Identification of Calmodulin-binding Proteins in Chicken Embryo Fibroblasts

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ABSTRACT We recently reported the detection of multiple classes of calmodulin-binding proteins in subcellular fractions of chicken embryo fibroblasts by using a gel binding procedure (Van Eldik, L. J., and W. H. Burgess, 1983, J. Biol. Chem., 258:4539-4547). In this report we identify many of these calmodulin-binding proteins and provide further evidence for the existence of multiple classes of calmodulin-binding proteins based on the interaction of these proteins with calmodulin and other calcium-modulated proteins. The fact that, in some cases, the same calmodulin-binding protein can bind troponin C and S100α suggests that similar functional domains may be present in these distinct calcium-modulated proteins. We also have used protocols based on purification steps for calmodulin-binding proteins and calmodulin-regulated activities from other systems, in conjunction with enzymatic assays and various immunological methods, to identify many of the calmodulin-binding proteins in chicken embryo fibroblasts. The identities of these proteins suggest in vivo roles for calmodulin in the regulation of cell shape and motility, cyclic nucleotide metabolism, and possibly nucleic acid and protein turnover in fibroblasts.

A major advance in our knowledge of the role of calcium in mediating cellular events was the demonstration that chemically homogeneous proteins have physiologically relevant calcium-binding activities (for a review see reference 1). Several classes of calcium-binding proteins have been proposed on the basis of the type of calcium-binding activity and structure they exhibit. The members of one class, the calcium-modulated proteins, are mostly intracellular proteins that reversibly bind calcium with dissociation constants in the micromolar range under physiological conditions and include parvalbumin, calmodulin, troponin C, and S100. No enzymatic activities have been reported for any of these proteins; rather, they are thought to mediate the effects of calcium through their interactions with other proteins.

We recently reported (2) the subcellular distribution of calmodulin and calmodulin-binding proteins in normal and virus-transformed chicken embryo fibroblasts (CEF). In those studies, we identified multiple classes of calmodulin-binding proteins in the crude subcellular fractions. We identified proteins that bind calmodulin in the presence of calcium but not in the presence of calcium chelator (EDTA) and proteins that bind calmodulin in the presence of calcium or EDTA. In this report, we have extended these observations to include an analysis of the binding of troponin C to proteins in the subcellular fractions and the identification of several of these calmodulin-binding proteins. Our results suggest that some functional domains are shared by members of the calcium-modulated proteins whereas others may be unique to calmodulin. The identification of the calmodulin-binding proteins suggests in vivo roles for calmodulin in the regulation of cell shape and motility, cyclic nucleotide metabolism, and perhaps nucleic acid and protein turnover in CEF.

MATERIALS AND METHODS

Materials: Acrylamide, N,N'-methylene-bis-acrylamide, N,N,N',N'-tetramethylethylenediamine, ammonium persulfate, Coomassie Brilliant Blue R-250, and high molecular weight gel standards were from Bio-Rad Laboratories (Richmond, CA). SDS was from BDH Chemicals Ltd. (Poole, England). Low molecular weight gel standards were obtained from Pharmacia Inc. (Piscataway, NJ). Cronex 4 x-ray film was from DuPont (E. I. DuPont de Nemours & Co., Newtown, CT) and 125I (Na125I; 100 mCi/ml) was obtained from Amersham Corp. (Arlington Heights, IL). Lactoperoxidase was from Calbiochem-Behring Corp. (La Jolla, CA); Trizma base, histones, and crystallized BSA were from Sigma Chemical Co. (St. Louis, MO). Vectastain ABC kits were obtained from Vector Laboratories, Inc., (Norwalk, CT). Deionized, pyrogen-
free water was from a Darco Water System (Durham, NC) or a Milli Q system (Millipore Corp., Bedford, MA). All other chemicals were reagent grade. Pure cellulose nitrate paper (0.45-μm pore size) was from Sartorius Filters, Inc. (Hillsboro, CA). The microslab gel apparatus was from Idea Scientific (Corvalis, OR).

