Nonrandom Chromosome Segregation in
Neocurtilla (Gryllotalpa) hexadactyla:
An Ultrastructural Study

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ABSTRACT During meiosis I in males of the mole cricket Neocurtilla (Gryllotalpa) hexadactyla, the univalent X₁ chromosome and the heteromorphic X₂Y chromosome pair segregate nonrandomly; the X₁ and X₂ chromosomes move to the same pole in anaphase. By means of ultrastructural analysis of serial sections of cells in several stages of meiosis I, metaphase of meiosis II, and mitosis, we found that the kinetochore region of two of the three nonrandomly segregating chromosomes differ from autosomal kinetochores only during meiosis I. The distinction is most pronounced at metaphase I when massive aggregates of electron-dense substance mark the kinetochores of X₁ and Y chromosomes. The lateral position of the kinetochores of X₁ and Y chromosomes and the association of these chromosomes with microtubules running toward both poles are also characteristic of meiosis I and further distinguish X₁ and Y from the autosomes. Nonrandomly segregating chromosomes are typically positioned within the spindle so that the kinetochoric sides of the X₂Y pair and the X₁ chromosome are both turned toward the same interpolar spindle axis. This spatial relationship may be a result of a linkage of X₁ and Y chromosomes lying in opposite half spindles via a small bundle of microtubules that runs between their unusual kinetochores. Thus, nonrandom segregation in Neocurtilla hexadactyla involves a unique modification at the kinetochores of particular chromosomes, which presumably affects the manner in which these chromosomes are integrated within the spindle.

The model for chromosome behavior during meiosis in all higher organisms is based upon Carothers’ (3) careful analysis of the behavior of heteromorphic chromosome pairs and a univalent sex chromosome in grasshopper spermatocytes. Each heteromorph was found to assort completely randomly with respect to every other heteromorphic pair and with respect to the single X chromosome. These results are in accord with the chromosomal theory of inheritance as first stated by Sutton (13); pairs of homologous chromosomes are arranged on the spindle at meiosis I without regard to one another and, therefore, assort independently, just as do unlinked genes in crosses.

Ironically, observations that contradict the generality of this model were reported by Payne (10, 11) at about the same time as the work of Carothers. In spermatocytes of the Northern mole cricket, a relative of the grasshopper, Payne recognized a heteromorphic chromosome pair formed by association of two chromosomes of markedly different sizes and found that the members of this pair segregate nonrandomly with respect to a univalent X chromosome; the larger member of the heteromorph always moves to the same spindle pole as the X chromosome at anaphase I. This nonrandom behavior was observed in every cell, despite the fact that Payne could discover no physical connection between the chromosomes involved. The accuracy of Payne’s observations has been confined by White (15) who studied fixed cells and by Camenzind and Nicklas (2) who observed living spermatocytes.

The truly remarkable nature of nonrandom segregation in the mole cricket was revealed by the study of living cells (2). Micromanipulation of the chromosomes involved supported the opinion of earlier workers that there is no physical connection between those chromosomes. Moreover, when the inappropriate orientation (X chromosome and larger member of the heteromorph oriented to opposite poles) was induced with the micromanipulation needle, the appropriate orientation was
actively restored; the sex chromosome usually reoriented sponta- neously so as to ensure its passage to the same spindle pole as the large member of the heteromorph.

Preferential distribution of unconnected chromosomes is not unique to the Northern mole cricket; it has been reported for a number of unrelated organisms (for review see references 7 and 8). However, fundamental differences between the various examples suggest that no single mechanism governs all cases of nonrandom chromosome behavior. In cray flies, for example, physically separate chromosomes that assume similar orientation on the spindle regularly move toward opposite spindle poles. It is, therefore, necessary to explain how poleward movement of one chromosome can induce oppositely directed movement of another chromosome (5). For the mole cricket, on the other hand, the fact that chromosomes orient on the spindle in nonrandom manner requires that we discover how orientation of the X chromosome can be influenced by the orientation of the heteromorph (2).

