Human $\beta_2$-microglobulin is a Substrate of Tissue Transglutaminase: Polymerization in Solution and on the Cell Surface

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ABSTRACT Incubation of purified human $\beta_2$-microglobulin ($\beta_2$-m) with tissue transglutaminase (Tgase) resulted in the formation of high molecular weight polymers revealed by sodium dodecyl sulfate polyacrylamide gel electrophoresis. In the presence of 30 mM $[^{14}C]$methylamine, the polymer formation was prevented, but incorporation of methylamine into $\beta_2$-m (equal to 1 methylamine per 1 molecule) could be observed. From the shedding of peripheral blood mononuclear cells occurring in the presence of Tgase, it is apparent that anti-$\beta_2$-m immunoadsorbent removed, in addition to human leukocyte antigen (HLA) and $\beta_2$-m, some other proteins. The enzyme could incorporate $[^{14}C]$methylamine into $\beta_2$-m of the shedding cells. On addition of rabbit anti-human $\beta_2$-m antibody, followed by fluoresceine-labeled goat anti-rabbit IgG antibody to human mononuclear blood cells, the otherwise homogeneous distribution of fluorescence turned into spots and patches on cells previously incubated with Tgase or Ca$^{2+}$-ionophore A23187.

Materials and Methods

Human $\beta_2$-microglobulin ($\beta_2$-m) isolated from culture media of human lymphoid cell lines was a kind gift of Dr. N. Tanigaki (Roswell Park Memorial Institute, Buffalo, New York). Guinea pig liver Tgase, which was purified according to Connellan et al. (5), exhibited 92% ± 8% of the reported specific activity upon assay by hydroxamate formation with benzyloxycarbonyl (2)-$\gamma$-glutaminylglycine. Active-site-inhibited Tgase prepared as previously described (15) showed no activity. An IgG fraction of monospecific rabbit anti-$\beta_2$-m-antiserum was obtained from Dako Immunoglobulins Ltd. (Copenhagen, Denmark) and diluted to 700 µg/ml with Parker medium TC-199 just before use. Fluoresceine-labeled IgG of goat antiserum against rabbit IgG (Hyland Diagnostics Div., Travenol Laboratories, Inc., Costa Mesa, Calif.) was used as a second antibody at a concentration of 500 µg/ml. Ca$^{2+}$-ionophore A23187, a gift of Eli Lilly and Company (Indianapolis, Indiana), was dissolved in dimethylsulfoxide at 1 mg/ml and stored at 4°C. $[^{14}C]$Methylamine (specific activity 40 mCi/mmol) was purchased from New England Nuclear (Boston, Massachusetts). Protein standards for molecular weight determination by sodium dodecyl sulfate (SDS) gel electrophoresis were obtained from Serva Fine Biochemicals, Inc., Garden City Park, N.Y. All other chemicals were either reagent grade or the best available.

Crosslinking of $\beta_2$-m by Tgase

At various intervals after the addition of 3 µl of Tgase (11 mg/ml) to a 150-µl solution of $\beta_2$-m (1.26 mg/ml in 30 mM Tris, pH 7.5, 100 mM NaCl, 5 mM CaCl$_2$, 2 mM DTT, 1 mM EDTA), 20-µl aliquots were withdrawn to process for SDS polyacrylamide gel electrophoresis (PAGE) on disc gels (17). When the incorporation of labeled methylamine into $\beta_2$-m was studied, 30 mM $[^{14}C]$methylamine was also included in the incubation mixture. After the usual electrophoresis, staining, and destaining procedures, the band corresponding to $\beta_2$-m was cut out.
from the gel, sliced, solubilized in Soluene-350 (Packard Instrument Co., Inc., Downers Grove, Ill.), and radioactivity was measured in a Packard liquid scintillation counter type 3320.

**Preparation of Human Peripheral Blood Mononuclear Cells**

Human peripheral blood mononuclear cells (PBMC) were separated from freshly drawn blood of healthy adults with a Ficoll-Uromiro (Pharmacia Uppsala, Sweden) gradient according to the method of Böyum (2). The preparation contained 90-95% lymphocytes as determined by morphology, <3% of the cells engulfed latex particles, and 98% of the cells were viable as indicated by trypan blue exclusion.

