Cbx4 maintains the epithelial lineage identity and cell proliferation in the developing stratified epithelium

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During development, multipotent progenitor cells establish lineage-specific programmers of gene activation and silencing underlying their differentiation into specialized cell types. We show that the Polycomb component Cbx4 serves as a critical determinant that maintains the epithelial identity in the developing epidermis by repressing nonepidermal gene expression programs. Cbx4 ablation in mice results in a marked decrease of the epidermal thickness and keratinocyte (KC) proliferation associated with activation of numerous neuronal genes and genes encoding cyclin-dependent kinase inhibitors (p16/p19 and p57). Furthermore, the chromodomain- and SUMO E3 ligase–dependent Cbx4 activities differentially regulate proliferation, differentiation, and expression of nonepidermal genes in KCs. Finally, Cbx4 expression in KCs is directly regulated by p63 transcription factor, whereas Cbx4 overexpression is capable of partially rescuing the effects of p63 ablation on epidermal development. These data demonstrate that Cbx4 plays a crucial role in the p63-regulated program of epidermal differentiation, maintaining the epithelial identity and proliferative activity in KCs via repression of the selected nonepidermal lineage and cell cycle inhibitor genes.

Introduction

During development, tissue differentiation relies on the establishment of specific patterns of gene expression, which is achieved by lineage-specific gene activation and silencing in multipotent stem cells and their progenies (Slack, 2008; Blanpain and Fuchs, 2014). The program of epidermal differentiation in mice begins at about embryonic day 9.5 (E9.5) and results in the formation of an epidermal barrier by E18.5 (Koster and Roop, 2007; Blanpain and Fuchs, 2009). The process of terminal differentiation in epidermal cells is executed by sequential changes of gene expression in the keratin type I/II loci, followed by the onset of expression of the epidermal differentiation complex genes encoding the essential components of the epidermal barrier (Fuchs, 2007). This program is governed by the coordinated involvement of several transcription factors (p63, AP-1, Klf4, Arnt, etc.), signaling pathways (Wnt, Bmp, Hedgehog, EGF, Notch, FGF, etc.), and epigenetic regulators (DNA/histone-modifying enzymes, Polycomb genes, higher order and ATP-dependent chromatin remodelers, and noncoding and microRNAs) that control expression of lineage-specific genes (Khavari et al., 2010; Botchkarev et al., 2012; Frye and Benitah, 2012; Perdigoto et al., 2014).

Among these regulatory molecules, the p63 transcription factor serves as a master regulator of epidermal development and controls expression of a large number of distinct groups of genes (Viganò and Mantovani, 2007; Vanbokhoven et al., 2011; Botchkarev and Flores, 2014; Kouwenhoven et al., 2015). p63 knockout (KO) mice fail to form stratified epithelium and express several epidermis-specific genes (Mills et al., 1999; Yang et al., 1999). In the epidermis, p63 regulates the expression of distinct chromatin-remodeling factors, such as Satb1 and Brg1, which, in turn, control the establishment of specific nuclear positioning and conformation of the epidermal differentiation complex locus required for full activation of keratinocyte (KC)-specific genes during terminal differentiation (Fessing et al., 2011; Mardaryev et al., 2014).

Epigenetic regulators exhibit both activating and repressive effects on chromatin in KCs: the histone demethylase Jmjd3, ATP-dependent chromatin remodeler Brg1, and genome
organizer Satb1 promote terminal KC differentiation, whereas the DNA methyltransferase DNMT1, histone deacetylases HDAC1/2, and Polycomb components Bmi1 and EzH1/2 stimulate proliferation of the progenitor cells via repression of the genes encoding cell cycle inhibitors, as well as inhibiting premature activation of terminal differentiation–associated genes (Sen et al., 2008, 2010; Ezzhkova et al., 2009; LeBoeuf et al., 2010; Fessing et al., 2011; Mardaryev et al., 2014).

Polycomb chromatin-remodeling proteins form two complexes (Polycomb repressive complex 1 and 2 or PRC1/2) that compact the chromatin and inhibit transcription by preventing binding of the transcription machinery to gene promoters (Simon and Kingston, 2013; Cheutin and Cavalli, 2014). Recent data reveal that binding of the noncanonical PRC1 complex containing histone demethylase KDM2B, PCGF1, and RING/YY1-binding protein (RYBP) promotes basal ubiquitylation of the H2A at lysine 119 (H2A-K119) at unmethylated CpG-rich DNA regions, which is sufficient to recruit the PRC2 complex (Blackledge et al., 2014; Cooper et al., 2014; Kalb et al., 2014). The PRC2 component EzH1/EzH2 histone methyltransferase promotes trimethylation of H3K27, followed by targeting of the Cbx proteins as a part of the canonical PRC1 complex to H3K27me3, which result in further increase of the H2AK119 ubiquitylation catalyzed by the PRC1 component Ring1b (Simon and Kingston, 2013; Cheutin and Cavalli, 2014; Perdigoto et al., 2014; Schwarz and Pirrotta, 2014).

In the epidermis, the Polycomb components Bmi1, EzH1/2, and Jarid2 stimulate proliferation of the progenitor cells via repression of the genes encoding cell cycle inhibitors, including the INK4A–INK4B locus, as well as inhibit premature activation of terminal differentiation–associated genes (Ezhkova et al., 2009; Meijetta et al., 2011). In addition, EzH1/2 restricts differentiation of the epidermal progenitor cells by repressing the Sox2 gene, which, in turn, promotes Merkel cell–specific differentiation (Bardot et al., 2013).

The Cbx4 gene belongs to the PRC1 family and has a chromodomain interacting with the H3K27me3 histone mark and mediating transcriptional repression together with the PRC2 complex (Li et al., 2007; Luis et al., 2011). Cbx4 also possesses SUMO E3 ligase activity that promotes sumoylation of other proteins, including DNA methyltransferase Dnmt3a, thus providing a link between Polycomb-mediated gene silencing and DNA methylation (Li et al., 2007). In normal human skin, the Cbx4 protein protects epithelial stem cells from senescence through PRC-dependent repression of the Ink4a locus, as well as controls their differentiation through PRC-independent mechanisms (Luis et al., 2011). These data suggest Cbx4 as a critical determinant regulating the activity of stem cells and their progenies in the skin. However, its role in the control of epidermal development and in establishing tissue-specific gene expression patterns still remains unclear.

