifugation at 500 g for 4 min and rinsed in phosphate-buffered saline (PBS), followed by 10% goat serum. These cells were resuspended in immune serum diluted 1:20 for 25 min at 25°C, rinsed in phosphate buffer saline without calcium (PBS), incubated in fluorescein-conjugated goat anti-rabbit IgG (Miles Laboratories Inc., Elkhart, Ind.) for 20 min at 37°C, and then rinsed extensively. Preparations were settled onto polylysine-coated coverslips and mounted in Elvaco (DuPont Instruments, Wilmington, Del.). Micrographs were taken with Kodak Plus X film developed in Dufine using Zeiss epifluorescence optics.

RESULTS

Lysis of Mitotic HeLa and CHO Cells

Treatment of mitotic cells (Fig. 1a) with the hypotonic buffer described above preserves spindle birefringence (BR) while the cells swell (Fig. 1b). The subsequent lysis with a detergent-containing MTP solution solubilizes >80% of the cell protein as judged by colorimetric protein estimation (7) and disperses the chromosomes, leaving MT-containing ghosts (Fig. 1c-e). HeLa cells accumulated with Nocodazole and lysed without release of the block produce large, calcium-sensitive asters (26), while later stages yield highly birefringent, spindle-shaped structures. The size of the asters and the BR of the spindle is affected by the concentration of polymerizable MTP in the lysis mixture. Regardless of this concentration, however, the isolated mitotic centers and associated MTs are enclosed within a thin refractile cage. The cages of lysed, metaphase HeLa cells average 18 μm in diameter and are similar in size to the intact cell before swelling. The MA of CHO cells is more difficult to preserve than that of HeLa cells: identical extraction conditions produce a similar refractile cage, but only occasionally will the cage contain a stabilized MA.

The refractile cage is by far the most stable structure in these extracted mitotic cells. Cells lysed in a wide variety of buffers promoting MT assembly yield labile spindles which readily dissolve, but the cage surrounding the MA invariably persists. Indeed, even under conditions in which we have biased the equilibrium towards a stabilization of the MT component by the addition of MTP at several milligrams per milliliter, no buffers have been found that will solubilize the cage but leave the MA intact.

Lysis of interphase cells using these procedures solubilizes...
FIGURE 1  Fig. 1a shows isolated mitotic HeLa cells detached from monolayer cultures by gentle shaking (DIC optics). Fig. 1b shows mitotic HeLa cells swollen in the hypotonic buffer described in Materials and Methods (polarization optics). Fig. 1 (c–e) shows lysed anaphase and early G1 HeLa cells lysed in the presence of MT protein as described in Materials and Methods, using phase (c), DIC (d), and polarization(e) optics. Cage structures are indicated with arrowheads. Extensive background polymerization of the MT protein has occurred in this preparation. Fig. 1 f shows interphase HeLa cells lysed as described in Materials and Methods (DIC optics). Bar, 10 μm. × 750.

>80% of the cellular protein (7). A nuclear structure remains surrounded by limited amounts of fibrillar cytoplasmic material (Fig. 1f), but no cage analogous to that of mitotic cells is seen. Similar preparations have been reported by several other laboratories (11, 49, 36, 21, 20). The interphase cells in our lysates are not active in nucleating organized birefringent arrays of MTs when lysed with MTP. Similar variation of MT-organizing ability with time in the cell cycle has been reported previously (60, 38, 50, 56).

Electron Microscopy of Detergent-extracted Mitotic Cells

Fig. 2a is a pair of stereo high-voltage electron micrographs (HVEMs) of a prometaphase HeLa cell lysed in assembly buffer containing MTP. We often observe multiple asters in the drug-blocked cells which are always enclosed within a thin fibrous cage. The large asters observed are the result of the elongation of endogenous MTs by the MTP in the lysis solution (26). The fibrous remnant clearly reflects the geometry of the unlysed cell. The asters in the drug-blocked prometaphase cell and the stabilized spindles in the metaphase cells are found within roughly spherical cages (Fig. 2a and b). Anaphase spindles lie in elongated cages (Fig. 3a) while telophase structures show a distinct furrow (Figs. 2c and 3b). The cages from detergent-extracted telophase cells are often collapsed, which may reflect the geometry of the cages at this stage or a tendency of these structures to deform during preparation. In the terminal phase of cell division, the two sister cell ghosts are connected by an intercellular bridge with a distinct midbody (Fig. 3c). At this stage, the cage has divided into two, one-half almost surrounding each daughter nucleus. The cages thus appear to be elastic and can be deformed in concert with the kinetic activities of the mitotic spindle and cleavage furrow.