**Source of Proteins:** Bovine brain calmodulin was prepared as described (3). Troponin C was prepared from rabbit skeletal muscle as described by Perry and Cole (4). Myosin was prepared from rabbit skeletal muscle according to the method of Fredrickson (5). Myosin light chain kinase (MLCK) was purified from chicken gizzard by using the procedure of Adoestein and Klec (6), and MLCK from turkey gizzard was a gift of Drs. E. Payne and R. Adoestein (National Institutes of Health). Mixed light chains were prepared from bovine cardiac muscle myosin by the method described by Blumenthal and Stull (7). We purified the regulatory light chain by applying mixed light chains (in 10 mM 2-(N-morpholino)propane-sulfonic acid [MOPS], 15 mM 2-mercaptoethanol, pH 7.0) to an Affigel blue (Bio-Rad Laboratories) column that was equilibrated in the same buffer. The column was washed, and regulatory light chains were eluted with a linear gradient of 0-0.4 M KCl in 10 mM MOPS, 15 mM 2-mercaptoethanol, pH 7.0. Phosphorylation b and phosphorylase kinase from rabbit skeletal muscle were provided, respectively, by Drs. R. Uiting and T. Chrisman (this institution). Bovine brain S100a was provided by Dr. D. Marshak (this institution). Carp muscle parvalbumin was provided by Dr. R. H. Kretsinger (University of Virginia).

**Preparation of Protein Fractions:** Subcellular fractions of CEF were prepared by differential centrifugation exactly as described (2). In some experiments, the CEF were prepared by the procedure of Levison (8). Ribosomal protein preparations were prepared from CEF as described by Palmiter (9). A bovine brain fraction of cyclic nucleotide phosphodiesterase that is depleted of calmodulin was prepared as previously described (10). A similar procedure was used to obtain the DEAE fraction from CEF except that DE-52 cellulose was used in place of DE-52 cellulose in the preparation of calmodulin-depleted fractions (11). The DE-52 cellulose was used in place of Sephadex A-50 and the gradient contained 0.1 M NaCl in the start buffer and 0.5 M NaCl in the limit buffer. Fractions were collected, dialyzed against 10 mM ammonium bicarbonate, and lyophilized.

**Gel Binding Procedure Using 125I-Labeled Calcium-modulated Proteins:** The gel binding procedure used in these studies is a modification of a previously described procedure (11). It is based on earlier reports by Burridge (12) for the detection of lectin-binding proteins and antigens after separation in SDS polyacrylamide gels and the studies of Lacks and Springerhorn (13) on the renaturation of enzymes after electrophoresis in SDS polyacrylamide gels. Protein samples were subjected to electrophoresis in the presence of 0.1% (wt/vol) SDS on 7.5 or 12.5% (wt/vol) polyacrylamide gels (0.4-mm thick) using the buffer system of Laemmli (14) and the microslab apparatus described by Matsuda and Burgess (15). We prepared samples for electrophoresis by adding an equal volume of sample buffer immediately before loading them onto the gel. After electrophoresis (~60 min at 90 V constant voltage), samples were placed into 135 × 130-mm plastic boxes (Chem-Products, Div. of American Hinge Corp., Wilton, CT) on a rotary shaker and fixed for 3 h (three changes of 100 ml each) in 25% (vol/vol) isopropanol, 10% (vol/vol) methanol, 10% (vol/vol) acetic acid; and dried and then exposed to x-ray film to detect those proteins in the gel that bind the iodinated protein. In some experiments, 5 mCi EDTA instead of 1 mCi CaCl2 was added to buffer A in the incubation and washes described above. In this protocol, all steps are the same before the incubation with 125I-labeled calcium-modulated proteins and either 1 mM CaCl2 or 5 mM EDTA. Thus we avoid any potential artifacts of differential renaturation in the presence or absence of excess divalent cations or chelators.