Here, we show that Cbx4 is a critical determinant regulating the establishment and maintenance of the epidermal differentiation program that represses nonepidermal lineage genes and controls cell proliferation/differentiation in the epidermis. We also show that Cbx4 serves as a direct p63 target and mediates its repressive effects on the expression of nonepidermal lineage genes and selected cell cycle inhibitor genes. These data illustrate how p63 transcription factor operates in concert with epigenetic regulator Cbx4 to establish and maintain the lineage-specific program in differentiating cells and in governing epithelial differentiation/function.
whereas immunostaining for active caspase 3 showed a lack of any changes in the KC apoptosis between WT and Cbx4 KO mice (Fig. S3, E and F). The reduced proliferation of E16.5 basal epidermal KCs was associated with increased expression of the cell cycle inhibitor genes (senescence-linked Ink4a locus p16/p19 and Cdkn1c/p57 gene; Fig. S3, A and B), which are well-known targets of the Polycomb proteins in several tissues, including the skin (Ezhkova et al., 2009; Luis et al., 2011).

As CBX4 was previously shown to prevent terminal differentiation in human epidermal progenitor cells independently of its Polycomb-associated repressive function (Luis et al., 2011), we explored the effect of Cbx4 ablation on the expression of early and late epidermal differentiation genes in the developing mouse epidermis. In the absence of Cbx4, cells of the embryonic ectoderm were able to acquire an epidermal fate and commence a differentiation program as indicated by positive K14 and K10 staining in the basal and suprabasal cells (Figs. 1 I and S4 A). However, Loricrin, a marker of terminally differentiating KCs, was prematurely expressed in the suprabasal cells located just above the basal cells in the Cbx4-deficient epidermis at E14.5, whereas its expression was not detectable in WT epidermis at this stage (Figs. 1 J and S4 A). In E16.5 WT embryos, the most superficial late suprabasal cells underwent terminal differentiation and expressed the key components of the cornified cell envelope such as Loricrin, Involucrin, and Filaggrin (Figs. 1 J and S4 A). In contrast to WT mice, the premature onsets of Loricrin, Involucrin, and Filaggrin expression were seen in the early suprabasal cells in E16.5 Cbx4 KO mice (Figs. 1 J and S4 A). Immunofluorescence data on the expression of the markers of terminally differentiated KCs were in concordance with quantitative real time–PCR (qRT-PCR) data showing the markedly elevated levels of the Loricrin, Involucrin, and Filaggrin mRNA expression in the LCM-captured basal epidermal cells of E16.5 Cbx4 KO mice versus WT controls (mean ± SD). n = 3. *** P < 0.001. Bars, 25 µm. ns, not significant.
transcripts in the E16.5 epidermis of Cbx4KO mice versus WT controls (Fig. 1 K). Thus, Cbx4 suppresses the expression of terminal differentiation–associated genes in the basal progenitor cells during murine epidermal development.

**Cbx4 represses nonepidermal lineage genes in KCs during epidermal development**

Next, we analyzed the distribution of H2AK119Ub and H3K27me3, two key histone modifications associated with Polycomb-mediated gene repression, in the epidermis of Cbx4KO mice. According to the model of Polycomb-mediated gene silencing, a basal level of H2AK119 ubiquitylation at unmethylated CpG-rich DNA regions is maintained by noncanonical PRC1 complexes lacking the Cbx proteins, whereas one of the Cbx family members serving as a part of the canonical PRC1 complex is only involved in the subsequent step in the Polycomb-mediated gene silencing and is required for binding to H3K27me3, followed by the Ring1b-mediated further increase of the H2AK119 ubiquitylation over its basal level (Simon and Kingston, 2013; Cheutin and Cavalli, 2014; Perdigoto et al., 2014; Schwartz and Pirrotta, 2014). In contrast to WT mice, epidermal cells in E14.5 and E16.5 Cbx4KO mice showed a significantly decreased level of H2AK119ub1 expression, which we considered as a basal level (Fig. 2, A and B). However, the expression levels of H3K27me3 were not changed in the epidermis of Cbx4KO mice compared with WT controls (Fig. 2, C and D). Also, expression of the histone methylase Kdm2b, a constituent of the noncanonical PRC1 complex (Schwartz and Pirrotta, 2014), was increased in the epidermis of Cbx4KO mice, whereas expression of Ring1b forming the canonical PRC1 complex together with Cbx4 and catalyzing the H2AK119 ubiquitylation was not changed (Fig. S2, A and B). Importantly, Cbx4 ablation changed neither H2AK119ub1 nor H3K27me3 expression in the dermal cells (Fig. S2 C), thus suggesting epidermis-specific effects of Cbx4 gene repression in the skin. These data were consistent with the data showing the significantly increased Cbx4 expression over its basal level mediated by PRC1 in epidermal cells.

Our data also demonstrate that the expression levels of H2AK119ub1 in the epidermis of E18.5 Cbx4KO and WT mice became quite similar (Fig. 2, A and B), which suggests that other Cbx proteins (Fig. S1 C) might substitute for Cbx4 in the canonical PRC1 complex in Cbx4KO mice. These data are also quite consistent with the data demonstrating that maximal differences in the epidermal thickness and proliferation between WT and Cbx4KO mice are seen at E16.5, whereas such differences become insignificant at E18.5 (Fig. 1, E–H).