**Shedding of Surface-labeled Human PBMC and Adsorption of the Shed Supernate on Sephadex Anti-β2-m Particles**

Supernates containing membrane components shed as a result of temperature shift (from 4°C to 37°C) were obtained from surface-labeled human PBMC by the method described by Sarmay et al. (13). 0.6 ml of PBMC suspension (7.5 x 10^7 cells/ml in serum-free Parker medium TC-199), previously surface-labeled by the lactoperoxidase technique (16) at room temperature, then washed and kept at 4°C, was incubated at 37°C for 60 min in the presence of 50 μg/ml active site-inhibited Tgase, 50 μg/ml active enzyme, or 0.1 μg/ml Ca²⁺-ionophore A23187. After centrifugation, the protein concentration (Bio-Rad Protein Assay, Bio-Rad Laboratories, Rockville Centre, N.Y.), as well as the concentration of β2-m (Phadebas β2-m RIA kit, Pharmacia Fine Chemicals, Div. of Pharmacia, Inc., Piscataway, N.J.), was determined in the supernate. Then, portions of the supernates (equal to 1.0 mg protein in each case) were mixed with 0.3-ml packed gel slurries of Sephadex anti-β2-m immunoabsorbent (3 x 10⁹ particles/ml; Pharmacia) which is previously washed in 10 mM Tris-HCl, pH 8.2 + 0.1 M NaCl + 1.5 mM EDTA. After 2 h of stirring at 37°C, the samples were further shaken at 4°C overnight. Afterwards, the slurries were centrifuged and washed free of unbound materials with the Tris-saline buffer. Specifically bound proteins were released by being boiled for 3 min in 100 μl of SDS-PAGE sample buffer containing 2% SDS and 2% 2-mercaptoethanol, then analyzed by SDS-PAGE. Essentially the same procedure was repeated when the Tgase-catalyzed incorporation of [¹⁴C]methylamine (final concentration: 1 mM) into the β2-m of shedding but noniodianated PBMC was studied. After electrophoresis, the gels were stained and sliced into 1-mm thick pieces; then radioactive measurements were performed (8).

**Patching of β2-m on PBMC**

Freshly prepared PBMC suspension was distributed into a series of plastic tubes (2 x 10⁶ cells/tube) in a volume of 200 μl of medium TC-199. In 100 μl of medium, various amounts of Tgase, active site-inhibited Tgase, Ca²⁺-ionophore A23187, respectively, or medium alone, were added to the cell suspensions, which were incubated for 15 min in a water bath at 37° C afterward. Then, the following steps were performed: (a) pelleting of the cells and washing them twice with medium TC-199 at room temperature; (b) resuspending each in a 50-μl solution of diluted anti-human β2-antibody and incubating for 30 min at room temperature; (c) washing them twice in ice-cold medium TC-199; (d) resuspending each in goat anti-rabbit IgG antibody labeled with fluoresceine and incubating in an ice bath for 30 min; (e) washing them twice in ice-cold medium TC-199; and (f) fixing them in 1% paraformaldehyde diluted in phosphate-buffered saline. (In two experiments, the cells were first treated according to step (b), and only then with 5 μg of Tgase; the following steps were the same.) After a last washing in medium TC-199, the cell pellet was resuspended in buffered glycerol and observed under a fluorescence microscope (Fluoval, Zeiss). The proportion of cells showing spots or patches was inferred from examination of 150-200 cells/sample.