The specific activities of the 125I-labeled calcium-modulated proteins used in these studies were in the range of 100-500 Ci/mmol. The length of exposure of the gels to x-ray film varied depending on the specific activity of the protein used and the amount of binding protein applied to the gel. In general a 24-h exposure was sufficient to detect the majority of binding proteins although a 48- or 72-h exposure was often necessary to detect minor components.

In some experiments proteins were transferred from the gel to nitrocellulose paper and then examined for 125I-calmodulin-binding activity. Electrophoresis was performed as above and the gels were washed in 50 mM Tris-HCl, pH 7.6, for 30 min (three changes of 100 ml each) and transferred to nitrocellulose paper using a Bio-Rad transblot apparatus. The chamber was filled with buffer containing 25 mM Tris-HCl, 192 mM glycine, 20% (vol/vol) methanol. Transfer was conducted at 180 mA constant current for 16 h at 4°C, after which the nitrocellulose was incuated for 2 h in 100 ml buffer A containing 1% (wt/vol) BSA and 0.2% (wt/vol) gelatin, rinsed with buffer containing 1 mM CaCl2, and then incubated with 125I-calmodulin as described above.

**Identification of Calmodulin-binding Proteins**

**Detection of DNA-binding Proteins:** Proteins were subjected to electrophoresis and transferred to nitrocellulose paper as described above. Detection of DNA-binding proteins was performed as described by Bowen et al. (16). Total DNA isolated from the sperm of the sea urchin Arbacia punctulata and stained with 125I by nick translation was provided by Dr. R. Zelinski (this institution).

**RESULTS**

**Binding of 125I-Calmodulin and 125I-Troponin C to Proteins in the Subcellular Fractions of CEF**

As previously described (2), we prepared subcellular fractions [N [nuclear], ML [mitochondrial/lysosomal], P [microsomal], and S [soluble]] from CEF. We examined the binding of 125I-calmodulin to proteins in the subcellular fractions in the presence of calcium (Fig. 1B). Clearly, many of the calmodulin-binding proteins in CEF localize specifically to the particulate fractions, primarily ML and P. As noted previously (2), many of the calmodulin-binding proteins in the particulate fractions bind 125I-calmodulin in the presence of calcium or EDTA (Fig. 1C).

To further examine the specificity and the molecular basis...
troponin C constituted a subset of those that bound calmodulin. Most of the proteins that bound 125I–troponin C, to the proteins in the subcellular fractions. The examination of the binding of another calcium-modulated protein, the interactions between calmodulin and these proteins, we

bind iodinated troponin C; i.e., unlabeled troponin C was in the presence of a 500-fold molar excess of unlabeled troponin C. In each panel, lane 1 contains the nuclear (N) fraction of CEF (~9.5 µg), lane 2, the mitochondrial-lysosomal (ML) fraction (~9.5 µg), lane 3, the microsomal (P) fraction (~9.5 µg), lane 4, the soluble (S) fraction (~9.5 µg), and lane 5, the molecular weight standards (labeled by arrows at left of A, Mr × 10^3). The autoradiograms shown are 72 h exposures.

of the interactions between calmodulin and these proteins, we examined the binding of another calcium-modulated protein, troponin C, to the proteins in the subcellular fractions. The results of one such experiment are shown in Fig. 1 D. Troponin C is both a structural and in some cases a functional analogue of calmodulin. Most of the proteins that bound 125I–troponin C constituted a subset of those that bound calmodulin. Many of these proteins were found in the P fraction and were ones that bound calmodulin even in the presence of EDTA. When gels were incubated with 125I–calmodulin in the presence of a 500-fold molar excess of unlabeled troponin C, 125I–calmodulin bound only to those proteins that did not bind iodinated troponin C; i.e., unlabeled troponin C was found to compete with 125I–calmodulin for those proteins that also bind 125I–troponin C (Fig. 1 E). These results indicate that some proteins may bind to a similar domain in both calmodulin and troponin C. The generality of this type of interaction is illustrated more clearly in the studies described below.