To determine the molecular targets of Cbx4 in the epidermal KCs during development, we correlated a microarray transcript profiling from E16.5 LCM-captured Cbx4-deficient basal cells with Cbx4 chromatin immunoprecipitation (ChIP) sequencing (ChIP-seq) analyses performed on primary epidermal KCs isolated from E16.5 WT mice. About 13.8% of genes up-regulated in the basal epidermal cells from Cbx4KO mice showed Cbx4 ChIP-seq binding, thus likely representing direct Cbx4 targets in KCs (Fig. 2, E–G). Because the Cbx4 chromodomains bind to H3K27me3, which results in a further increase of the H2AK119 ubiquitylation over its basal levels catalyzed by the PRC1 component Ring1b (Li et al., 2007; Luis et al., 2011), we merged data for direct Cbx4 targets and ChIP-seq data for H3K27me3 and found that 60.9% of Cbx4 direct target genes in primary epidermal KCs show peaks for both Cbx4 and H3K27me3 within 100 kb of their transcription start sites (Fig. 2 E and Table S5). Furthermore, 12.1% of these genes showed the overlapping ChIP-seq peaks for Cbx4 and H3K27me3, thus pointing to these genes as direct targets for Cbx4-mediated repression (Table S5).

The DAVID bioinformatics tool for gene ontology functional annotation revealed an enrichment of the genes involved in the control of neuronal development among the Cbx4 targets (Fig. 2, F and G; and Table S5). In primary epidermal KCs, several neuronal and other lineage-specific genes (Neft, Neurog3, Mobp, En2, and Lhx4) contained both Cbx4 and H3K27me3 ChIP-seq peaks in their regulatory regions (Fig. 2 H). Validation by qRT-PCR showed increased transcript levels of the selected Cbx4 target genes in the Cbx4-null epidermal cells (Fig. 2 I), including the Neft and Mobp genes encoding a neurofilament light polypeptide and myelin-associated oligodendrocyte basic protein, structural components of the neurons, and Schwann cells, respectively (Montague et al., 1997; Jordanova et al., 2003). In addition, expression of the Neurog3, Olig2, En2, and Lhx4 genes, encoding the corresponding transcription factors controlling the development of nervous system and some other non–KC cell lineages (Hubert and Westphal, 2000; Wallén and Perlmann, 2003; Pelling et al., 2011; Mitew et al., 2014), was markedly increased in the epidermis of Cbx4KO mice compared with WT controls (Fig. 2 I).

Interestingly, Neft transcript levels showed an inverse correlation with Cbx4 expression in the WT epidermis during E11.5–E16.5 (Figs. 1 A, 2 I, and S1 C). Further analysis revealed that, in addition to the dermal nerve fibers, an ectopic expression of the Neft protein was seen in both basal and suprabasal epidermal layers of E16.5 Cbx4KO mice, whereas an exclusively neuronal Neft expression was seen in WT mice (Fig. 2 J). Furthermore, ChIP–quantitative PCR (ChIP-qPCR) analyses showed enrichments for Cbx4, H2AK119ub1, and H3K27me3 at the regulatory region of the Neft gene, whereas a lack of Cbx4 enrichment was seen at the regulatory region of the Lor gene that did not belong to the category of the Cbx4 direct targets and served as a control in this experiment (Fig. 2 K). In addition, shRNA-mediated Cbx4 gene silencing resulted in a significant decrease in the Cbx4 and H2AK119ub1 levels at the regulatory region of the Neft gene in primary epidermal KCs, whereas the H3K27me3 levels were not changed (Figs. 2 L and S2 D). These data were consistent with the data demonstrating the decrease in the H2AK119ub1 levels in the epidermis of Cbx4KO versus WT mice (Fig. 2, A and B) and suggest that Cbx4 as a part of the canonical PRC1 complex facilitates repression of its target Neft gene. However, Cbx4 ablation did not affect Merkel cell numbers in the developing epidermis or expression of the transcription factors (Atoh1 and Sox2) involved in their differentiation (Fig. 4, B and C; and Table S5; Van Keymeulen et al., 2009; Bardot et al., 2013; unpublished data).

**Chromodomain- and SUMO E3 ligase–dependent Cbx4 activities differentially regulate proliferation, differentiation, and expression of nonepidermal genes in KCs**

As Cbx4 uniquely possesses, among PRC1-associated Cbx subunits, both Polycomb and non-Polycomb SUMO E3 ligase activities (Li et al., 2007; Luis et al., 2011), we next
questioned whether its effects on epidermal proliferation, differentiation, and repression of nonepidermal genes seen during murine skin development are associated with one of these Cbx4 activities. We infected primary epidermal KCs isolated from newborn WT mice with retroviral constructs expressing either WT Cbx4 (Cbx4WT), chromodomain-mutated Cbx4 (Cbx4CDM), or E3 ligase–deficient Cbx4 (Cbx4ΔSIM; Luis et al., 2011).

Interestingly, transfection of cells with Cbx4WT or Cbx4CDM constructs did not show any effect on the number of proliferating Ki67+ cells, whereas a prominent decrease in proliferation was seen in the cells expressing the E3 ligase–deficient Cbx4ΔSIM construct (Fig. 3, A and B). These data were consistent with the data showing the decrease of epidermal proliferation in Cbx4KO mice (Fig. 1, G and H) and suggest that the effects of Cbx4 on proliferation in cultured KCs are mediated predominantly by its E3 ligase domain.
Given that Cbx4 suppresses senescence in human epidermal KCs (Luis et al., 2011), we examined the mechanisms underlying the Cbx4-dependent regulation of the senescence-linked cell cycle inhibitor genes and performed ChIP-qPCR analyses that revealed Cbx4 binding to the Cdkn2a (p16/p19) regulatory regions in primary epidermal KCs (Fig. S3 C). Transfection of primary epidermal KCs with Cbx4WT or Cbx4CDM constructs showed that the Cbx4 chromodomain is involved in regulating the expression of Cdkn2a and Cdkn1c (p57; Fig. S3 D). Together with our data demonstrating an increase in the expressions of p16/p19, p57, and the senescence marker γ-H2AX in the epidermis of Cbx4KO mice versus WT controls (Fig. S3, A, B, and E), these data demonstrated an involvement of the Cbx4 chromodomain in the control of expression of the senescence-associated p16/p19 and p57 genes. These data were in full concordance with the data of Luis et al. (2011) that show similar effects of the Cbx4 chromodomain on the expression of senescence-associated genes in human epidermal KCs.