Camenzind and Nicklas (2) suggested that the structural organization of the spindle could dictate nonrandom orientation of particular chromosomes if there exist specific differences between the spindle fibers of non-randomly segregating chromosomes and those of the remaining chromosomes of the complement. Our study was undertaken to discover if there are ultrastructural indications of such differences. We have found that: an electron-dense material is associated with the kinetochores of the X chromosome and of the smaller chromosome of the heteromorphic pair and that microtubules are arrayed about these particular kinetochores in an unusual fashion. We discuss the manner in which these structural peculiarities might be related to nonrandom chromosome behavior.

MATERIALS AND METHODS

Living specimens of Neocurtilla (Gryllotalpa) hexadactyla Perty (4) were collected near Grassy Pond in the Sandhills Wildlife Management Area in the vicinity of Southern Pines, N. C.

All of the preparative procedures used in this study have been described previously. To select cells in the desired stages of spermatogenesis, living cultures of testicular tissue were prepared for light microscopic observations as described by Nicklas et al. (9). Cells were observed on an inverted phase contrast microscope and photographed by time-lapse cinemicrography. During observation and recording, fixation of selected cells was initiated by microinjection of fixative into the vicinity of the target cell and was completed by immersion of the coverslip bearing the attached target cell in a large volume of fixative. This routine of microinjection followed by "macrofixation" and subsequent steps of preparation for electron microscopy were essentially those of Nicklas et al. (9). However, the composition of fixatives, buffers, etc. and the schedule of fixation differed. Our conditions were as follows: (a) Microfixation: 6% glutaraldehyde in PIPES-NaCl buffer (100 mM PIPES [Sigma Chemical Co., St. Louis, Mo., 0.3% NaCl, pH 6.8-6.9) for 4-5 min. (b) Macrofixation: 3% glutaraldehyde in PIPES-NaCl buffer for 30 min. (c) Rinses: PIPES-NaCl buffer; two changes of buffer at 10 min intervals followed by overnight storage in buffer at 4°C. (d) Second Fixation: 2% osmium tetroxide in PIPES-NaCl buffer for 0.75-1 hr. (e) Rinses: distilled water; two changes at 10 min intervals. (f) Uranyl acetate treatment: 0.5% uranyl acetate in veronal-acetate buffer for 0.75-1 hr (12).

Fixatives were prepared immediately before use. As an assurance of the quality of the glutaraldehyde used, fixatives were prepared from a 70% stock solution stored in sealed ampoules (Ladd Research Industries, Inc., Burlington, Vt.); only those lots having an absorption ratio A230/ A360 < 0.2 were used (1).

Serial sections through entire cells were examined in a Siemens 101 electron microscope, and micrographs of each section including nucleus or spindle were prepared using Kodak 70-mm roll film (Kodak #2772). The overall analysis was based upon the study of serial negatives prepared at original magnifications ranging from × 2,300 to × 3,900 and viewed using the × 14 or × 31 additional magnification provided by an aerial roll film viewer (Hoppmann Corp., Spring-
The First Meiotic Division

The features that distinguish nonrandomly segregating chromosomes in *N. hexadactyla* are most evident during the later stages of the first meiotic division. Therefore, the description of metaphase and anaphase cells precedes that of the earlier prometaphase and prophase cells.

KINETOCHORE STRUCTURE AND SPINDLE ORGANIZATION IN METAPHASE CELLS: Four metaphase cells were examined in serial sections; they are designated cells A–D. In each, massive aggregates of extremely electron-dense material are located on the X1 and Y chromosomes, as illustrated in cell A (Figs. 2, 3, and 4). These aggregates are of indeterminate structure but are characterized by an elongate profile and by the irregular, labyrinthine relationship among the patches of electron-dense material composing them. The electron-dense material marks the X1 and Y kinetochore regions — only here are microtubules associated with these chromosomes.