Scanning micrographs of lysed mitotic cells confirm the three-dimensional characteristics of these fibrous cages (data not shown). Interphase cells lysed and prepared for microscopy by similar methods exhibit a distinct nucleus with attached fibrous protein which is often found as a cap on the nucleus (Fig. 3d) or as a maze of fibers surrounding the nucleus.

Examination of the cages at higher magnification reveals a heterogeneous population of fibers (Fig. 4). Many fields contain arrays of fibers showing an approximately uniform diameter of 8–11 nm characteristic of intermediate filaments (31, 24, 46). Other fibers, however, appear beaded with diameters ranging from 10 to 30 nm. This nonuniform appearance is not presently understood, but it may result from variable amounts of cytoplasm adhering to the fibers during lysis and fixation. In addition to the MTs extending from the mitotic centers, short segments of MTs are observed extending from other regions of the lysed structures in disorganized arrays. These MTs are most likely the result of nonspecific nucleation of the MTP used in the lysis solutions by the material left in the extracted cells.

Electron micrographs of lysed mitotic CHO cells reveal a similar filamentous component in the detergent-extracted mitotic ghosts (Fig. 5a). The cages are composed primarily of beaded fibers although large numbers of disorganized MTs are present. The CHO cages appear to have a lower density of fibers than those prepared from HeLa cells and do not maintain the distinct geometry of the mitotic cell after lysis. Nonetheless, scanning electron micrographs confirm the roughly spherical shape of the mitotic ghosts (Fig. 5b).

We have studied the structure of the isolated cages with thin sections and electron microscopy, but the residue of material left after detergent extraction is so sparse that the images are not useful in comparison with the HVEMs presented in Figs. 2 and 3. Thin sections of unlysed and unswollen cells are,
FIGURE 2  Three pairs of stereo micrographs of lysed mitotic HeLa cells. Mitotic cells were lysed in the presence of MTP and processed for high-voltage electron microscopy as described in Materials and Methods. All pairs were tilted by 3° about a vertical axis between the taking of the two pictures. Fig. 2a is a pair of micrographs of lysed drug-blocked cells. Fig. 2b is of a lysed metaphase cell. Fig. 2c is of a lysed telophase cell. The midbody appears to have formed in this cell before the cleavage furrow completed cytokinesis. The cage structures are indicated with arrowheads. x 3,000.
FIGURE 3 HVEMs of lysed mitotic and interphase HeLa cells. Cage structures are indicated with arrowheads. Fig. 3 a shows an anaphase cell with the hint of a forming cleavage furrow. Fig. 3 b shows a telophase cell with a distinct cleavage furrow. In this preparation the organized MTs in the ghost were lost. Fig. 3 c shows a pair of early G1 cells connected by an intercellular bridge with a distinct midbody ring. Fig. 3 d shows a pair of later G1 daughter cells with distinct nuclei and a cap of filamentous material. The remnant of the midbody is attached to one daughter cell (arrow). The high concentration of MTP in this preparation nucleated what we interpret as a large number of nonspecific MTs. The daughter cells appear to have some symmetry about the midbody. Bar, 2 μm. × 3,000.

However, informative about the physiological distribution of fibrous elements that appear to constitute the cage. We have focused on CHO cells for this work because HeLa cells contain a variety of intermediate filaments that cannot be distinguished in the electron microscope. The cytoplasm of CHO cells in metaphase contains large bundles of 10-nm filaments situated about midway between the periphery of the spindle and the cell cortex (Fig. 5 c and d). These bundles loosely encircle the spindle and, as such, are likely candidates for components of the cage seen in the lysed cells. The enlarged diameter of the cage relative to the position of the fiber bundles in unlysed cells may be the result of the swelling process that precedes lysis.

Several of the populations of mitotic cells prepared for light and electron microscopy were obtained by accumulating mitotic populations with the use of the readily reversible antimitotubule drug Nocodazole (27, 62). Antimicrotubule drugs have been shown to produce alterations in cell architecture which, in addition to the depolymerization of MTs, include a rearrangement of intermediate filaments and a disorganization of cellular organelles (14, 24, 31). However, mitotic cells lysed without exposure to Nocodazole display insoluble filamentous cages that appear identical to those collected with the use of low concentrations of Nocodazole (compare Figs. 2 a, 3 b and Fig. 3 a, c). It is therefore unlikely that structures observed here are the result of the synchronization procedures employing 0.04 μg/ml of Nocodazole.