To assess whether targeting of Cbx4 in the primary KCs affects cell differentiation, we analyzed the expression of two established terminal differentiation-associated markers (Involucrin and Filaggrin) after infection of cells with the WT or mutated Cbx4-expressing retroviruses. Cbx4WT-expressing cells showed a decrease in protein and gene expression of Involucrin in comparison with control cells (Fig. 3, C and D). However, Cbx4ΔSIM-treated KCs showed a prominent increase in Involucrin protein and gene expression compared with Cbx4WT and Cbx4CDM constructs (Fig. 3, C, D, and G). Furthermore, transcripts of Flg were also increased upon Cbx4ΔSIM overexpression compared with the Cbx4WT and Cbx4CDM constructs (Fig. 3 H). Thus, similar to human epidermal progenitor cells (Luis et al., 2011), expression of terminal differentiation markers in mouse KCs occurs predominantly under the control of non-Polycomb SUMO E3 ligase activity of Cbx4.

To further elucidate which of the Cbx4 activities were likely responsible for repression of the nonepidermal lineage genes, we analyzed Nefl expression in the response to overexpression of the Cbx4 constructs harboring either chromodomain- or SUMO E3 ligase–mutated domains. A marked increase of both Nefl transcript and protein expression was detected in the Cbx4WT-expressing cells compared with Cbx4WT and Cbx4ΔSIM (Fig. 3, E, F, and I). Thus, Cbx4 represses neuronal gene Nefl in epidermal KCs predominantly via the Polycomb-mediated inhibitory pathway.
Cbx4 is a direct p63 target mediating its inhibitory activity on nonepidermal genes in KCs

p63 is a major transcriptional regulator of epidermal development with multiple functions, including a direct positive transcriptional regulation of epidermal genes such as keratins and adhesion molecules (Koster and Roop, 2007; Vanbokhoven et al., 2011; Botchkarev and Flores, 2014; Kouwenhoven et al., 2015). A recent study has also highlighted an important role for p63 in suppression of nonepidermal lineage genes (De Rosa et al., 2009). To test whether p63-dependent suppression of the nonepidermal genes is mediated by Cbx4, we compared the transcriptomes of epidermal cells isolated by LCM from E18.5 Cbx4 KO and p63 KO embryos. A significant portion (34.7%) of the genes (716 out of 2,059) up-regulated in the p63-deficient KCs were also up-regulated in Cbx4 KO basal epidermal cells, suggesting a functional link between p63 and Cbx4 (Fig. 4 A and Table S2). Furthermore, ChIP-seq analyses showed that 72 of these genes, including the Nefl gene, served as direct Cbx4 targets in KCs (Fig. 4 A and Table S2). The expression changes of the Nefl gene were further validated by qRT-PCR in p63-deficient KCs isolated from p63 KO embryos and p63 siRNA–treated primary mouse KCs (Fig. 4, B and C). Similar to Cbx4 KO epidermal cells, p63-deficient skin epithelial cells showed strongly up-regulated Nefl protein expression compared with WT cells (Fig. 4, D and E), further substantiating functional cooperation between p63 and Cbx4 in the repression of nonepidermal genes in the developing epidermal KCs.

To further investigate the relationships between p63 and Cbx4, we analyzed Cbx4 expression in p63 KO embryonic skin. Both the Cbx4 transcript and protein levels were markedly reduced in the p63-deficient KCs (Fig. 4, F–H). Interestingly, in p63 KO dermal cells, Cbx4 protein expression was detectable at similar levels to control mice (Fig. 4 G), suggesting a KC-specific Cbx4 regulation by p63 transcription factor. By using a PatSearch tool (Grillo et al., 2003), we identified a highly conserved region ~5.5 kb upstream of the Cbx4 transcription start site containing several putative p63-binding sites (Fig. 4 I). Furthermore, ChIP revealed binding of the p63 transcription factor to the identified region in primary mouse KCs (Fig. 4 J). We cloned a 2.4-kb Cbx4 5′ UTR fragment, containing the putative p63-binding sites as an enhancer, into the pG13 promoter plasmid (pG13-Cbx4-luc) and coexpressed it with either ΔNp63-expressing or empty vector in the immortal KC HaCaT cell line. A marked increase in luciferase activity was seen in cells coexpressing pG13-Cbx4-luc with pΔNp63 vectors compared with a control plasmid (pFLAG-CMV2; Fig. 4 K), thus showing a direct transcriptional regulation of Cbx4 by p63 in KCs.

Ectopic Cbx4 expression partially rescues the epidermal phenotype in embryonic skin explants of p63-deficient mice

To assess whether the restoration of Cbx4 expression is capable of partially rescuing the skin phenotype of p63 KO mice, E13.5 skin explants isolated from p63 heterozygous (p63+/−) mice were cultured ex vivo and infected with p63 shRNA–expressing lentiviruses in combination with either Cbx4–expressing or control lentiviruses (Fig. 5, A–E). Compared with the p63+/− skin explants treated with control viruses, treatment with p63 shRNA in combination with control lentiviruses resulted in a marked decrease of the epidermal thickness, cell proliferation, and expression of epidermal keratins K14 and K10, thus reproducing in part the skin phenotype of p63 KO mice (Fig. 5, A–E). However, cotreatment of p63+/− skin samples with p63 shRNA– and Cbx4–expressing lentiviruses resulted in a significant increase of epidermal thickness, cell proliferation, and K14/K10 expression compared with the samples treated with p63 shRNA and control lentiviruses (Fig. 5, A–E). The restoration of epidermal proliferation after ectopic Cbx4 expression was consistent with data showing reduced cell proliferation in the epidermis of Cbx4KO mice compared with WT controls (Fig. 1, G and H). Furthermore, the Nefl protein was strongly decreased in the epithelium of p63-deficient embryonic skin explants transduced with Cbx4 (Fig. 5, D and E), suggesting that Cbx4 is indeed capable of inhibiting its expression in epidermal KCs ex vivo. Therefore, Cbx4 plays a role in mediating the p63-dependent program of gene repression during epidermal development and is capable of partially restoring the epidermal phenotype of p63-deficient mice.