The remaining kinetochores in metaphase cells, those of the autosomes (Fig. 5) and of the X2 chromosome, are discrete fibrillar masses less dense than the chromosomes; they are found at the poleward extreme of the chromosome, sometimes in a cuplike depression. Those that lie near the unusual X1 or Y kinetochore regions may be marked by a single electron-opaque patch, as is the case for one autosomal kinetochore and the X2 kinetochore in cell A and for two autosomal kinetochores in cell B (Fig. 6). In general, though, autosomal and X2 kinetochores are free of such electron-dense material; none appears in cells C and D (because a number of sections of cell D were lost, the kinetochores of only nine of its 10 autosomal pairs were examined). During other division stages (see descriptions of meiosis II and mitosis), X1 and Y kinetochores are discrete fibrillar structures indistinguishable from the autosomal and X2 kinetochores just described. Therefore, we attempted to recognize such structure within the X1 and Y kinetochore regions during metaphase of meiosis I. We conclude that if X1 and Y kinetochores per se are similar to autosomal and X2 kinetochores, this is obscured by the aggregate of electron-dense material in the kinetochore region.

The X1 and Y chromosomes at metaphase I differ further from autosomes and the X2 chromosome in the position of the kinetochore on the chromosome and in their associations with microtubules. The autosomes and the X2 chromosome display exactly the sort of kinetochore-microtubule-pole relationship which is characteristic of organisms with localized kinetochores (6) (Figs. 2, 5, and 6); kinetochores on oriented metaphase chromosomes are situated so that they face directly toward one centriolar pole and microtubules that emanate from the kinetochore run directly toward that pole. In contrast, the kinetochore regions of X1 and Y chromosomes (Figs. 2, 3, and 4) are not pointed directly toward only one pole. Instead, the elongate electron-dense kinetochore region is lateral to the body of the chromosome; it is, thus, aligned parallel to the interpolar axis and cannot be considered to face only one of the two poles. Furthermore, the microtubules that impinge on the electron-dense aggregate are directed not toward the nearer spindle pole exclusively, but microtubule profiles are arrayed about both ends of the aggregate as well as in the interstices between the electron-dense patches (Figs. 3 and 4; see also Fig. 8). Three-dimensional reconstructions prepared so as to include all microtubule profiles between the chromosome and the near pole demonstrate that a microtubule bundle proceeds from the kinetochore region poleward. Presumably, this bundle is involved in the chromosome's orientation to that pole. It is possible that some or all of the microtubules that compose that bundle actually pass through the electron-dense aggregate and emerge at its opposite end. This would account for the microtubules found at the two ends of the aggregate and in among the electron-dense patches (Figs. 3 and 4). Alternatively, separate bundles of microtubules may associate with the opposite ends of the kinetochore region. We could not evaluate these alternatives because the density of the electron-opaque material in the kinetochore region makes it impossible to recognize any microtubules which may run through it.

At metaphase, nonrandomly segregating chromosomes are positioned in the spindle in a particular manner. Most obvious, of course, is the characteristic orientation of the physically unlinked X1 and X2 chromosomes to one pole, whereas the Y chromosome (which is paired with the X2 chromosome) is oriented to the opposite pole. Also, as illustrated in earlier publications (e.g., references 2 and 10), the X2Y pair is shifted off the equator so that the Y chromosome is closer to a pole than is X2 (Figs. 1 and 2). More subtle spatial relationships are also discernible. Three-dimensional reconstructions of all four metaphase cells demonstrate that the X2Y chromosome pair is centered within the spindle volume so that it lies almost exactly on a line drawn between the centrioles at opposite spindle poles. Moreover, in three of the four cells (cells A, B, and D), the relationship between the central X2Y pair and the more peripheral univalent X1 chromosome is as illustrated in Figs. 2 and 7 a and b. In Fig. 2, because both X2 and Y chromosomes are included in the same section, it is plain that the kinetochoric sides of these chromosomes are aligned more or less along the same spindle axis. In cells B and D, this relationship is not so immediately apparent because X1 and Y appear in different section planes. However, the reconstructions show that the chromosomes are situated exactly as in cell A. Cell C is an exception in this regard; the chromosomes in question are 90° out of the arrangement that would bring their kinetochoric sides onto the same axis (Fig. 7 c and d).