**FIGURE 1** Crosslinking of β2-m by tissue Tgase. Proteins were separated on 10% polyacrylamide SDS gels after incubation at 37°C and subsequent denaturation. 1: Guinea pig liver Tgase incubated for 60 min without Ca²⁺. 2: Tgase incubated for 60 min in the presence of Ca²⁺. 3: β2-m. 4: β2-m and Tgase incubated for 60 min without Ca²⁺. 5-7: β2-m and Tgase incubated for 10, 30, and 60 min, respectively, in the presence of Ca²⁺. 8: β2-m and Tgase incubated for 60 min in the presence of Ca²⁺ and 30 mM [¹⁴C]methylamine (83,500 cpm/nmol); 25.2 μg of β2-m was run on the gel; and the cpm measured in the β2-m band was 179,825.
RESULTS

The appearance of high-molecular-weight polymers on the top of 10% SDS polyacrylamide gels with a concomitant decrease of $\beta_2$-m was observed when samples obtained from the Ca$^{2+}$-containing incubation mixture of $\beta_2$-m and Tgase were electrophoresed (Fig. 1). The reaction was time-dependent and accompanied by the disappearance of $\beta_2$-m monomers as well. Although the self-polymerization (1) of the enzyme was complete within 60 min, there were still $\beta_2$-m monomer molecules left. The latter could not be polymerized, even by the addition of more enzyme (results not included in the figure). When 30 mM $[^{14}C]$methylamine was also included in the incubation mixture, the polymerization of $\beta_2$-m did not take place (Fig. 1, gel 8). However, there was an incorporation of labeled methylamine into $\beta_2$-m. From the measured radioactivity incorporated into the known amount of $\beta_2$-m monomer, the number of methylamine molecules coupled to $\beta_2$-m was estimated to be an average of one methylamine molecule per $\beta_2$-m monomer.

$\beta_2$-m, like several other proteins, is shed into the media of PBMC when the cells are exposed to a temperature shift (13). Under our experimental conditions, the protein and $\beta_2$-m concentrations in the sheddings of PBMC varied between 2.0 and 2.2 mg/ml and 50 and 80 $\mu$g/ml, respectively. When the radioactive proteins shed from iodinated PBMC in the presence of Tgase and bound to anti-$\beta_2$-m immunoadsorbent were analyzed on SDS-PAGE, a 33,000, a 70,000, and a higher (not entering into 10% gel) molecular-weight protein were found in addition to the 45,000 (human leukocyte antigen [HLA]) and 12,000 ($\beta_2$-m) molecular weight proteins found in the control sample (Fig. 2). In the case of Ca$^{2+}$-ionophore A23187, the two additional peaks were also detected (33,000 and 70,000 mol wt).

In a separate experiment, noniodinated PBMC were subjected to shedding in the presence of 1.0 mM $[^{14}C]$methylamine and either Tgase or Ca$^{2+}$-ionophore A23187. $\beta_2$-m in the shed supernates was bound to anti-$\beta_2$-immunoadsorbent and then run on SDS-PAGE in the presence of internal standard molecular-weight proteins. The following values in counts per minute (cpm) were measured in the band corresponding to $\beta_2$-m: 85 (shedding in the presence of active site inhibited enzyme), 19,221 (active enzyme), and 3,152 (Ca$^{2+}$-ionophore A23187), showing the apparent incorporation of $[^{14}C]$methylamine into $\beta_2$-m of the shedding PBMC by the enzyme.

$\beta_2$-m can not be patchedor capped on the cell surface by adding only anti-$\beta_2$-m antibodies to the cells, not even when a second antibody (directed against the anti-$\beta_2$-m antibody) is added to the anti-$\beta_2$-m antibody-coated cells in the cold (12). This was confirmed in our experiments with PBMC (Fig. 3a). However, when PBMC were incubated with Tgase at 37°C for 15 min before the coating procedure, the fluorescence, originating from the fluoresceine-labeled secondary antibody, was not found to be homogeneously distributed on the cell surface but was concentrated in spots and patches of various sizes (Fig. 3b, c, and d). Fig. 4 shows the increase in the percentage of spotted/patched cells in suspensions treated with varying enzyme concentrations and with Ca$^{2+}$-ionophore. When the enzyme was added to anti-$\beta_2$-m antibody-coated cells, the portion of spotted/patched cells was even less than in the control experiment. The addition of active site-inhibited Tgase did not result in any change in the homogeneous pattern of fluorescence on the cell surface.