Discussion

Polycomb-dependent transcriptional repression is a powerful mechanism that shapes gene expression programmers in essentially all cell types of living organisms during development and postnatal homeostasis, whereas alterations in the patterns of Polycomb-dependent gene repression contribute to many pathological conditions including carcinogenesis (Simon and Kingston, 2013). Polycomb PRC1 and PRC2 are multiprotein complexes, and despite the fact that the role of some Polycomb genes, such as Ezh1/2, in execution of terminal differentiation programmers in the epidermis and hair follicles was shown previously (Ezhkova et al., 2009, 2011), the effects of the other PRC1/2 components on the establishment of lineage-specific patterns of gene repression during skin development and epidermal differentiation remain unclear. Here, we show that PRC1 component Cbx4 plays a unique role in the establishment and maintenance of the KC lineage identity and represses nonepidermal lineage (neuronal) genes in the epidermal progenitor cells, as well as controls proliferation and inhibits premature differentiation in basal epidermal KCs.

We demonstrate that among different PRC1 genes, Cbx4 shows the most prominent changes in its expression in the embryonic epidermis during its stratification, where Cbx4 is seen in basal and suprabasal KCs. Genetic Cbx4 ablation results in a marked decrease of epidermal thickness and proliferation, as well as in the premature appearance of the terminal differentiation markers in the immediate suprabasal epidermal cells. Interestingly, Ezh2KO mice also show a decrease of epidermal proliferation and premature terminal differentiation (Ezhkova et al., 2009). Similar to Ezh2KO mice (Ezhkova et al., 2009), Cbx4-null mice show an increase of the p16/p19 and p57 cell cycle inhibitor genes in the epidermis, which, at least in part, explains the stimulatory effects of Cbx4 on epidermal proliferation. However, in contrast to Ezh2KO mice, Cbx4 genetic ablation results in a marked decrease of the epidermal thickness, suggesting that Cbx4 plays a unique yet partially overlapping role with Ezh2 in the control of epidermal development.

The decreased cell proliferation observed in the epidermis upon Cbx4 ablation is consistent with our previous results that show reduced cell proliferation and thymus hypoplasia in Cbx4KO mice (Liu et al., 2013). In contrast to other Cbx proteins, Cbx4 uniquely possesses both Polycomb and non-Polycomb
SUMO E3 ligase activities (Li et al., 2007; Luis et al., 2011). We show here that in cultured primary epidermal KCs, only the non-Polycomb E3 ligase–deficient Cbx4 (Cbx4ΔSIM) is capable of inhibiting proliferation, suggesting that the SUMO E3 ligase activity of Cbx4 contributes to the alterations of epidermal proliferation in Cbx4 KO mice. Interestingly, KCs treated with Cbx4ΔSIM also show a marked increase in the expression of terminal differentiation–associated markers, such as Involucrin or Filaggrin, thus suggesting that non-Polycomb SUMO E3 ligase Cbx4 activity might also inhibit the premature activation of terminal differentiation genes in Cbx4ΔSIM mice.

Figure 4. Cbx4 serves as a direct p63 target in epidermal KCs. (A) Overlap of the Cbx4 target genes commonly up-regulated in laser-captured epidermis of E16.5 Cbx4KO and p63KO mice. [B and C] qRT-PCR validation of Nefl gene up-regulation in the epidermal progenitor cells in p63KO mice [B] and primary epidermal KCs expressing p63 shRNA [C; mean ± SD]. n = 3. (D) Increased Nefl protein expression in the epidermis of p63 mutant mice (arrowheads). Nefl-expressing dermal nerve fibers are shown by arrows. Dashed lines separate epidermis and dermis. (E) The number of Nefl+ epidermal cells is significantly increased in p63KO mice (mean ± SD). n = 3. (F and G) Cbx4 transcript [F] and protein [G] are markedly down-regulated in the epidermis of p63KO mice (mean ± SD). n = 3. Dashed lines separate epidermis and dermis. (H) A significant reduction in the number of Cbx4+ cells in p63KO skin compared with WT skin (mean ± SD). n = 3. (I) Conserved putative p63-binding sites are identified by the PatSearch tool in the mouse and human Cbx4 5′ UTR. Asterisks indicate consensus sequences for p63-binding sites, and core sequence elements are indicated by red asterisks. (J) ChIP-qPCR showing p63 binding to the identified region in the mouse Cbx4 5′ UTR. (K) A 2.4-kb region in the Cbx4 5′ UTR increases minimal SV40-promoter activity (pGL3-Cbx4-luc) upon coexpression with a ΔNp63-expressing construct compared with an empty plasmid (pFLAG-CMV2). Bars, 25 µm. **, P < 0.01; ***, P < 0.001.

Our data also reveal that the Cbx4 chromodomain is involved in the repression of the senescence-associated Cdkn2a gene, whereas Cbx4 ablation results in the accumulation of H2AX as one of the senescence markers (Turinetto and Gia-chino, 2015) in epidermal KCs. These data are in full concordance with the data of Luis et al. (2011) that show similar effects of the Cbx4 chromodomain on the expression of senescence-associated genes in human epidermal KCs.
the levels of H3K27me3 were not altered in the epidermis of Cbx4KO mice, Cbx4 ablation resulted in a marked decrease in H2AK119Ub in the epidermis (presumably to its basal levels) compared with WT controls. The fact that Cbx4-null mice showed reduced levels of H2AK119Ub only in the epidermis, whereas H2AK119Ub levels in the dermal cell were not altered upon Cbx4 ablation compared with WT controls, points to the specificity of this interaction.

Interestingly, Cbx4 ablation is also accompanied by an increase of epidermal expression of Kdm2b histone demethylase that forms a noncanonical PRC1 complex together with PCGF1 and RYBP and promotes basal ubiquitylation of H2AK119 at unmethylated CpG-rich DNA regions (Blackledge et al., 2014; Cooper et al., 2014; Kalb et al., 2014). These data suggest that interaction of the noncanonical PRC1 complex with gene targets is likely not altered in the epidermis of Cbx4KO mice and that the Cbx4 chromodomain is specifically involved in the execution of the canonical PRC1-mediated gene repression in the epidermis via interaction with H3K27me3 followed by H2AK119 ubiquitylation.