We looked for structural features that might account for a definite spatial relationship between X1 and Y chromosomes. For both cells A and B, the reconstructions suggest that a few microtubules link the kinetochore regions in question. The clearest example is presented by cell A, in which microtubule profiles running in a very different direction from the majority of spindle microtubules trace a direct line between X1 and Y kinetochore regions (Fig. 8). In cell B, also, a small bundle of microtubules extends between these kinetochore regions. This was not the case in cell C where the kinetochoric sides of X1 and Y chromosomes are not on the same axis. Similar examination of cell D was impossible because of section losses.

INAPPROPRIATE CHROMOSOME ORIENTATION: We examined a single cell showing a well-defined metaphase plate but with the nonrandomly segregating chromosomes in inappropriate orientation, i.e., with X1 oriented to the same pole as the Y chromosome (Fig. 9 a and b). Again, the kinetochores of chromosomes X1 and Y are marked by patchy electron-dense aggregates and these face a common axis, as in the typical metaphase cells already described. However, the X1 kinetochore region is distinctly different in this cell; it is not elongated along the interpolar axis and all of the associated microtubules run directly toward the nearest pole (Fig. 9 c). The only feature of the X2Y chromosome pair worthy of note is its somewhat eccentric placement in the spindle so that it does not lie on the central interpolar axis as it does in more typical metaphase
FIGURE 3  Kinetochore region of the X₁ chromosome in metaphase of meiosis I, cell A. The chromosome is shown in the same orientation as in Fig. 2; as determined from serial sections, the nearer pole is above and to the left of the area included in these micrographs. The patches of electron-dense material found in the kinetochore region (kr) are arranged in an irregular mass; they appear in 14 serial sections and, thus, constitute a sizeable elongate aggregate (sections a, 4; b, 6; c, 7; and d, 9 of the series are shown). Microtubules (arrows) are found at the end of the kinetochore region closer to a pole as well as at its opposite end and in among the electron-dense patches. Bar, 1 μm. X 24,000.

ANAPHASE: One early anaphase cell was studied in detail. At the time of fixation, the X₂ and Y chromosomes had already disjoined and moved a significant distance poleward, but only two of the 10 autosome pairs were clearly disjunct. The patchy electron-dense substructure of X₁ and Y kinetochore regions, the array of microtubules at both ends of those electron-dense aggregates, the position of X₂ and Y chromosomes on the central spindle axis, and turning of the kinetochoric sides of the X₁ and Y chromosomes toward a common axis are just as they are in typical metaphase cells. The spindle region between X₁ and Y kinetochore regions contains no microtubules with the directionality which would be expected if microtubules formed a link between these chromosomes. All autosomal kinetochores are free of electron-dense material.

The only cell fixed at a later stage in anaphase was, unfortunately, not very well preserved (poor visibility of microtubules, chromatin fusion). Nevertheless, it is worth noting that single small electron-dense patches appear on the kinetochores of several chromosomes but that no massive aggregations of such material occur; thus, X₁ and Y chromosomes are not distinctive in this cell.

PROMETAPHASE: In the three prometaphase cells examined (designated cells E, F, and G), the X₁ and Y chromosomes are not recognizable from inspection of single sections because the kinetochore regions of all chromosomes have a similar appearance. These resemble the peculiar X₁ and Y kinetochore regions formed a link between these chromosomes. All autosomal kinetochores are free of electron-dense material.