**Binding of 125I–Calmodulin and 125I–Troponin C to Purified and Partially Purified Protein Fractions**

Using the gel binding procedure described in Materials and Methods, we also examined the binding of 125I–calmodulin to purified proteins. As shown in Fig. 2 B, we detected binding of 125I–calmodulin to purified MLCK (lane 2) and to the β and γ subunits (Mr = 128,000 and 45,000) of phosphorylase kinase (lane 1). A physiological role for calmodulin in the regulation of these two enzymes appears to be well established. Both enzymes have been purified to chemical and enzymatic homogeneity and the calcium-binding subunit has been shown directly to be calmodulin (23, 24). The binding of 125I–calmodulin to the β subunit of phosphorylase kinase was more apparent when NaCl was not included in the incubation or wash buffer (Fig. 2 C, lane 7). The reversible interaction of calmodulin with the homoenzyme is reported to occur through binding to the α and β subunits whereas the calmodulin that binds as a “tight” or integral subunit of the homoenzyme does so by interaction with the γ subunit (24).

We also detected 125I–calmodulin binding to purified phosphorylase b (Fig. 2 B, lane 3) and to the heavy chain of myosin (Fig. 2 B, lane 4). The physiological significance of the interaction between calmodulin and these two proteins has not been determined. As was the case with the β subunit of phosphorylase kinase, the binding of 125I–calmodulin to phosphorylase b and myosin heavy chain was more readily detected when NaCl was not included in the incubation and wash buffers (Fig. 2 C, lanes 3 and 4). The binding of 125I–calmodulin to all of the proteins shown in Fig. 2 was inhibited when CaCl2 was replaced by 5 mM EDTA in the incubation and wash buffers, or when the gels were washed with EDTA-containing buffers after incubation with 125I–calmodulin in the presence of CaCl2. Similar results were obtained when a 500-fold molar excess of unlabeled calmodulin was included in the incubation with the iodinated protein or when incubations were done in the presence of 1 mM MgCl2 or 1 mM MnCl2.

Calmodulin has four structural domains that are similar to one another in amino acid sequence and to the four structural domains of skeletal muscle troponin C (25). In addition to these structural domains, calmodulin and troponin C, as well as S100, are proposed to have functionally similar drug-binding domains (22). Calmodulin has been shown to substitute functionally for troponin C under certain in vitro conditions (26). The results of the experiments on the binding of 125I–calmodulin and 125I–troponin C to common proteins in the subcellular fractions of CEF also suggest the presence of functionally similar protein-binding domains in these calcium-modulated proteins. To further investigate these potential similarities, we examined the binding of 125I–troponin C and 125I–S100α to purified proteins using the same conditions as were used for calmodulin.

We detected binding of 125I–troponin C to several of the proteins that also bound 125I–calmodulin. For example, as shown in Fig. 3 B, phosphorylase kinase (lane 1), phosphorylase b (lane 3), myosin heavy chain, and an Mr = 35,000 polypeptide in the myosin sample (lane 4) bound both 125I–troponin C and 125I–calmodulin. In contrast, MLCK bound...
**Figure 2** The binding of $^{125}$I-calmodulin to purified proteins. A shows a Coomassie Blue-stained 7.5% (wt/vol) acrylamide SDS gel. B and C show corresponding autoradiograms of gels that were incubated after electrophoresis with $^{125}$I-calmodulin in the presence of (B) 1 mM CaCl$_2$ and 0.2 M NaCl or (C) 1 mM CaCl$_2$ and no added NaCl. The autoradiogram in B is a 72-h exposure; C is a 24-h exposure. In each panel, lane 1 contains phosphorylase b kinase from rabbit skeletal muscle (5 μg), lane 2, MLCK from turkey gizzard (1 μg), lane 3, phosphorylase b from rabbit skeletal muscle (2 μg), and lane 4, myosin from rabbit skeletal muscle (3 μg). (M, $\times 10^{-3}$ in this and the following gels.)