However, a correlation between the Cbx4 and Ezh1/2 ChIP-seq targets in epidermal KCs might help in addressing the roles for PRC1 and PRC2 complexes in repressing the activities of the distinct gene groups in KCs. Our data reveal a contribution of the Cbx4 chromodomain to the repression of non-KC lineage (neuronal) genes in the epidermis, which is quite consistent with the data obtained from the Ezh2KO mice (Ezhkova et al., 2009). By merging the microarray data obtained from the epidermis of Cbx4KO mice with Cbx4 ChIP-seq and H3K27me3 ChIP-seq data, we demonstrate an enrichment of the genes associated with nervous system development among the direct Cbx4 targets in KCs. We show that several key genes encoding structural components of the neurons and Schwann cells (Nefl and Mobp) and transcription factors involved in the control of nervous system development (Neurog3, Olig2, En2, and Lhx4) are up-regulated in the epidermis of Cbx4KO mice compared with WT controls.

Epidermal and neuronal lineages share the common ectodermal origin (Patthey and Gunhaga, 2011; Benitah and Frye, 2012), and mechanisms that control gene repression during specification of these two lineages in the embryo are unclear. Ezh2 is strongly expressed in neural stem cells and is down-regulated during their differentiation into neurons and astrocytes, whereas its expression is increased during neural stem cell differentiation into the oligodendrocyte lineage (Sher et al., 2008). Analyses of the Ezh2 targets in neural stem cells (Sher et al., 2012) reveal that several genes encoding transcription factors involved in the control of nervous system development (En2, Lhx4, Olig2, Pax3, and Pitx2) also represent the direct Cbx4 targets in epidermal KCs. These data support an idea that Cbx4, through its chromodomain, and Ezh2 operate in concert to repress neuronal genes and maintain the epithelial fate in the epidermal progenitor cells, whereas Ezh2 also represses their differentiation into the Merkel cell lineage independently of Cbx4 (Bardot et al., 2013).

Specification and differentiation of the epidermal progenitor cells during development is supported by the p63 transcription factor. Cbx4 partially rescues an epidermal phenotype in p63KO mice. (A and B) Hematoxylin/AP staining [A] and immunodetection of Ki67+ proliferating cells [B] in E13.5 ex vivo embryonic skin explants of heterozygous p63+/-KO mice treated with p63 shRNA- and Cbx4-expressing lentiviruses. Dashed lines separate epidermis and dermis. (C) Cbx4 overexpression in p63-deficient epidermal cells significantly increases epidermal thickness and cell proliferation (mean ± SD). n = 3. (D and E) Quantitative immunofluorescence stainings with anti-K14, anti-K10, and anti-Nefl antibodies show down-regulation of K14/K10 and up-regulation of Nefl (arrows) in the epithelium of p63-deficient skin explants. Cotreatment of the explants with p63 shRNA- and Cbx4-expressing lentiviruses results in up-regulation of K14/K10 expression and down-regulation of Nefl expression in p63-deficient epidermis. Integrin-β4 (Itgb4) expression outlines the basement membrane of the epidermis. DAPI counterstain (blue) shows the nuclei (mean ± SD). n = 3. **, P < 0.01; ***, P < 0.001. (F) Scheme summarizing the p63-/Cbx4-mediated effects on gene repression in the developing epidermis. Bars, 25 µm.
factor that serves as a master regulator of epidermal development (Koster and Roop, 2007; Vanbokhoven et al., 2011; Botchkarev and Flores, 2014; Kouwenhoven et al., 2015). Increased evidence demonstrates that p63 regulates several components of the epigenetic machinery (Brg1, Lsh, and Satb1) to regulate chromatin remodeling and establish the epidermal differentiation program (Fessing et al., 2011; Keyes et al., 2011; Mardaryev et al., 2014).

We show here that p63 also operates as a direct upstream regulator of Cbx4, which mediates its effects on cell proliferation and is involved in the repression of non-KC lineage genes. We showed previously that p63 directly interacts with Cbx4 in thymocytes, suggesting a possibility for both proteins to form a transcription complex to regulate expression of target genes (Liu et al., 2013). Consistently, comparison of the microarray data from the skin epithelium of p63KO mice and epidermis of Cbx4KO mice reveals that ~35% of the genes up-regulated in the skin epithelium of p63KO mice are also up-regulated in the epidermis of Cbx4 mutants, whereas ~10% of these genes represent direct Cbx4 targets in KCs. Furthermore, we show that Cbx4 expression is markedly decreased in the skin epithelium of p63KO mice, p63 shows binding to the Cbx4 regulatory region and stimulates Cbx4 promoter activity, and Cbx4 is capable of partially rescuing alterations of KC proliferation upon p63 ablation in skin organ cultures.

Collectively, these data provide compelling evidence that Cbx4 operates as an important part of the p63-regulated program of establishing and maintaining epithelial cell fate in epidermal progenitors and that Cbx4-mediated repression of non-KC lineage genes serves as a crucial component of this program (Fig. 5 F). However, additional analyses are required to fully understand the complexity of Cbx4-dependent gene repression in epidermal KCs. In particular, the role of Cbx4 in the repression of other lineage-specific genes, including mesodermal genes in KCs, and the potential contribution of these genes in the control of epithelial–mesenchymal transition need to be carefully explored. Also, the roles of other members of the Cbx family, including Cbx6 and Cbx7, which might substitute for Cbx4 in the canonical PRC1 complex and partially compensate for the effects of Cbx4 ablation in epidermal KCs, need to be clarified.

Because skin contains a large variety of the progenitor cells for distinct cell lineages (epithelial, neuronal, and mesenchymal; Hunt et al., 2009), future research in this direction will shed light on the role of distinct Cbx family members in the control of progenitor cell differentiation and/or reprogramming and isolated from the skin. This will also help to design novel approaches for modulating gene expression programmes in healthy and diseased skin via targeting the activity of Cbx genes.