FIGURE 2  Metaphase of the first meiotic division, cell A. (a) In this phase micrograph taken immediately after microfixation, the X₁ univalent (X₁), the heteromorphic bivalent formed by association of a large (X₂) and a small (Y) chromosome, and autosomes (A) are distinguishable. As is typical for this stage of meiosis, X₁ has already approached one pole and the X₂ chromosome is oriented to the same pole. Note that the X₂Y pair is shifted off the spindle equator so that the Y chromosome lies closer to a pole than does X₂. Bar, 10 μm. X 2,100. (b) This section includes the kinetochore regions (arrows) of the four chromosomes identified in a. The kinetochore regions of X₁ and Y chromosomes appear as elongate electron-dense masses lateral to the body of the chromosomes. Note that the kinetochoric sides of these two chromosomes face toward a common axis—a line from pole to pole lying between the X₂Y pair and the X₁ chromosome. The kinetochores of the X₂ chromosome and of an autosome are difficult to discern because of their moderate electron density and the low magnification of this micrograph. See Fig. 5 for a micrograph of the autosomal kinetochore at higher magnification. Bar, 1 μm. X 11,200.
FIGURE 4 Kinetochore region of the Y chromosome in metaphase of meiosis I, cell A. The chromosome is oriented as in Fig. 2; the nearer pole is directly below the area included in these micrographs. The Y kinetochore region (kr) is similar to the X, kinetochore region shown in Fig. 3 in its electron density, complicated elongate structure, and its association with microtubules (arrows). The kinetochore is massive and continues in a series of 12 sections (sections a, 5; b, 7; c, 8; and d, 10 are shown). Bar, 1 μm. X 24,000.

FIGURE 5 An autosomal kinetochore at metaphase of meiosis I, cell A. This is another section of the autosomal kinetochore indicated in Fig. 2. The kinetochore (k) is a moderately electron-dense, fibrillar mass embedded in a cuplike depression of the chromosome. Microtubules (arrows) run from the kinetochore toward the nearer pole. Bar, 1 μm. X 38,900.

in metaphase cells insofar as they have a mottled or patchy character, but they are far less electron-opaque (Figs. 10 and 11). Because cell G is in a relatively advanced stage of pro-metaphase (all chromosomes in approximately equatorial position, six of 11 chromosome pairs bipolarly oriented) and well-defined bundles of microtubules are associated with the majority of kinetochores, it is clear that the observed kinetochore structure is not a feature restricted to earliest prometaphase or to kinetochores which are not associated with microtubules.

Two-dimensional reconstructions permit identification of the univalent X, chromosome and the X2Y chromosome pair (Fig. 12) in all three cells. By this means, we can see that the X, and Y kinetochore regions are more massive than those of other chromosomes and that they are elongate and lateral to the body of the chromosome just as in metaphase. Also, as in metaphase, microtubules are associated with both ends of the kinetochore regions; this was evident for both X, (Fig. 10) and Y chromosomes in cell E and for the X, chromosome in cell F.

Because prometaphase is the stage during which chromosomes assume appropriate orientation on the spindle, association between X, and Y chromosomes during this stage might dictate their nonrandom behavior at later stages. In this context, we note that the X, univalent and the X2Y pair are not obviously connected in any of the three cells but that the kinetochores of X, and Y chromosomes are turned toward each other in two of the three cells (Fig. 12).
FIGURE 6 Kinetochore of an autosome and the Y kinetochore region in metaphase of meiosis I, cell B. A patch of electron-dense material (arrow) marks the kinetochore of an autosome (A) immediately adjacent to the massive electron-dense kinetochore region (kr) of the Y chromosomes (Y). The autosome-associated electron-dense material is confined to the single patch illustrated and appears in only three consecutive sections. Bar, 1 μm. x 23,800.

FIGURE 7 The relationship between nonrandomly segregating chromosomes as determined from three-dimensional reconstructions of the spindle. (a) In metaphase cells A, B, and D, the central X2Y pair (X2Y) and the more peripheral X1 chromosome (X1) lie in the spindle with their kinetochore regions (arrows) aligned along a common spindle axis; if the spindle were viewed from one pole, as in b, X1 and Y kinetochore would appear to face each other across one radius of the spindle. (c) In metaphase cell C, the X1 and Y chromosomes have a different relationship; if the X2Y pair is viewed so that the Y kinetochore region is on the side of the chromosome facing the reader, a nonkinetochoric side of the chromosome lies on a common axis with the X1 kinetochore region. When viewed from the pole as in d, the X1 kinetochore region appears to face a side of the X2Y pair ~90° away from the Y kinetochore region.