DISCUSSION

The results indicate that $\beta_2$-m is a substrate of Tgase. The result is not surprising because the number of glutamine and lysine residues in the primary amino acid sequence of $\beta_2$-m is considered to be exceptionally high (6). However, not all of the $\beta_2$-m monomers were converted to multimers by the action of the enzyme on the purified protein. Similarly, there were always $\beta_2$ monomers left in the supernate of surface-labeled PBMC shed in the presence of Tgase. In addition, although high concentrations of the enzyme resulted in a substantial increase in the number of spotted/patched cells after Tgase treatment in the antibody-coating experiments, not all cells were affected.

There are several possible explanations for these results. First, the glutamine substrate specificity of Tgase is known to be determined by the amino acid neighbors of the glutamine residue in the polypeptide chain (9). Our observation, that an average of only one methylamine molecule was incorporated
per $\beta_2$-m monomer in the isotope incorporation experiment, suggests the presence of few (perhaps only one) appropriate glutamine residue(s) for the enzyme in the sequence. Also, deamination of $\beta_2$-m may occur during the preparation of the protein or in vivo (there have been reports showing isomeric forms of $\beta_2$-m [3, 14]), and eliminate the appropriate glutamine residue(s) of $\beta_2$-m from some of the molecules. Second, the conformation of $\beta_2$-m (5) may allow the Tgase-catalyzed formation of intramolecular $\epsilon$-(gamma-glutamyl) lysine crosslinks. A recent study (4) depicts the tertiary structure of $\beta_2$-m as having two $\beta$-sheets on top of each other kept together in a sandwich-like manner by hydrophobic bonds. On the edges of the $\beta$ sheets one finds the sequences Glu-Pro-Lys and Lys-Ile-Gln. These sequences are so located that internal isopeptide bonds may be easily formed by Tgase, which would prevent the molecule from participating in the polymerization process.

Third, various $\beta_2$-m “populations” (weakly or strongly attached, complexed to HLA, diffusely packed, etc.) may differ in their availability to interact with the enzyme and other macromolecules on the cell surface.

The catalytic effect of Tgase on $\beta_2$-m may be explained in different ways, which do not necessarily exclude one another. (a) The polymers formed may consist of only crosslinked $\beta_2$-m. On several occasions, we could see the transient appearance of a faint band of $\beta_2$-m dimer on the gel. Its disappearance and our unsuccessful attempts to find other products of the polymerization process can be explained by assuming that the dimer (or trimer, etc.) form is just as good a substrate to the enzyme as the monomeric one. (b) $\beta_2$-m may be copolymerized with Tgase, although it should be noted that changing the $\beta_2$-m:enzyme ratio in separate experiments did not alter the final portion of polymerized $\beta_2$-m (data not shown in figures). (c) In the cellular experiments, $\beta_2$-m may be crosslinked to other membrane protein(s). The presence of newly formed anti-$\beta_2$-m antibody-reactive proteins other than $\beta_2$-m and HLA antigen in the supernate of PBMC shed as a result of Tgase action seems to support this possibility. (d) The change in the distribution of $\beta_2$-m on the surface of PBMC may be the result of the enzyme-catalyzed polymerization of another protein which is closely associated with $\beta_2$-m. We have shown that several proteins (besides $\beta_2$-m) can be labeled on the cell surface using Tgase and a radioactive amine (8).

Both $\beta_2$-m and Tgase are widely distributed and have already been implicated (in some cases with little evidence) in a number of biological functions. At present, it would be premature to speculate on the interrelationship of the two proteins in these functions. Nevertheless, our finding, that Ca$^{2+}$-ionophore A23187 activating Tgase in PBMC led to crosslinkage of
FIGURE 4 The percentage of cells showing spots or patches in differently treated suspensions of PBMC after the addition of rabbit anti-human β₂-m antibody, followed by fluorescein-labeled goat anti-rabbit immunoglobulin. 2 x 10⁸ cells/tube was incubated at 37°C for 15 min with medium TC-199 (1), 25 µg of active site-inhibited Tgase (2), 2.5 µg of Tgase (3), 5 µg Tgase (4), 25 µg of Tgase (5), or 0.03 µg of Ca²⁺-ionophore A23187 (7), respectively, before the addition of the antibodies. In the case of 6, the incubation of PBMC with Tgase took place after the addition of anti-human β₂-m and subsequent washing.

REFERENCES