Materials and methods

Animals and tissue collection

All animal studies were performed under protocols approved by a Home Office Project License. C57BL/6 mice were purchased from Charles River. Skin samples were collected from mice at distinct days of embryonic and postnatal development (E11.5, E14.5, E16.5, E18.5, P1.5, and P56). p63KO embryos were obtained by breeding p63<sup>−/−</sup> animals purchased from The Jackson Laboratory. Generation of Cbx4KO mice was described previously (Liu et al., 2013). In brief, the N-terminal region of the Cbx4 gene including the first two exons and a 0.9-kb upstream region was targeted for Cbx4 gene disruption. The null allele was gained upon Cre-loxP excision by crossing mice carrying the floxed allele with Ella-Cre transgenic mice to achieve global gene KO, and the mice were bred on the C57BL/6J-129Sv genetic background. Genotyping of mice was performed using PCR, as described previously (Liu et al., 2013). For each developmental stage, six to seven samples were collected. Tissue samples were snap frozen in liquid nitrogen, embedded into optimal cutting temperature medium (VWR), and processed for immunofluorescent or LCM and microarray analyses as described in the following paragraph.

LCM, microarray, and qRT-PCR analysis

LCM of mouse basal KCs at selected stages of development was performed followed by RNA isolation, amplification, and microarray analysis as previously described (Sharov et al., 2006, 2009). In brief, 8-µm-thick frozen skin cryosections were dehydrated to preserve RNA integrity, and LCM was performed from the epidermis of WT, Cbx4KO, and p63KO mice at E11.5, E16.5, P0.5, and P56 using an LCM system (Arcturus; Life Technologies). Total RNAs were isolated using an RNA isolation kit (PicoPure; Life Technologies) followed by two rounds of linear RNA amplification using an RNA amplification kit (Ribopure; Life Technologies). Equal amounts of RNA from each sample were labeled by Cy3 using a fluorescent labeling kit (Agilent Technologies). Quality and size distribution of the targets was determined by an RNA 6000 Nano Lab-on-a-chip assay (Agilent Technologies), and quantification was determined using a microscale spectrophotometer (NanoDrop). After one round of linear amplification in all analyses, Universal Mouse Reference RNA (Agilent Technologies) was used as a control. All microarray analyses were performed by MGenex, LC, using the 41K Whole Mouse Genome 60-mer Oligo Microarray kit (Agilent Technologies). All microarray data were normalized to the corresponding data obtained from the reference RNA. Two independent datasets were obtained from WT and transgenic mice, and p-values were calculated by Feature Extraction software (version 7.5; Agilent Technologies) using distribution of the background intensity values to signal intensity and using a Student’s t test.

For qRT-PCR analysis of RNA isolated and amplified after LCM, PCR primers were designed on Beacon Designer software (PRE MIER Biosoft International; Table S3). RT-PCR was performed using iQ SYBR green Supermix and the MyiQ Single-Color Real-Time PCR Detection System (Bio-Rad Laboratories). Differences between samples and controls were calculated using the Gene Expression Macro program (Bio-Rad Laboratories) based on the ΔΔct equitation method with glyceraldehyde 3-phosphate dehydrogenase and peptidylprolyl isomerase A genes as internal housekeeping controls.

Plasmids

Cbx4WT and Cbx4-mutated retroviral plasmids were previously validated (Luis et al., 2011). In brief, Cbx4 cDNA was cloned into pMSCV and pMSCV-Puro-IRES-GFP. The chromodomain was mutated using the QuickChange II XL Site-Directed Mutagenesis kit (Agilent Technologies) to create the F11A and W35L double mutants. In Cbx4ΔSIM plasmid (a gift from D. Wotton, University of Virginia, Charlottesville, VA), Cbx4 cDNA was mutated to harbor internal deletion of nucleotides at the 784–795 and 1,381–1,404 positions, corresponding to both SIM1 (IVIV) and SIM2 (EVIIVLDSD) amino acid sequences, respectively (Merrill et al., 2010). Cbx4 knockout was done using a validated shRNA construct (5′-CGCGCGACACCACTGAACC TGGTGATTCTCAGATACCAAAGGTTACTGTTGTCTTTTG-3′) cloned into pLKO.1-GFP lentiviral plasmid (Morey et al., 2012). pGIPZ–p63 shRNA lentiviral plasmid (V3LMv_508694; GE Healthcare) contains a 5′-TGATCTCTACGAACTATC-3′ sequence and was used to target p63, pHAGE2-EF1-mCbx4 lentiviral plasmid was produced by cloning PCR-amplified mouse Cbx4 cDNA into pHAGE2-EF1 vector (a gift from G. Mostoslavsky, Boston University, Boston, MA).
Production of the Cbx4 and p63 expression and knockdown viruses

Cbx4WT, Cbx4CDM, and Cbx4ΔSIM-expressing retroviruses were produced by transfection of Phoenix-E/HEK293T packaging cells. Cbx4 shRNA– and p63 shRNA–expressing lentiviruses were produced by cotransfection of HEK293T cells with pLKO.1-EGFP-Cbx4 shRNA or pGIPZ–p63 shRNA and packaging (pPAX2) and envelope (pMD2.G) plasmids. For Cbx4-expressing lentiviruses, HEK293T cells were cotransfected with pHAGE2-EIF1-mCbx4 and helper plasmids (pTAT, pREV, pHagg2(GAG/Pol), and pVSV-G) as published elsewhere (Mostoslavsky et al., 2005). For Cbx4-mutated retroviruses, Phoenix-E cells were transfected with pMSCV-PIG-CB4X-WT, pMSCV-PIG-CB4X-CDM, and pMSCV-PIG-CB4X-ΔSIM plasmids as published previously (Luis et al., 2011). Cell culture medium containing viruses was collected 24 h, 48 h, and 72 h after transfection, followed by precipitation of the viral particles using PEG-it solution (System Biosciences) as per the manufacturer’s protocol.

Primary mouse KC culture

Epidermis from newborn C57BL/6 was separated from dermis by overnight digestion with 0.25% trypsin at 4°C. Epidermal sheath was added to prechilled, supplemented, low-calcium KC culture medium (MEM, 4% FBS, 0.05 mM CaCl₂, 0.4 μg/ml hydrocortisone, 5 μg/ml insulin, 10 mg/ml EGF, 10−10 M cholera toxin, 2 × 10−4 M T3, 2 mM L-glutamine, 100 U/ml penicillin, and 100 μg/ml streptomycin). To make a single-cell suspension, the epidermal tissue was chopped with scissors and triturated, followed by filtering through a 70-μM silicon strainer and plating onto the surface of 6-well plates containing Williams’ medium E. Within 2 h after the culture was set up, cells were transfected with the retroviral/lentiviral particles. Cells were collected for subsequence RT-PCR or immunofluorescence analyses 48 h after infection.