Polypeptide Composition of the Fibrous Cage

The electrophoretic profiles of the detergent-extracted mitotic and interphase HeLa cells are similar and reveal several
major polypeptides which range in mol wt from 43,000 to 60,000 daltons (Fig. 6). There is also a high molecular weight polypeptide of ~260,000 daltons. The profiles of lysed CHO cells are less complex than those of HeLa cells and contain major polypeptides of 58,000 and ~260,000 mol wt. Our morphological studies reveal that the MA is missing from the cell ghosts that have been through two cycles of detergent extraction in the absence of exogenous tubulin (data not shown). The extraction of the spindle is reflected in the absence of major tubulin polypeptides in the isolated ghosts. Comparison of the major polypeptides present in the detergent-extracted cages and those of whole-cell extracts suggests that the proteins of the cage are among the most abundant proteins of the cell in both HeLa and CHO cells.

Immunological Characterization of the Fibrous Cage from Mitotic Cells

In an effort to identify some protein components of the fibrous cage, two different antisera directed against 10-nm filaments from cultured cells were obtained from other investigators. Antisera prepared against intermediate filament proteins from hamster cells were the generous gifts of Robert Goldman (53) and of Richard Hynes (30). Both sera were used to stain electrophoretic profiles of the detergent-extracted mitotic cages according to the procedure of Burridge (11) (Fig. 7). The serum from Hynes' laboratory binds to the major 58,000-dalton polypeptide from mitotic HeLa and CHO cells. This antiserum was raised against a 58,000-dalton polypeptide from hamster fibroblasts that has been implicated as the subunit of the intermediate filaments by several laboratories and has been named "vimentin" (18). By the same assay, the antiserum of Starger et al. (53), which was raised using intact intermediate filaments as an antigen, recognizes the high 260,000-dalton polypeptide present in both the HeLa and CHO preparations.

Isolated cages prepared from drug-blocked mitotic HeLa and CHO cells stain with both sera (Fig. 8). The fluorescent structures appear as spherical cages with occasional deformations of the surface. No individual filaments can be distinguished in our preparations, probably because of the high density of the fibers in these structures.

DISCUSSION

Stabilized mitotic centers or spindles, as isolated here from HeLa cells, are surrounded by a fibrous cage that is missing or greatly reduced in similarly treated interphase cells. Apparently, the cage forms as the cell enters mitosis. The cage changes shape as the cell proceeds through mitosis to accommodate the elongating spindle, and during cytokinesis it is partitioned into the two daughter structures. Lysed mitotic CHO cells reveal similar fibrous cages, but only occasionally do they contain a stabilized spindle. When visualized in the electron microscope, the cages isolated from HeLa and CHO cells consist of heterogeneous fibrous material that includes fibers 10 nm in diameter.

Analysis of the fibrous cages by SDS gel electrophoresis shows that they are chemically complex, but compared with cell sap they are greatly enriched for polypeptides with mobilities characteristic of intermediate filaments. The rather heterogeneous morphology of the cage fraction is consistent with the biochemical complexity that they display. Our serological studies of the cages from HeLa and CHO cells do, nonetheless, confirm that they contain intermediate (10-nm) filaments of the vimentin type. In HeLa cells, which are of epithelial origin, the cages are particularly well developed and may also contain intermediate filaments of the keratin type.
Intermediate filaments, defined as unbranched cytoplasmic fibers with a diameter of ~10 nm, have been described in a variety of cells. Biochemical and immunological characterizations of these fibers in several laboratories have led to the conclusion that they occur in at least five distinct classes. Muscle cells, neurons and glial cells all contain filaments that appear 10 nm in diameter (5, 8, 13, 31, 35, 44, 45, 61). The fibers of these cells are immunologically distinct, even comparing cells from the same organism. Cells of epidermal origin contain insoluble filaments 6–8 nm in diameter called tonofilaments (17, 19, 21, 32, 54). These fibers pervade the cytoplasm and are found in several differentiated states. The constituent polypeptides of these fibers have been identified as keratins, and they include a family of at least six polypeptides with mol
The planar geometry may result from the flat shape of these endothelial cells which differs from the rounded shape of the mitotic HeLa and CHO cells we have studied. Blose (6) has demonstrated that the ring of intermediate filaments is not cleaved if the cleavage furrow is inhibited with cytochalasin. Similarly, the cage of intermediate filament in HeLa cells remains spherical when cells are arrested at the onset of mitosis, suggesting that the intermediate filaments are passively deformed by the kinetic activities of the mitotic spindle and cleavage furrow. Thin-section electron micrographs of intact mitotic CHO cells in which the intermediate filaments are readily identifiable demonstrate that the filaments appear in the cytoplasm around the perimeter of the cell, however, they do not appear to interact with the cell membrane.