**Figure 3** The binding of $^{125}$I-troponin C to purified proteins. A shows a Coomassie Blue-stained 7.5% (wt/vol) acrylamide SDS gel. B and C show corresponding autoradiograms of gels that were incubated after electrophoresis with $^{125}$I-troponin C in the presence of 1 mM CaCl$_2$ and 0.2 M NaCl or (C) $^{125}$I-calmodulin in the presence of 1 mM CaCl$_2$, 0.2 M NaCl, and a 500-fold molar excess of unlabeled troponin C. Both autoradiograms are 24-h exposures. In each panel, lanes 1–4 contain the same samples as described in Fig. 2.

$^{125}$I-calmodulin but showed little $^{125}$I-troponin C binding (cf. lane 2, Figs. 2B and 3B). Similarly, proteins in crude bovine brain extracts that bound $^{125}$I-troponin C were generally distinct from those that bound $^{125}$I-calmodulin. Unlike calmodulin, the binding of $^{125}$I-troponin C to these proteins was not diminished by substitution of 5 mM EDTA for 1 mM CaCl$_2$ in the incubation and wash buffers. In addition, binding of $^{125}$I-troponin C to the β subunit of phosphorylase kinase (cf. lane 1, Figs. 2B and 3B) was easily detected even in the presence of 0.2 M NaCl. When a 500-fold molar excess of unlabeled troponin C was included in the incubation with $^{125}$I-calmodulin (Fig. 3C), unlabeled troponin C did not compete with $^{125}$I-calmodulin for binding to MLCK but did compete for binding to phosphorylase b, phosphorylase kinase, and myosin. These observations are consistent with the results obtained using $^{125}$I-troponin C alone. The binding patterns observed for $^{125}$I-S100α are similar to those observed using $^{125}$I-troponin C in that $^{125}$I-S100α bound phosphorylase kinase, phosphorylase b, and myosin but not MLCK. The binding patterns of $^{125}$I-S100α to proteins in crude bovine brain extracts also are qualitatively similar to those obtained using $^{125}$I-troponin C (data not shown).

Carp muscle parvalbumin is a calcium-modulated protein that contains calcium-binding structures, but not drug-binding domains, that are similar to those in calmodulin, troponin C, and S100α (22). In contrast to the results obtained using $^{125}$I-troponin C and $^{125}$I-S100α, we could detect only weak binding in the presence of 1 mM CaCl$_2$ of $^{125}$I-parvalbumin to phosphorylase b, and little or no binding to MLCK (Fig. 4B). In the presence of 5 mM EDTA, the binding of $^{125}$I-parvalbumin to phosphorylase b was enhanced (Fig. 4C, lane 1).

In conclusion, the binding patterns observed for $^{125}$I-calmodulin, $^{125}$I-troponin C, $^{125}$I-S100α, and $^{125}$I-parvalbumin cannot be explained by the fact that they are all relatively small, acidic, calcium-binding proteins. Consistent with this observation is the fact that α-lactalbumin, a small, acidic and calcium-binding protein that is not clearly related structurally or functionally to calmodulin, troponin C, S100α, or parvalbumin did not inhibit the binding of $^{125}$I-calmodulin to the proteins examined, even when present at a 1000-fold molar excess. Finally, there is a specific set of calmodulin-binding proteins that bind troponin C and S100α. However, there are also other proteins that bind troponin C or S100α but do not bind calmodulin.
Identification of the Calmodulin-binding Proteins in CEF

The results described above indicate that different classes of calmodulin-binding proteins can be detected by gel binding experiments using calmodulin and related proteins. The physiological significance of these findings as they relate to CEF cannot be determined without first identifying the binding proteins in the various subcellular fractions.

The apparent molecular weights and predominantly N-fraction localization of the two binding proteins in lane 1 of Fig. 1, B and C suggest that these proteins are histones. As previously described (2), some nuclear lysis occurs when this particular subcellular fractionation protocol is used. Fig. 5A shows the binding of 125I-calmodulin to a nuclear preparation of CEF where the integrity of the nuclei was maintained as judged by total DNA determinations. This preparation (cf. Fig. 1, B, lane 1 and Fig. 5, A, lane 2) is enriched for these two calmodulin-binding proteins. These proteins co-migrated with histone standards (Fig. 5, B) and exhibited similar DNA-binding activities (Fig. 5, C). It should be noted that one of the histones (H2a) was washed out of the gel during the 125I-calmodulin-binding protocol (cf. Fig. 5, A and B). This result is similar to those of Irie and Sezaki (27) who observed that histone H2a was preferentially extracted from acrylamide gels when the gels were fixed with 25% isopropanol, 10% acetic acid before staining.