FACS

A single-cell suspension of newborn epidermal KCs was prepared as described in the previous paragraph. To ensure analysis and sorting of viable cells with intact chromatin, KCs were stained with UV Live/Dead Fixable Dye (Life Technologies) for 30 min on ice before fixation with 1% PFA for 10 min at RT. Fixed cells were labeled with CD49f-phycoerythrin and Sca-1–FITC antibodies (Table S4) for 1 h on ice. CD49f+/Sca-1+ basal KCs were gated after exclusion of dead cells and sorted on a cell sorter (MoFlo XDP; Beckman Coulter) as described elsewhere (Jensen et al., 2010). Sorted cells were pelleted at 2,000 g and stored at −80°C.

Luciferase reporter assay

To make the pG3-Cbx4-luc plasmid, a 2.4-kb Cbx4 enhancer region (−5,954/−3,523 bp upstream of the Cbx4 transcription start site) was PCR amplified from genomic DNA and cloned into the Luciferase reporter plasmid pG3 promoter (Promega) at BamHI–SalI sites using the In-Fusion HD Cloning System (Takara Bio Inc.). For the reporter assay, HaCaT cells were seeded into white-bottomed/white-walled 96-well plates at 10,000 cells/well, and transfections were performed after overnight incubation at 37°C. Cells were transfected with 200 ng DNA total plus 10 ng pRLTK Renilla Control vector (Promega): 100 ng pG3-Cbx4-luc and 100 ng pΔNp63 or empty pFLAG-CMV2 vectors. After 48 h of incubation at 37°C, cells were washed with PBS, and the assay was performed using the Dual-Glo Luciferase Assay System (Promega) according to the manufacturer’s protocol. Firefly luciferase activity was normalized against the Renilla luciferase activity, and the data represent three independent triplicates.

Histology and immunohistochemistry

For histology and histomorphometry, 8-μm cryosections were fixed in 4% PFA for 10 min at RT followed by hematoxylin staining and AP activity visualization using histoenzymatic reaction (Botchkareva et al., 1999, 2000; Ahmed et al., 2011, 2014). Epidermal thickness was measured in 10 microscopic fields using ImageJ software (National Institutes of Health). For immunohistochemistry, 8-μm cryosections were fixed in 4% PFA for 10 min at RT and incubated with primary antibodies (Table S4) overnight at 4°C, followed by application of corresponding Alexa Fluor 488 and Cy-555–coupled secondary antibodies (1:200; Life Technologies) for 60 min at 37°C. Cell nuclei were visualized with 4′,6-diamidino-2-phenylindol. Images were acquired on a microscope (Eclipse 50i; Nikon) with 20×/0.50 NA and 40×/0.75 NA Plan Fluor objectives (Nikon) using a digital camera (ExiAqua; QImaging) and Image-Pro Express 6.3 image analysis software (QImaging).

Comparative analysis of Ki67+, Ivl+, Nefl-, and Cbx4+ cells was done by evaluating the ratio of the number of positive cells to DAPI+ cells, as described previously (Sharov et al., 2003, 2005; Mardaryev et al., 2011). For quantification of Ki67+ and Cbx4+ cells in tissue cryosections, 10 microscopic fields (20×) from three WT and Cbx4KO skin sections were included in the analysis. For quantification of Ki67+, Ivl+, and Nefl- cells cultured in vitro, five microscopic fields (20×) taken from the center and each of the four sides of the well were used in the analysis. Data were pooled, the means ± SD were calculated, and statistical analysis was performed using a two-tailed t test (α = 0.05).

Immunofluorescence intensity was determined using ImageJ software as described previously (McClay et al., 2014). In brief, red or green fluorescent signals were collected from experimental tissues in RGB format using the same exposure conditions. To measure the fluorescence intensity at each pixel, the RGB images were converted to 8-bit grayscale format. Regions of interest of distinct size were selected within WT or Cbx4KO epidermis and dermis, and the corrected values of total cell fluorescence were calculated for each selected area using the following formula: corrected total cell fluorescence = integrated density − (area of selected cell × mean fluorescence of background readings). For pairwise comparisons, a two-tailed t test (α = 0.05) was used. Where multiple samples were compared, a one-way analysis of variance (ANOVA) was used, followed by the Newman–Keuls test (α = 0.05).

ChIP-seq and ChIP-qPCR assays

ChIP was performed using epidermal KCs isolated from newborn mouse skin and anti-H3K27me3, anti-Cbx4, and anti-H2AK119ub1 antibodies (Table S5). In brief, a single-cell suspension of mouse epidermal KCs was prepared after overnight digestion in 5 mg/ml dispase (Roche) at 4°C, followed by a 5-min treatment with 0.2% trypsin (Life Technologies). Basal epidermal KCs were sorted by FACS and further processed using a ChIP-IT High Sensitivity kit (Active Motive). 5 × 10⁸ PFA-fixed cells were used as a starting material. Indexed ChIP-seq libraries were generated using NEBNext reagents (New England Biolabs, Inc.), and ChIP libraries were sequenced on the HiSeq 2500 platform (Illumina), producing 30–70 million reads per library. Sequencing reads were aligned to the mm9 mouse genome assembly (University of California, Santa Cruz, Genome Browser). Specific areas of binding were identified with Sicer using default settings. For ChIP-qPCR, precipitated DNA was analyzed using primers designed for the promoter regions of Cbx4, Lor, Nefl, and Cdks2a genes (Table S3). ChIP-qPCR data were pooled, means ± SD were calculated, and statistical analysis was performed using a one-way ANOVA test as described previously (Mardaryev et al., 2011).

Embryonic tissue culture

Whole dorsal skin culture from E13.5 p63<sup>c−</sup> embryos were prepared and maintained as previously described (Botchkarev et al., 1999). In brief, skin samples were dissected from embryos and cultured in the 6-well plates containing Williams’ medium E. Within 2 h after the cul-