PROPHASE: We examined one cell in diakinesis, the final prophase stage before nuclear envelope breakdown signals the onset of prometaphase. Kinetochores are not recognizable on the loosely condensed chromosomes in this nucleus (Fig. 13). In two-dimensional reconstruction, individual chromosomes are easily recognized. Because 12 chromosomes (10 autosome pairs, the X2Y pair, and chromosome X1) appear as discrete entities in the reconstruction, there is evidently no close association between X1 and the X2Y pair such as might promote their coordinated orientation in the ensuing prometaphase and nonrandom segregation at anaphase.

The Second Meiotic Division

The nonrandom behavior of X1, X2, and Y chromosomes in the first meiotic division assures that cells in the second meiotic division carry either the X1 or the Y chromosome, never both. Therefore, if the differentiation of the kinetochores of these particular chromosomes persists in the second division, we can
mitotic. Bar, 1 μm. X 10,900. (c) Another section of the kinetochore region shown in b. It is clear that microtubules (m) run between the kinetochore region and the near pole (p). In the serial sections, all of the microtubules associate with this kinetochore material. The kinetochores of six of the eight autosomes examined are so marked, four of them at each of their two kinetochores, whereas two others have a patch at only one kinetochore.

**Mitosis**

Kinetochore structure was examined in spermatogonial mitosis to discover whether X1 and Y chromosomes manifest peculiar kinetochore structure even when their behavior is not exceptional. For one spermatogonial metaphase and one spermatogonial anaphase cell, all kinetochores were examined in serial sections. All resemble the undifferentiated autosomal kinetochores described for the first meiotic metaphase and anaphase; no electron-dense patches appear on any of them. Thus, X1 and Y kinetochores are not distinctive during mitosis.

**DISCUSSION**

Any hypothesis formulated to explain nonrandom segregation in *N. hexadactyla* must take into account the information derived from experimental study of living cells (2) together with the following structural facts established by our study.

First, distinctive kinetochore regions are found on only two of the nonrandomly segregating chromosomes, the unlinked X1 and Y chromosomes, which orient to opposite poles.

Second, the kinetochore regions of X1 and Y chromosomes become progressively more distinctive during meiosis I. In mitosis and meiosis II, X1 and Y kinetochores are as small as the kinetochores of all other chromosomes and they interact with microtubules in the same way as do other kinetochores. In prometaphase of meiosis I, changes in X1 and Y kinetochore regions are already evident. They are appreciably larger than those numerous microtubules at the nonpolar end of the X1 kinetochore region which lie at angles precluding their inclusion in a microtubule bundle running between the two chromosomes. p, Pole. Bar, 1 μm.
FIGURE 10. The X₁ kinetochore region in prometaphase of meiosis I, cell E. The presence of only one kinetochore region (kr) on this chromosome identifies it as the X₁ univalent (X₁) (see also Fig. 12). The kinetochore region is similar to X₁ kinetochore regions in metaphase cells in its inhomogeneous, patchy substructure, elongate profile, position lateral to the body of the chromosome and association with microtubules (arrows) at opposite ends. Yet, it differs from metaphase X₁ kinetochore regions in being no more electron-dense than the chromosome. Bar, 1 μm. X 38,000.

FIGURE 11. The kinetochore of an autosome (A) in prometaphase of meiosis I, cell G. This autosomal kinetochore (k), like all autosomal kinetochores in the prometaphase cells examined, is similar to the prometaphase kinetochore regions of X₁ and Y chromosomes (cf. Fig. 10) in its inhomogeneous, patchy appearance. Nevertheless, the position of the kinetochore at a chromosome end, the presence of a second kinetochore at the opposite end of the bivalent, and the microtubules (arrow), which clearly terminate at the kinetochore, all permit an unambiguous distinction between autosomal and X₁ or Y kinetochore regions. Bar, 1 μm. X 28,500.