As can be seen in Fig. 1, there were many proteins in the P fraction with apparent molecular weights ranging from 15,000 to 40,000 that bound 125I-calmodulin and 125I-troponin C (lanes 3 in Fig. 1, B–D). The general pattern is similar to gel profiles of various heterogeneous nuclear ribonucleoprotein particle preparations and also to one-dimensional gel profiles of ribosomal protein preparations (28, 29). The data shown in Fig. 5 demonstrate that these proteins are not of nuclear origin since they are not present in the nuclear preparation. However, the apparent molecular weights and subcellular distribution were found to be similar to those of ribosomal proteins. We used several procedures for the preparation of eucaryotic ribosomal proteins to determine whether these binding proteins may indeed be ribosomal proteins. The results of one such experiment are shown in Fig. 6. A comparison of lanes 1 and 2 illustrates that many but not all of the 125I-calmodulin-binding proteins in the P fraction of CEF were present in a ribosomal protein preparation from CEF and suggests that these binding proteins may be ribosomal proteins.

Burrage et al. (30) detected the presence of nonerythroid spectrin in CEF by using antibodies against brain spectrin (fodrin) and immuno-autoradiography on gels. Several laboratories have demonstrated binding of 125I-calmodulin to nonerythroid spectrin (fodrin) from various cell types including bovine brain (31, 32). The P and S fractions of CEF were analyzed for the presence of calmodulin-binding proteins that cross-react with bovine brain spectrin (fodrin) antibodies. The results of one such experiment are shown in Fig. 7. Proteins with apparent molecular weights of ~240,000 and 150,000 that cross-react with the antibodies and also bind 125I-calmodulin were detected in the CEF fractions. A protein of apparent molecular weight 150,000 has been postulated to be a proteolytic fragment of bovine brain spectrin (fodrin) (30).

Several proteins with apparent molecular weights ~60,000...
that bind $^{125}$I-calmodulin in the presence of calcium but not in the presence of chelator were detected in the S fraction of CEF. It is possible that these proteins are calmodulin-activatable cyclic nucleotide phosphodiesterase and calcineurin, a calmodulin-activatable phosphatase (33), since both of these calmodulin-binding proteins have calmodulin-binding subunits of $M_r$ ~60,000 (34). To test this possibility, we prepared from bovine brain a partially purified fraction with calmodulin-activatable cyclic nucleotide phosphodiesterase activity and calmodulin-activatable nitrophenylphosphate phosphatase activity. The binding of $^{125}$I-calmodulin and calcineurin antibodies to proteins in this fraction and to proteins in the S fraction of CEF are shown in Fig. 8A and B. In both fractions, proteins that cross-react with the calcineurin antibodies and also bind $^{125}$I-calmodulin were detected. When CEF were subjected to a purification protocol similar to the one used to prepare calmodulin-activatable phosphodiesterase from bovine brain, fractions from the DEAE column were obtained that have calmodulin-activatable cyclic nucleotide phosphodiesterase activity and contain $^{125}$I-calmodulin-binding proteins that co-migrate with $^{125}$I-calmodulin-binding proteins in the bovine brain fraction (Fig. 8, C and D).

Finally, $^{125}$I-calmodulin-binding activity similar to that exhibited by purified MLCK was detected in one of the fractions (Fig. 8D, lane 4). This fraction contains MLCK activity using cardiac regulatory light chain as a substrate. When the subcellular fractions of CEF were examined for MLCK immunoreactivity by immunoblot analysis, a band that co-migrates with MLCK purified from chicken gizzard was detected in the S fraction (data not shown). MLCK immunoreactivity was also present in CEF extracts as judged by competition radioimmunoassay using $^{125}$I-labeled chicken gizzard MLCK as tracer (Fig. 9).