HeLa cells have recently been identified as containing intermediate filaments immunologically related to the keratin-containing tonofilaments in addition to the vimentin filaments (20). Similar filaments were shown to be lacking in fibroblasts such as the CHO cells. These fibers were demonstrated to be insoluble in detergent and also to collect around the MA to form a “basket” during cell division. It is highly likely that our isolated HeLa cell mitotic cages include prekeratin-containing intermediate filaments as well as vimentin-containing fila-

wt from 40,000 to 60,000. Antibody produced against soluble keratin (prekeratin) will decorate filamentous arrays in cultured cells of epithelial origin, including HeLa cells, but this probe does not stain the intermediate filaments of fibroblasts (17, 19, 55). A fifth distinct system of intermediate filaments forms networks in the cytoplasm of a number of interphase cells (6, 18, 25, 30, 52). These networks appear to originate from a juxtanuclear position and collapse around the nucleus to form perinuclear whorls when interphase microtubules are depolymerized by antimicrotubule reagents (15, 24). A 58,000-dalton polypeptide has been identified as a subunit of these filaments by several laboratories (18, 25, 30) and has been named vimentin by Franke et al. (18). Vimentin fibers are insoluble in the presence of nonionic detergents at both low and high ionic strength (4, 30, 52, 49). They have been identified as a major component of cytoskeletal preparations from a wide variety of cell types (4, 30, 19).

The immunofluorescent staining of isolated mitotic cages from HeLa and CHO cells and the radioimmune recognition of appropriate polypeptides in the electrophoretic profiles of the isolated cages by antisera directed against the vimentin fibers demonstrates that this class of intermediate filaments is present in our detergent-extracted preparations from both epithelial cells and fibroblasts. The localization of intermediate filaments of this type in the isolated mitotic cages even after extraction of the spindle is not surprising, considering the large number of reports that intermediate filaments remain insoluble after detergent extraction. The metaphase organization of the filaments in a cage that surrounds the MA suggests that the interphase arrays collapse around the mitotic spindle as the cell enters into mitosis.

Previous immunofluorescence studies have described vimen-
ments. These tonofilaments may account for the polypeptides of ~40,000–55,000 daltons present in the lysed HeLa cells and not present in the lysed CHO cells (11). This second class of intermediate filaments may also contribute to the greater stability of the lysed HeLa cells compared to the lysed CHO cells.

Several laboratories have proposed that the distribution of vimentin filaments is closely tied to that of the MTs, a suggestion based both on the similarity of their spatial distribution in many cells and on the rearrangement of intermediate filaments that occurs when MTs are depolymerized by antimicrotubule drugs (19, 24, 30). The rearrangement of the intermediate filaments into a cage surrounding the MA may be an analogous result of the physiological depolymerization of the cytoplasmic MTs as the mitotic spindle assembles. It is noteworthy, however, that intermediate filaments are conspicuously absent from the spindle itself, except for the telophase interzone (40) where they are presumably co-mingled with the spindle tubules by the action of the cleavage furrow. It seems unlikely, therefore, that mitotic MTs interact in any direct way with intermediate filaments.

No specific function for the intermediate filaments in cultured cells has yet been identified, although many investigators have suggested that they play a structural role in the cytoplasm. The filaments may be associated with the well-known exclusion of cytoplasmic organelles from the area of the mitotic spindle (22), or they may form a cage because there is no place else for them to go. More speculative but intriguing is the possibility that the retention of these filaments and whatever additional material is associated with them in the absence of cytoplasmic MTs or microfilament bundles allows them to serve a skeletal role in mitotic cells. When the microfilament bundles, the cytoplasmic MTs, and the nuclear envelope break down in prophase, much of the interphase organization of a cell is lost. Nonetheless, cells reestablish their interphase morphology shortly after division. Perhaps the mitotic cage serves as a framework that helps redefine the cytoplasmic geometry after cytokinesis.

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