FIGURE 12. Chromosomes in prometaphase of meiosis I, cells E, F, and G. For the sake of clarity, only chromosome outlines and the kinetochore regions (dotted lines) of X₁ and Y chromosomes are included in these drawings, which were traced from two-dimensional reconstructions of complete spindle regions. In each cell, the X₂Y (X₂;Y) pair is unambiguously identifiable on the basis of its drumsticklike shape. The X₁ chromosome (X₁) is identified as the only chromosome in the cell bearing but one elongate, lateral kinetochore region, as expected for this univalent. The X₁ and Y kinetochore regions are both elongate and lateral to the body of the chromosome, exactly as in metaphase cells. a, Cell E; b, cell F; c, cell G.

It was thought that pairing between the X₁ univalent and the X₂Y chromosome pair in early meiotic stages might provide a relatively simple device for their nonrandom orientation (15, 16); if present, the pairing could constrain X₁ and Y kineto-
chores to face oppositely so that the sort of explanations offered for bipolar orientation of ordinary meiotic bivalents \(^8^\) would be applicable. The experiments of Camenzind and Nicklas (2) demonstrated that reorientation leading to appropriate segregation proceeds without pairing, but the possibility that initial orientation requires pairing remained open. Now, with the knowledge that the \(X_1\) univalent and the \(X_2 Y\) pair are not associated either before (diakinesis) or during (prometaphase) the time chromosomes are establishing their spindle connections, we can dismiss the idea that pairing is ever concerned with the preferential distribution of these chromosomes.

We must instead consider that nonrandom segregation in \(N.\ hexadactyla\) is a function of extraordinary properties of the \(X_1\) and \(Y\) kinetochores and their associated microtubules. The inference that nonrandom segregation is somehow dependent on the unique nature of \(X_1\) and \(Y\) kinetochores is justified by the contrast between various divisions. \(X_1\) and \(Y\) kinetochore regions are grossly different from other kinetochores only during the first meiotic division in males, precisely when preferential orientation and distribution occur, but they are not at all distinctive in the second meiotic division or in mitosis when all chromosomes behave similarly. Although diverse kinetochrome types are known from a variety of organisms \(6\), this mole cricket is the first example in which massive aggregates of electron-dense material distinguish particular kinetochores; thus, although kinetochores like those of the autosomes and the \(X_2\) chromosome are known, the distinctive \(X_1\) and \(Y\) kinetochore regions as observed during the first meiotic division in males are unlike any previously described.

The differentiation of \(X_1\) and \(Y\) kinetochore regions during meiosis I involves increases in size and electron density which presumably occur because those kinetochores associate with an electron-dense substance. The electron-dense material responsible for the conspicuous appearance of \(X_1\) and \(Y\) chromosomes at metaphase I and anaphase I is obviously not an integral part of the chromosome nor is it absolutely specific for these particular kinetochores, as is indicated by the presence of solitary patches of electron dense material on autosomal or \(X_2\) kinetochores lying near the massive electron-dense aggregates on \(X_1\) and \(Y\) chromosomes (metaphase cells A and B) and by the

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**Figure 13** Phosphase of meiosis I. (a) In diakinesis, the chromosomes (1 and 2) are loosely condensed and each is in contact with regions of the nuclear envelope (ne) free of nuclear pores. Nuclear pores (arrows) are concentrated in the remaining chromosome-free areas of the envelope and sizeable flocculent electron-dense masses (m) appear on the cytoplasmic side of the envelope in these areas \((14)\). Bar, 1 \(\mu\text{m} \times 19,400\). (b) In an earlier prophase cell (chromosomes more loosely organized and nucleolus less dispersed than in diakinesis), the association of electron-opaque material with individual pores is clear (arrows). The presence of aggregates of similar electron-dense substance in the cytoplasm of metaphase cells (not illustrated) indicates the persistence of the material in later stages of meiosis. Bar, 1 \(\mu\text{m} \times 36,700\).
occurrence of such patches on numerous chromosomes when massive electron dense aggregates are not present (the late anaphase I cell and the metaphase II cell). Nonetheless, X₁ and Y chromosomes are the only ones ever associated with substantial quantities of electron-dense material, and the electron-dense material appears on both X₁ and Y chromosomes even when they are inappropriately oriented. We can do no more than speculate as to what role, if any, this substance may play in chromosome behavior. An obvious and interesting possibility is that the electron-dense material confers unusual properties on the X₁ and Y chromosomal fibers and that stable spindle structure (hence, stable orientation of X₁ and the X₂Y pair) is possible only when one atypical chromosomal fiber is included in each half spindle (cf. reference 2). Alternatively, the electron-dense material may not directly alter the properties of spindle fibers; instead its presence may be a secondary effect which derives from presently unrecognized special properties of the X₁ and Y chromosomes. In any case, the specificity of association of this material with particular kinetochores does at least signal that those chromosomes have unique properties which produce an affinity between kinetochore and electron-dense material and probably determine the observed pattern of non-random segregation.