DISCUSSION

The results presented here and elsewhere (2) demonstrate that multiple classes of calmodulin-binding proteins can be identified. There are proteins that bind calmodulin in the presence of added calcium, ones that bind calmodulin in the presence of calcium or chelator, ones that appear to bind only calmodulin, and others that bind calmodulin and other calcium-modulated proteins. These results suggest that calmodulin may contain multiple functional domains for interaction with binding proteins, that some of these domains may be shared by troponin C, S100a, or parvalbumin, and that others may be unique to calmodulin.

The studies on the binding of $^{125}$I-troponin C to proteins in the subcellular fractions of CEF indicate that many of the “calmodulin-binding proteins” in the particulate fractions are not exclusive for calmodulin. However, the proteins in the subcellular fractions of CEF that bind both calmodulin and troponin C cannot be discounted merely as “nonspecific” binding proteins. For example, we have demonstrated here that calmodulin, troponin C, and S100a bind to phosphorylase, myosin, and subunits of phosphorylase kinase. The inter-

**FIGURE 7** Identification of spectrin from CEF as an $^{125}$I-calmodulin-binding protein. Proteins were separated on a 7.5% (wt/vol) acrylamide SDS gel, transferred to nitrocellulose paper, and incubated with spectrin antibodies (A) or $^{125}$I-calmodulin (B). In each panel, lane 1 contains the S fraction from CEF and lane 2 contains the P fraction from CEF. Proteins with apparent molecular weights of 240,000, 150,000, and 60,000 are labeled with arrows.

**FIGURE 8** Detection of calcineurin immunoreactivity and calmodulin binding to proteins from bovine brain and CEF. In A and B, proteins were separated on a 7.5% (wt/vol) acrylamide SDS gel, transferred to nitrocellulose paper, and incubated with either calcineurin antibodies (A) or $^{125}$I-calmodulin (B). Lane 1 contains a fraction from bovine brain with calmodulin-activatable phosphodiesterase and phosphatase (calcineurin) activities and lane 2 contains the S fraction from CEF. The arrow labeled 60 points to the region of the gel containing proteins of apparent molecular weight 60,000. Panel C shows a Coomassie-Blue-stained 7.5% (wt/vol) acrylamide SDS gel and D shows the corresponding autoradiogram depicting the binding of $^{125}$I-calmodulin. In C and D, lane 1 contains molecular weight standards (indicated at left), lane 2 contains the DEAE fraction from CEF with maximal calmodulin-activatable phosphodiesterase activity, lane 3 contains the DEAE fraction from bovine brain with maximal calmodulin-activatable phosphodiesterase activity, and lane 4 contains the DEAE fraction from CEF with maximal calmodulin-activatable MLCK activity. The arrows point to the regions of the gel containing MLCK (M) or proteins of apparent molecular weight 60,000 (60).
action of calmodulin with phosphorylase has recently been demonstrated by other procedures (35), yet the significance of this interaction is not known. The γ subunit of phosphorylase kinase, whose interaction with calmodulin has been shown by other procedures and is considered to be physiologically relevant, also binds troponin C and S100α in the gel binding procedure. Although we believe this procedure is an excellent method for screening protein fractions for potential calmodulin-binding activity, caution must be used in assigning or discounting physiologically relevant calmodulin-binding activity on the basis of gel binding data alone. Similarly, calmodulin-Sepharose interactions (36) or cross-linking of added calmodulin (37) do not, by themselves, provide any indication as to whether or not proteins bind calmodulin in vivo. Gel overlay analysis combined with enzymology, protein chemistry, immunochemistry, and cell biology studies can provide, as shown here, insight into potential biological roles of calmodulin.