The different behavior of autosomes and the nonrandomly segregating chromosomes must depend most directly on the very different chromosome-microtubule interactions observed. The kinetochore microtubules of autosomes and the X₂ chromosome run only toward the near pole. It is obvious that these ordinary chromosomes are integrated in the spindle syntelically; that is, both sister kinetochores on the two chromatids composing a chromosome connect to the same pole via a single chromosomal fiber. In contrast, appropriate orientation of the unlinked X₁ and Y chromosomes is at least correlated with an unusual association of microtubules with not only the end of the kinetochore region closest to a pole but also with the opposite end. This peculiar relationship is the rule for appropriately oriented X₁ and Y chromosomes in metaphase cells and is already evident in prometaphase (X₁ and Y chromosomes in cell E; and X₁ chromosome in cell F). Conceivably, the array of microtubules in the vicinity of the extraordinary X₁ and Y kinetochore regions indicates amphitelic orientation (sister kinetochores connected to opposite poles via microtubules running toward opposite poles). Because meiotic univalents are often amphitetrically oriented (for review see reference 8), this would not be unexpected for the univalent X₁, and in this light it is the Y chromosome, which is anomalous. However, we cannot conclude that either X₁ or Y is in fact amphitetically oriented because we could not establish that separate bundles of microtubules emanate from each end of the kinetochore, with each bundle connected to a different sister kinetochore region. The [See page 272.]}
ochore.

Whereas the microtubule bundles associated with X₁ and Y chromosomes are unmistakably different from the ordinary kinetochore microtubule bundles in the same spindle, it is not clear that the integration of such atypical fibers within the spindle would necessarily constrain the X₁ and Y chromosomes to segregate to opposite poles. The position of X₁ and Y relative to each other may be influenced by the microtubules that we found to run directly between X₁ and Y kinetochores regions lying in opposite half spindles (metaphase cells A and B). If these microtubules constitute a mechanically significant link between X₁ and Y chromosomes, they could certainly cause these chromosomes to lie with their kinetochoric sides facing a common axis, as is typically observed. Whether such linkage also promotes the orientation of X₁ and Y chromosomes to opposite poles is a matter for conjecture. Perhaps, for example, the elongation of microtubules, or sliding between interdigitated microtubules, which extend between X₁ and Y kinetochores increases the distance between those kinetochores so that they must come to lie in opposite half spindles. Thus, it is at least conceivable that long-range interactions between X₁ and Y chromosomes via microtubules provides a structural basis for nonrandom segregation.

In summary, we have suggested that nonrandom segregation in *N. hexadactyla* could, in principle, depend upon a) half-spindle effects—only one chromosome bearing a modified kinetochore can be accommodated in half spindle and upon b) “interzonal” effects—the linkage of modified kinetochores via microtubules that cross the equator influences the relative position of these kinetochores. These are not mutually exclusive possibilities; for example, initial orientation of modified kinetochores to opposite poles may be promoted by half-spindle effects, whereas stabilization of that orientation is dependent upon interzonal effects. This idea is consistent with experimental data which demonstrate that the X₁ chromosome does not achieve stable orientation when the X₁Y pair is inappropriately oriented or has been removed from the cell (2).

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