We have extended our initial studies on the detection of calmodulin-binding proteins with the gel binding procedure and have now identified many of the calmodulin-binding proteins and calmodulin-regulated activities in CEF. Some of these have been identified in many cell types other than CEF, some have been identified in CEF but their calmodulin-binding properties have not been investigated, and others have not been previously reported to be calmodulin-binding proteins in any cell type. For example, MLCK, cyclic nucleotide phosphodiesterase, and calmodulin-dependent phosphotase (calcinurin) from many cell types have been shown to be calmodulin-binding proteins with calmodulin-regulated activities. We have demonstrated the presence of MLCK in CEF by kinase activity, by immunoreactivity in immunoblots and competition radioimmunoassay, and by 125I-calmodulin binding. We have identified 125I-calmodulin-binding proteins with apparent molecular weights in the range of 55,000–60,000 and we can detect calmodulin-activatable cyclic nucleotide phosphodiesterase activity and calcineurin immunoreactivity in CEF. However, we are not yet able to assign cyclic nucleotide phosphodiesterase activity or calcineurin activity to a particular 125I-calmodulin-binding protein in CEF.

Nonerythroid spectrin (fodrin) in CEF has been identified by immunoreactivity with cell extracts after electrophoresis (30). We have correlated immunoreactivity with 125I-calmodulin-binding activity in the S, and to a lesser extent the P, fraction of CEF. Similarly, we have correlated DNA binding activity with 125I-calmodulin-binding activity in nuclei preparations of CEF, and have shown that these proteins co-migrate with histone standards. Finally, we have shown that many of the 125I-calmodulin binding proteins in the P fraction of CEF can be identified in ribosomal protein preparations of CEF.

Overall, the results presented here on the identification of calmodulin-binding proteins suggest specific roles for calcium and calmodulin regulation of cyclic nucleotide metabolism and cell shape and motility in CEF. The physiological significance of the interaction of calmodulin with some of these proteins, such as the ribosomal proteins and the histones, has not been established. Clearly, further studies in this area are needed. However, there are reports of effects of calcium on protein synthesis, and calcium and calmodulin on nucleic acid synthesis (38–41). Some of the calmodulin-binding proteins identified in these fractions may represent sites of regulation.

The results obtained here suggest that many particulate calmodulin-binding proteins interact with a calmodulin domain that may be found in the structurally related protein troponin C, and that several binding proteins interact with calmodulin in a less calcium-dependent, possibly calcium-independent, manner. It should be noted that although some of the protein-protein interactions shown here appear to be relatively calcium independent, any potential regulation of these proteins may indeed be calcium dependent. Related to our results, Andreasen et al. (42) recently purified a membrane protein from bovine cerebral cortex whose apparent affinity for calmodulin is higher in the presence of calcium chelators than in the presence of calcium. In contrast, most of the soluble calmodulin-binding proteins from CEF and several particulate calmodulin-binding proteins do not bind troponin C and bind calmodulin in the presence of excess calcium but not in the presence of EDTA. These results are consistent with the possibility that calmodulin could be "compartmentalized" in the cell by virtue of its interaction with particulate binding proteins (2, 11, 42, 43) or through the use of different binding domains on the calmodulin molecule. These interactions could occur even at very low calcium concentrations and this compartmentalized calmodulin could still be potentially accessible to other calmodulin-binding proteins via a different binding domain. In the case of particulate bound calmodulin, this could include accessibility to "soluble" calmodulin-binding proteins.

In summary, these studies provide insight into the potential role of calmodulin in CEF cell function and a framework for future studies addressing the specificity of its interactions with binding proteins. The correlation of structural and functional domains in the calmodulin molecule and further characterization of the interactions with the various classes of binding proteins are necessary for a complete understanding of the
molecular mechanisms of action of calcium-modulated proteins. In addition, the results presented here identify specific targets or activities for potential calcium and calmodulin regulation in CEF. That some of the activities may change as a function of the cell cycle and upon viral transformation of CEF, coupled with the demonstrated changes in calmodulin levels during the cell cycle and upon viral transformation (2, 44–46), indicate future directions for a more complete understanding of the role of calmodulin in the regulation of cell function.

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